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(11)

**EP 1 106 061 A1**

(12)

**EUROPEAN PATENT APPLICATION**  
published in accordance with Art. 158(3) EPC

(43) Date of publication:

13.06.2001 Bulletin 2001/24

(51) Int Cl.7: **A01K 67/027, C12N 5/10,  
C12N 15/06**

(21) Application number: **99938578.4**

(86) International application number:  
**PCT/JP99/04518**

(22) Date of filing: **23.08.1999**

(87) International publication number:  
**WO 00/10383 (02.03.2000 Gazette 2000/09)**

(84) Designated Contracting States:

**AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU  
MC NL PT SE**

Designated Extension States:

**AL LT LV MK RO SI**

(30) Priority: **21.08.1998 JP 23616998**

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**(54) METHOD FOR MODIFYING CHROMOSOMES**

(57) The present invention relates to a method for producing a modified foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

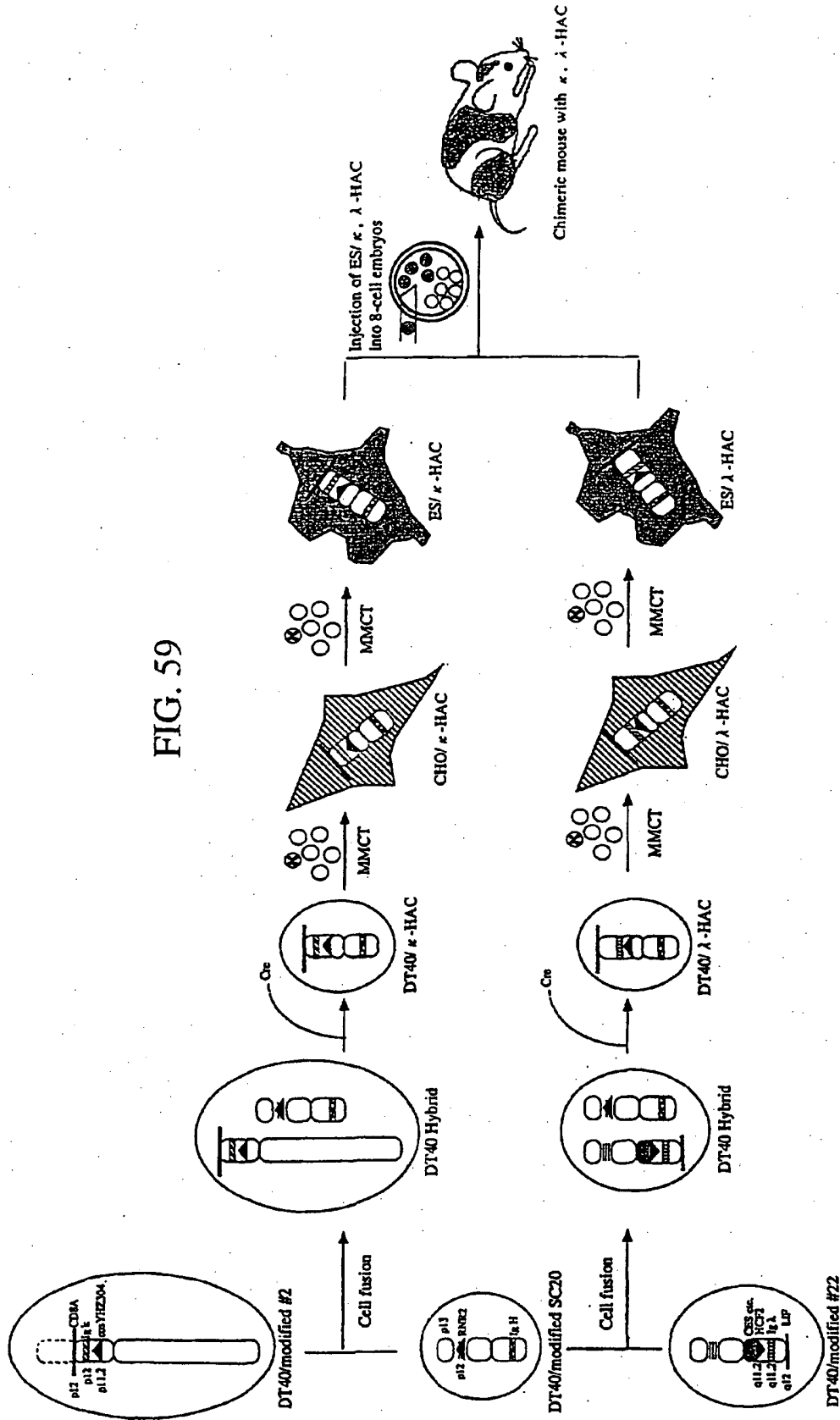
- (a) preparing a microcell comprising a foreign chromosome(s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment(s) into a cell with high homologous recombination efficiency through its fusion with said microcell;
- (b) in said cell with high homologous recombination efficiency, inserting a targeting vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof, and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking said desired site; and
- (c) in said cell with high homologous recombination efficiency, causing deletion and/or translocation to occur at the marked site of said foreign chromo-

some(s) or a fragment(s) thereof.

The invention also relates to a method for producing a chimeric non-human animal or a non-human animal, which comprises a modified foreign chromosome(s) or a fragment(s) thereof; the chimeric non-human or non-human animal; a recombinant chromosome(s) or a fragment(s) thereof, which is obtained by deletion and/or translocation of a chromosome(s) or a fragment(s) thereof; a method for modifying a foreign chromosome(s) or a fragment(s) thereof in a cell; an artificial chromosome vector, which comprises a centromere sequence derived from human chromosome #14 and a recognition sequence for a site-directed recombination enzyme; and a recombinant chromosome(s) or a fragment(s) thereof obtained by deletion and/or translocation of a chromosome(s) or fragment(S) thereof.

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## Description

## Technical Field

**[0001]** The present invention relates to a method for modifying a chromosome(s) or fragment(s) thereof. The present invention also relates to chimeric non-human animals, a method for producing the same and a method for using the same. The present invention allows chimeric non-human animals to retain a foreign giant DNA fragment(s) of at least 1 Mb and to express the gene(s) on such a fragment(s), which was impossible heretofore. Hence, the following becomes possible by using the method.

- Production of animals which retain and express a full length of a gene encoding a biologically active substance, for example, a full length of human antibody gene. The biologically active substance, for example, a human-type antibody is useful as a pharmaceutical product.

- Analysis of functions of human giant genes (e.g., histocompatibility antigen, dystrophin, etc.) in animals.

- Production of model animals with human dominant hereditary disease and a disease due to chromosomal aberration.

**[0002]** The present invention relates to pluripotent cells in which endogenous genes are disrupted, use of the same, and a method for producing chimeric non-human animals and use of the animals. If a foreign chromosome or a fragment thereof containing a gene encoding a gene product identical with or homologous to the gene product encoded by the disrupted endogenous gene is transferred into the pluripotent cell of the present invention as a recipient cell so that a desired functional cell or a desired chimeric non-human animal is produced from the cell, the transferred gene can be expressed efficiently without differentiation of the pluripotent cell into a germ cell. Even if a germ cell of the non-human animal is affected or the pluripotent cell cannot be differentiated into a germ cell by the disruption of the endogenous gene or the introduction of a foreign gene, a functional cell, or a chimeric non-human animal, a tissue or a cell of the animal can retain and express a foreign giant DNA fragment in excess of the heretofore unattainable 1 Mb (a million bases) in conditions of a deficiency in the endogenous gene and a decrease in the production of an endogenous gene product by producing the desired functional cell or non-human animal from the pluripotent cell.

## Background Art

**[0003]** Techniques of expressing foreign genes in animals, that is, techniques of producing transgenic animals are used not only for obtaining information on the gene's functions in living bodies but also for identifying DNA sequences that regulate the expression of the genes (e.g., Magram et al., *Nature*, 315:338, 1985), for developing model animals with human diseases (Yamamura et al., "Manual of model mice with diseases" published by Nakayama Shoten, 1994), for breeding farm animals (e.g., Muller et al., *Experientia*, 47:923, 1991) and for producing useful substances with these animals (e.g., Velandar et al., *P.N.A.S.*, 89:12003, 1992). Mice have been used the most frequently as hosts for gene transfer. Since mice have been studied in detail as experimental animals and the embryo manipulating techniques for mice have been established, they are the most appropriate kind of mammals for gene transfer.

**[0004]** Two methods are known for transferring foreign genes into mice. One is by injecting DNA into a pronucleus of a fertilized egg (Gordon et al., *P.N.A.S.*, 77:7380, 1980). The other is by transferring DNA into a pluripotent embryonic stem cell (hereinafter referred to as "ES cell") to produce a chimeric mouse (Takahashi et al., *Development*, 102:259, 1988). In the latter method, the transferred gene is retained only in ES cell-contributing cells and tissues of chimeric mice whereas it is retained in all cells and tissues of progenies obtained via ES cell-derived germ cells. These techniques have been used to produce a large number of transgenic mice up to now.

**[0005]** However, there had been a limit of the size of DNA capable of being transferred and this restricts the application range of these techniques. The limit depends on the size of DNA which can be cloned. One of the largest DNA fragments which have ever been transferred is a DNA fragment of about 670 kb cloned into a yeast artificial chromosome (YAC) (Jakobovits et al., *Nature*, 362:255, 1993). Recently, introduction of YAC containing an about 1 Mb DNA fragment containing about 80 percent of variable regions and portions of constant regions (C $\mu$ , C $\delta$  and C $\gamma$ ) of a human antibody heavy-chain was reported (Mendes et al., *Nature Genetics*, 15:146, 1997). These experiments were carried out by fusing a YAC-retaining yeast cell with a mouse ES cell. Although it is believed that foreign DNA of up to about 2 Mb can be cloned on YAC (Den Dunnen et al., *Hum. Mol. Genet.*, 1:19, 1992), the recombination between homologous DNA sequences occurs frequently in budding yeast cells and therefore, in some cases, a human DNA fragment containing a large number of repeated sequences is difficult to retain in a complete form. In fact, certain recombinations occur in 20-40% of the clones of YAC libraries containing human genomic DNA (Green et al., *Genomics*, 11:584, 1991).

**[0006]** In another method that was attempted, a metaphase chromosome from a cultured human cell was dissected under observation with a microscope and the fragment (presumably having a length of at least 10 Mb) was injected

into a mouse fertilized egg (Richa et al., Science, 245:175, 1989). In the resulting mice, a human specific DNA sequence (Alu sequence) was detected but the expression of human gene was not confirmed. In addition, the procedure used in this method to prepare chromosomes causes unavoidable fragmentation of DNA into small fragments due to the use of acetic acid and methanol in fixing the chromosome on slide glass and the possibility that the injected DNA exists as an intact sequence is small.

[0007] In any event, no case has been reported to date that demonstrates successful transfer and expression in mice of uninterrupted foreign DNA fragments having a length of at least 1 Mb.

[0008] Useful and interesting human genes which are desirably transferred into mice, such as genes for antibody (Cook et al., Nature Genetics, 7:162, 1994), for T cell receptor (Hood et al., Cold Spring Harbor Symposia on Quantitative Biology, Vol. LVIII, 339, 1993), for histocompatibility antigen (Carrol et al., P.N.A.S., 84:8535, 1987), for dystrophin (Den Dunnen et al., supra) are known to be such that their coding regions have sizes of at least 1 Mb. Since human-type antibodies are important as pharmaceutical products, the production of mice which retain and express full lengths of genes for human immunoglobulin heavy chains (~1.5 Mb, Cook et al., supra), and light chain  $\kappa$  (~3 Mb, Zachau, Gene, 135:167, 1993), and light chain  $\lambda$  (~1.5 Mb, Fripiat et al., Hum. Mol. Genet., 4:983, 1995) is desired but this is impossible to achieve by the state-of-the-art technology (Nikkei Biotech, July, 5, 1993).

[0009] Many of the causative genes for human dominant hereditary disease and chromosomal aberration which causes congenital deformity (Down's syndrome, etc.) have not been cloned and only the information on the approximate location of the genes on chromosome is available. For example, when a gene of interest is found to be located on a specific G band, which is made visible by subjecting a metaphase chromosome to Giemsa staining, the G band has usually a size of at least several Mb to 10 Mb. In order to transfer these abnormal phenotypes into mice, it is necessary to transfer chromosomal fragments of at least several Mb that surround the causative genes, but this is also impossible with the presently available techniques.

[0010] Hence, it is desired to develop a technique by which a foreign DNA longer than the heretofore critical 1 Mb can be transferred into a mouse and expressed in it.

[0011] DNA longer than 1 Mb can be transferred into cultured animal cells by the techniques available today. Such transfer is carried out predominantly by using a chromosome as a mediator. In the case of human, chromosomes have sizes of about 50-300 Mb. Some methods for chromosome transfer into cells have been reported (e.g., McBride et al., P.N.A.S., 70:1258, 1973). Among them, microcell fusion (Koi et al., Jpn. J. Cancer Res., 80:413, 1989) is the best method for selective transfer of a desired chromosome. The microcell is a structural body in which one to several chromosomes are encapsulated with a nuclear membrane and a plasma membrane. A few chromosomes (in many cases, one chromosome) can be transferred by inducing a microcell with an agent that inhibits the formation of spindle in a specific kind of cell, separating the microcell and fusing it with a recipient cell. The resulting libraries of monochromosomal hybrid cells containing only one human chromosome have been used for mapping known genes and specifying the chromosomes on which unknown tumor-suppressor genes and cellular senescence genes exist (e.g., Saxon et al., EMBO J., 5:3461, 1986). In addition, it is possible to fragment a chromosome by irradiating a microcell with  $\gamma$ -rays and to transfer part of the fragments (Koi et al., Science, 260:361, 1993). As described above, microcell fusion is considered to be an appropriate method for transferring DNA larger than 1 Mb into a cultured animal cell.

[0012] The expectation that a mouse could be generated from a cultured cell turned to a real fact when the ES cell which has stable pluripotency was discovered (Evans et al., Nature, 292:154, 1981). Foreign genes, various mutations and mutations by targeted gene recombination could be introduced into the ES cell, making it possible to perform a wide variety of genetic modifications in mice (e.g., Mansour et al., Nature, 336:348, 1988). The ES cell can be used to produce a mouse having a disrupted target gene by gene targeting techniques. The mouse is mated with a transgenic mouse having a gene of interest to produce a mouse that expresses the gene of interest efficiently. For example, a mouse having a disrupted endogenous antibody gene can be mated with a mouse having a human antibody gene transferred to produce a mouse that expresses the human antibody efficiently. A normal diploid cell has alleles. A transgenic mouse having one allele of an mouse antibody heavy-chain gene disrupted expresses an increased level of human antibody in its serum. A mouse having both alleles of mouse antibody heavy-chain gene disrupted expresses a further remarkably increased level of human antibody (S.D. Wagner et al., Genomics, 35:405-414, 1996).

[0013] Some researchers have developed a technique in which one allele of a target gene is disrupted, and then the concentration of a selective drug is increased, thereby deleting both alleles of the target gene (double knock-out). However, this technique holds the possibility of a decrease in the ability of the target gene-deficient cell to differentiate into a germ cell because the target gene-deficient cell obtained by the high-concentration-selective-culture method is cultured in vivo for a long period and because the drug-selection pressure is severe (Takatsu Taki, Experimental Medicine, supplement, Biomanual UP Series Basic Techniques for Immunological Study, Yodo-sha, 1995). In another case, if two kinds of selective drugs are used for double knocking-out, for example, if a neomycin-resistant cell is subjected to a double knock-out treatment with hygromycin, the double drug-resistant ES cell is rarely differentiated to produce a mutant mouse (Watanabe et al., Tissue Culture 21, 42-45, 1995). ES cells may lose their differentiation and growth capabilities under certain culture conditions. When a gene targeting procedure is performed twice, ES cells do not lose



the ability to differentiate into germ cells of a chimeric mouse but the second homologous recombination frequency is extremely low (Katsuki et al., *Experimental Medicine*, Vol. 11, No. 20, special number, 1993). Hence, when a target gene-deficient homozygote is produced, particularly when at least two target genes are targeted, a mouse deficient in each target gene is produced and then the produced mice are mated with each other to produce a homozygote mouse deficient in at least two genes (N. Longberg et al., *Nature*, 368:856-859, 1994). If genes to be disrupted exist close to each other and if a mouse deficient in at least two genes cannot be obtained by mating, heterozygote mice deficient in the two target genes are produced from ES cells and they are mated to produce homodeficient mice (J.H. van Ree et al., *Hum Mol Genet* 4:1403-1409, 1995).

[0014] An attempt to differentiate a pluripotent ES cell into a functional cell in vitro has been made (T. Nakano et al., *Science*, 265:1098-1101, 1994, A.J. Potocnik et al., *The EMBO Journal*, 13:5274-5283, 1994). The cultivation system used in this attempt, for example, a system in which the differentiation into a mature B cell can be induced is expected to be used in the identification of unknown growth and differentiation factors which will work in development and differentiation processes of B cells.

[0015] As long as the transfer of giant DNA is concerned, it has been believed that the size of the aforementioned foreign DNA fragment which can be cloned into a YAC vector is the upper limit. The prior art technology of chromosome transfer for introducing a longer DNA into cultured cells has never been applied to gene transfer into mice and this has been believed to be difficult to accomplish (Muramatsu et al., "Transgenic Biology", published by Kodansha Scientific, p.143 -, 1989).

[0016] The reasons are as follows.

- The transfer of a human chromosome into a mouse ES cell of a normal karyotype as a recipient cell would be a kind of transfer of chromosomal aberration. Up to now, it has been believed that genetic aberration at chromosomal levels which is large enough to be recognizable with microscopes is generally fatal to the embryogeny in mice (Gropp et al., *J. Exp. Zool.*, 228:253, 1983 and Shinichi Aizawa, "Biotechnology Manual Series 8, Gene Targeting", published by Yodosha, 1995).
- Available human chromosomes are usually derived from finitely proliferative normal fibroblasts or differentiated somatic cells such as cancer cells and the like. It was believed that if a chromosome derived from such a somatic cell was transferred into an undifferentiated ES cell, the transferred chromosome might cause differentiation of the ES cell or its senescence (Muller et al., *Nature*, 311:438, 1984; Sugawara, *Science*, 247:707, 1990).
- Only few studies have been reported as to whether a somatic cell-derived chromosome introduced into an early embryo can function in the process of embryonic development as normally as a germ cell-derived chromosome to ensure the expression of a specific gene in various kinds of tissues and cells. One of the big differences between the two chromosomes is assumed to concern methylation of the chromosomal DNA. The methylation is changed according to differentiation of cells and its important role in the expression of tissue-specific genes has been suggested (Ceder, *Cell*, 53:3, 1988). For example, it has been reported that if a methylated DNA substrate is introduced into a B cell, the methylated DNA is maintained after replication and suppresses a site-directed recombination reaction which is essential to the activation of an antibody gene (Hsieh et al., *EMBO J.*, 11:315, 1992). In addition, it was reported that higher levels of de novo methylation occurred in established cell lines than in vivo (Antequera et al., *Cell*, 62:503, 1990). On the basis of the studies reported, it could not be easily expected that an antibody gene in a human fibroblast or a human-mouse hybrid cell which was likely to be methylated at a high level would be normally expressed in a mouse B cell.

[0017] It should be noted that there are two related reports of Illmensee et al. (*P.N.A.S.*, 75:1914, 1978; *P.N.A.S.*, 76:879, 1979). One report is about the production of chimeric mice from fused cells obtained by fusing a human sarcoma cell with a mouse EC cell and the other is about the production of chimeric mice from fused cells obtained by fusing a rat liver cancer cell with a mouse EC cell. Many questions about the results of the experiments in these two reports were pointed out and thus these reports are considered unreliable (Noguchi et al., "Mouse Teratoma", published by Rikogakusha, Section 5, 1987). Although it has been desired to perform a follow-up as early as possible, as of today when 17 years have passed since the publication of these reports, successful reproduction of these experiments has not been reported. Hence, it is believed that foreign chromosomes cannot be retained and the genes on the chromosomes cannot be expressed in mice by the method described in these reports.

[0018] Under these circumstances, it has been believed to be difficult to transfer a giant DNA such as a chromosomal fragment and express it in an animal such as mouse. Actually, no study has been made about this problem since the Illmensee's reports.

[0019] Therefore, an object of the present invention is to provide chimeric non-human animals which retain foreign chromosomes or fragments thereof and express genes on the chromosomes or fragments, and their progenies, and a method for producing the same.

[0020] It is also an object of the present invention to provide pluripotent cells containing foreign chromosomes or

fragments thereof and a method for producing the pluripotent cells.

[0021] Another object of the present invention is to provide tissues and cells derived from the chimeric non-human animals and their progenies.

[0022] A further object of the present invention is to provide hybridomas prepared by fusing the cells derived from the chimeric non-human animals and their progenies with myeloma cells.

[0023] A still further object of the present invention is to provide a method for producing a biologically active substance that is an expression product of the gene on a foreign chromosome or a fragment thereof by using the chimeric non-human animals or their progenies, or their tissues or cells.

[0024] It is also an object of the present invention to provide pluripotent cells which can be used as recipient cells into which a foreign chromosome(s) or a fragment(s) thereof is transferred in the production of chimeric non-human animals retaining the foreign chromosome (s) or fragment(s) thereof and expressing a gene(s) on the foreign chromosome(s) or fragment(s) thereof.

[0025] A further object of the present invention is to provide a method for using the pluripotent cells.

[0026] A further object of the present invention is to provide a method for modifying a chromosome(s) or fragment (s) thereof.

#### Disclosure of Invention

[0027] As a result of the various studies conducted to achieve the above objects, the inventors succeeded in transferring chromosomes or fragments thereof derived from human normal fibroblast cells into mouse ES cells and obtaining clones which were capable of stable retention of the chromosomes or fragments. Moreover, they produced from these ES clones those chimeric mice which retained human chromosomes in normal tissues and which expressed several human genes including human antibody heavy-chain genes. It has become possible to make that animals retain and express giant DNA fragments by the series of these techniques, although this has been impossible by conventional techniques. Moreover, the inventors succeeded in obtaining embryonic stem cells having both of antibody heavy-chain and light-chain genes knocked out. The inventors have also developed a novel method for artificially modifying chromosomes.

[0028] The subject matter of the present invention is as follows:

1. A method for producing a chimeric non-human animal, which comprises preparing a microcell containing a foreign chromosome(s) or a fragment(s) thereof and transferring the foreign chromosome(s) or fragment(s) into a pluripotent cell by fusion with the microcell.

2. A method for producing a pluripotent cell containing a foreign chromosome(s) or a fragment(s) thereof, which comprises preparing a microcell containing a foreign chromosome(s) or a fragment(s) thereof and transferring the foreign chromosome(s) or fragment(s) thereof into a pluripotent cell by fusion with the microcell.

In the method of item 1 or 2, the foreign chromosome(s) or fragment(s) thereof may be larger than 670 kb, further, at least 1 Mb (one million base pairs). The foreign chromosome or fragment thereof may contain a region encoding an antibody. The microcell containing a foreign chromosome(s) or a fragment(s) thereof may be induced from a hybrid cell prepared by the fusion of a cell from which the foreign chromosome(s) or fragment(s) thereof is (are) derived, with a cell having a high ability to form a microcell. The microcell containing a foreign chromosome (s) or a fragment(s) thereof may be induced from a cell prepared by a further fusion of the microcell induced from the hybrid cell with a cell having a high ability to form a microcell. The cell from which the foreign chromosome(s) or fragment(s) thereof is(are) derived may be a human normal diploid cell. The cell having a high ability to form a microcell may be a mouse A9 cell. The pluripotent cell can be selected from embryonal carcinoma cells, embryonic stem cells, embryonic germ cells and mutants thereof. It is preferred that the foreign chromosome or fragment thereof contains a gene of interest and that the pluripotent cell has a disrupted endogenous gene identical with or homologous to said gene of interest on the foreign chromosome or fragment thereof. It is also preferred that the foreign chromosome or fragment thereof contains at least two genes of interest and that the pluripotent cell has disrupted endogenous genes identical with or homologous to said genes of interest on the foreign chromosome or fragment thereof. In the pluripotent cell, one or both alleles of an endogenous gene identical with or homologous to the gene of interest on the foreign chromosome or fragment thereof may be disrupted. The gene of interest may be an antibody gene. The antibody gene may be one or more sets of antibody heavy-chain and light-chain genes. In the method of item 1 or 2, it is preferred that the foreign chromosome or fragment thereof contains a gene of interest and that the foreign chromosome or fragment thereof is transferred into a pluripotent cell having an endogenous disrupted gene identical with or homologous to the gene of interest and then, a chimera is produced from the pluripotent cell by using an embryo of a non-human animal in a strain deficient in an endogenous gene identical with or homologous to the gene of interest. The non-human animal in a strain deficient in an endogenous gene identical with or homologous to the gene of interest can be produced by homologous recombination in gene

targeting. Preferably, the chimeric non-human animal retains the foreign chromosome(s) or fragment(s) thereof, expresses the gene(s) on the foreign chromosome(s) or fragment(s) thereof, and can transmit the foreign chromosome(s) or fragment(s) thereof to its progeny. The chimeric non-human animal is preferably a mammal, more preferably a mouse.

3. A pluripotent cell containing a foreign chromosome(s) or a fragment(s) thereof.

In the pluripotent cell, the foreign chromosome(s) or fragment(s) thereof may be larger than 670 kb. In the cell of item 3, the foreign chromosome or fragment thereof may contain a gene of interest and the pluripotent cell has an endogenous disrupted gene identical with or homologous to the gene of interest on the foreign chromosome or a fragment thereof. The foreign chromosome or fragment thereof may contain at least two genes of interest and the pluripotent cell has disrupted endogenous genes identical with or homologous to the genes of interest on the foreign chromosome or a fragment thereof. In the pluripotent cell, one or both alleles of an endogenous gene identical with or homologous to the gene of interest may be disrupted. The foreign chromosome or fragment thereof may contain an antibody gene. The antibody gene may be one or more sets of antibody heavy-chain and light-chain genes. The pluripotent cell can be selected from embryonal carcinoma cells, embryonic stem cells, embryonic germ cells and mutants thereof. The cells of item 3 can be used to produce chimeric non-human animals.

4. A chimeric non-human animal retaining a foreign chromosome (s) or a fragment (s) thereof and expressing a gene(s) on the foreign chromosome(s) or fragment(s) thereof, or its progeny retaining the foreign chromosome(s) or fragment(s) thereof and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof.

In the chimeric non-human animal or its progeny, the foreign chromosome(s) or fragment(s) thereof may be larger than 670 kb. The foreign chromosome or fragment thereof may contain a gene of interest and the animal may have an endogenous disrupted gene identical with or homologous to the gene of interest. The foreign chromosome or fragment thereof may contain at least two genes of interest and the animal may have disrupted endogenous genes identical with or homologous to said genes of interest. In the chimeric non-human animal or its progeny, one or both alleles of an endogenous gene identical with or homologous to the gene of interest may be disrupted. The gene of interest may be an antibody gene. The antibody gene may be one or more sets of antibody heavy-chain and light-chain genes.

5. A non-human animal which can be produced by mating the chimeric non-human animals or their progenies of item 4, said non-human animal retaining the foreign chromosome (s) or fragment (s) thereof and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof, or its progeny retaining the foreign chromosome(s) or fragment(s) thereof and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof.

6. A non-human animal retaining the foreign chromosome(s) or fragment(s) thereof and expressing a gene(s) on the foreign chromosome(s) or fragment(s) thereof, which can be produced by mating the chimeric non-human animal or its progeny of item 4, or the non-human animal or its progeny of item 5, with a non-human animal in a strain deficient in said gene(s) or a gene homologous thereto, or its progeny retaining the foreign chromosome(s) or fragment(s) thereof and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof.

7. A tissue from the chimeric non-human animal or its progeny of item 4 or from the non-human animal or its progeny of item 5 or from the non-human animal or its progeny of item 6.

8. A cell from the chimeric non-human animal or its progeny of item 4 or from the non-human animal or its progeny of item 5 or from the non-human animal or its progeny of item 6.

The cell may be a B cell, a primary culture cell derived from an animal tissue or a cell fused with an established cell.

9. A hybridoma prepared by the fusion of the B cell with a myeloma cell.

10. A method for producing a biologically active substance, which comprises expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof in the chimeric non-human animal or its progeny of item 4, the non-human animal or its progeny of item 5 or the non-human animal or its progeny of item 6, or a tissue or a cell thereof, and recovering the biologically active substance as an expression product.

In the method, the cell of the chimeric non-human animal may be a B cell. The B cell may be immortalized by fusion with a myeloma cell. The chimeric non-human animal cell may be fused with a primary culture cell derived from an animal tissue or fused with an established cell line. The biologically active substance may be an antibody. The antibody is preferably an antibody of a mammal, more preferably a human antibody.

11. A biologically active substance which can be produced by the method of item 10.

12. A non-human animal retaining at least one human antibody gene larger than 670 kb and expressing the gene.

The non-human animal of item 12 preferably retains at least one human antibody gene of at least 1 Mb and expresses the gene. The human antibody gene may be a human heavy-chain gene, a human light-chain  $\kappa$  gene, a human light-chain  $\lambda$  gene, or a combination thereof. The non-human animal of item 12 may be deficient in a non-human animal antibody gene identical with or homologous to the human antibody gene. The deficiency of non-human animal antibody gene may be caused by disrupting the non-human animal antibody gene by homologous recombination.

13. A hybridoma prepared by the fusion of a spleen cell of the non-human animal of item 12 with a myeloma cell.  
 14. An antibody produced by the hybridoma of item 13.  
 15. A non-human animal expressing at least one class or subclass of human antibody.

The non-human animal of item 15 may be deficient in an endogenous antibody gene identical with or homologous to the expressed human antibody gene. The class or subclass of human antibody may be IgM, IgG, IgE, IgA, IgD or a subclass, or a combination thereof.

16. A non-human animal retaining a foreign DNA(s) larger than 670 kb and expressing a gene(s) on the foreign DNA(s).

The non-human animal of item 16 may be deficient in an endogenous gene identical with or homologous to the expressed gene on the foreign DNA. The non-human animal of item 16 may retain a foreign DNA(s) of at least 1Mb and express the gene(s) on the foreign DNA(s). The non-human animal may be deficient in an endogenous gene identical with or homologous to the expressed gene on the foreign DNA.

17. A method for producing a transgenic non-human animal, which comprises preparing a microcell containing a foreign chromosome(s) or a fragment(s) thereof, transferring the foreign chromosome(s) or fragment(s) into a cultured cell derived from a blastocyst by fusion with the microcell and transplanting the nucleus of the cultured cell into an enucleated unfertilized egg.

18. A pluripotent cell in which at least two endogenous genes are disrupted.

In the cell of item 18, each of the endogenous genes may be disrupted in one or both alleles. The disrupted endogenous genes may be antibody genes. The disrupted antibody genes may be antibody heavy-chain and light-chain genes. The pluripotent cell can be selected from embryonal carcinoma cells, embryonic stem cells, embryonic germ cells and mutants thereof.

19. A method of producing the cell of item 18 by at least two homologous recombinations.

[0029] The method of item 19 may comprise the steps of:

disrupting one allele of the endogenous gene in the pluripotent cell by homologous recombination using a drug-resistant marker gene;  
 culturing the pluripotent cell in the presence of the drug to select drug-resistant cells; and  
 screening the selected drug-resistant cells to yield a cell in which both alleles of the endogenous gene have been disrupted.

[0030] In the method of item 19, one allele of the endogenous gene in the pluripotent cell may be disrupted by homologous recombination using a drug-resistant marker gene and the other allele of the endogenous gene may be disrupted by another homologous recombination using a drug-resistant marker gene. The same drug-resistant marker gene may be used in the two homologous recombinations. Alternatively, different drug-resistant marker genes may be used in the two homologous recombinations.

[0031] Furthermore, the present invention provides a method of using the pluripotent cell as a recipient cell into which a foreign gene(s) or a fragment(s) thereof, or a foreign chromosome(s) or a fragment(s) thereof are to be transferred. The foreign gene(s) or fragment(s) thereof may be incorporated in a vector such as a plasmid, a cosmid, YAC or the like. Alternatively, the foreign chromosome(s) or fragment(s) thereof may be contained in a microcell. The foreign chromosome(s) or fragment(s) thereof is preferably, but not limited to, one that contains a gene(s) identical with or homologous to the endogenous gene(s) disrupted in the pluripotent cell. The term "homologous gene" means herein a gene encoding the same kind of protein or a protein having a similar property in the same or different species of a given organism.

[0032] Moreover, the present invention provides a method of using the pluripotent cell for producing a chimeric non-human animal.

[0033] The present invention also provides a method of producing a pluripotent cell containing a foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

preparing a microcell containing the foreign chromosome(s) or fragment(s) thereof; and  
 fusing the microcell with said pluripotent cell having at least two endogenous genes disrupted, whereby said foreign chromosome(s) or fragment(s) thereof is transferred into said pluripotent cell.

[0034] The present invention further provides a method of producing a chimeric non-human animal, which comprises the steps of:

preparing a microcell containing a foreign chromosome(s) or a fragment(s) thereof; and  
 fusing the microcell with said pluripotent cell having at least two endogenous genes disrupted, whereby said foreign

chromosome(s) or fragment(s) thereof is transferred into said pluripotent cell.

**[0035]** In the aforementioned two methods, the foreign chromosome(s) or fragment(s) thereof may have a length(s) of at least 1 Mb (100 million base pairs). The foreign chromosome(s) or a fragment(s) thereof may contain a region encoding an antibody. The microcell containing the foreign chromosome(s) or fragment(s) thereof may be induced from a hybrid cell prepared by the fusion of a cell containing the foreign chromosome(s) or fragment(s) thereof, with a cell having a high ability to form a microcell. The microcell containing the foreign chromosome(s) or fragment(s) thereof may be induced from a cell prepared by a further fusion of the microcell induced from the hybrid cell, with a cell having a high ability to form a microcell. The cell containing the foreign chromosome(s) or fragment(s) thereof may be a human normal diploid cell. The cell having a high ability to form a microcell may be a mouse A9 cell. In the methods of producing a chimeric non-human animal, a foreign chromosome(s) or a fragment(s) thereof containing gene(s) identical with or homologous to the endogenous gene(s) disrupted in the pluripotent cell may be transferred into the pluripotent cell having the disrupted at least two endogenous genes and then, a chimera of the cell with an embryo of a non-human animal in a strain deficient in a gene(s) identical with or homologous to said endogenous gene(s) may be prepared. The chimeric non-human animal deficient in a gene identical with or homologous to the endogenous gene disrupted in said pluripotent cell may be produced by homologous recombination in gene targeting. The chimeric non-human animal may be such that it retains the foreign chromosome(s) or fragment(s) thereof, expresses a gene(s) on the foreign chromosome(s) or fragment(s) thereof, and can transmit the foreign chromosome(s) or fragment(s) thereof to its progeny. The chimeric non-human animal may be a mammal, preferably a mouse.

**[0036]** The present invention also provides a pluripotent cell containing a foreign chromosome(s) or a fragment(s) thereof, which is obtainable by a method of producing a chimeric non-human animal, which method comprises the steps of:

preparing a microcell containing the foreign chromosome(s) or fragment(s) thereof; and

fusing the microcell with said pluripotent cell having at least two endogenous genes disrupted, whereby said foreign chromosome(s) or fragment(s) thereof is transferred into said pluripotent cell. The present invention further provides a method of using the cell for producing a chimeric non-human animal.

**[0037]** The present invention also provides a chimeric non-human animal retaining a foreign chromosome(s) or a fragment(s) thereof and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof, which is obtainable by one of the aforementioned methods of producing a chimeric non-human animal, or its progeny. The present invention also provides a non-human animal retaining a foreign chromosome(s) or a fragment(s) thereof and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof which is obtainable by mating between the chimeric non-human animals or its progenies, or its progeny. The present invention further provides a tissue from the aforementioned chimeric non-human animal or its progeny, or the aforementioned non-human animal or its progeny. The present invention still more provides a cell from the aforementioned chimeric non-human animal or its progeny, or the aforementioned non-human animal or its progeny. The cell may be a B cell.

**[0038]** The present invention also provides a hybridoma prepared by the fusion of the cell from the aforementioned chimeric non-human animal or its progeny, or the aforementioned non-human animal or its progeny with a myeloma cell.

**[0039]** The present invention provides a non-human animal or its progeny retaining a foreign chromosome(s) or a fragment(s) thereof and expressing a gene(s) on the foreign chromosome(s) or fragment(s) thereof, which is obtainable by mating said chimeric non-human animal or its progeny or said non-human animal or its progeny retaining the foreign chromosome(s) or fragment(s) thereof and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof, with a non-human animal in a strain deficient in a gene(s) identical with or homologous to said gene(s).

**[0040]** Furthermore, the present invention provides a method of producing a biologically active substance, which comprises expressing a gene(s) on a foreign chromosome(s) or a fragment in the chimeric non-human animal or its progeny, or the non-human animal or its progeny, or a tissue or a cell thereof and recovering the biologically active substance as the expression product. The cell of the chimeric non-human animal or its progeny, or the non-human animal or its progeny may be a B cell. The B cell may be immortalized by fusion with a myeloma cell. The biologically active substance may be an antibody. The antibody may be an antibody of mammal, preferably a human antibody.

**[0041]** Moreover, the present invention provides a method of producing a biologically active substance, which comprises expressing a gene(s) on a foreign chromosome(s) or a fragment in a offspring or a tissue and a cell thereof, wherein the offspring is produced by mating the chimeric non-human animal or its progeny, or the non-human animal or its progeny retaining the foreign chromosome(s) or fragment(s) thereof with a non-human animal in a strain deficient in a gene identical with or homologous to said genes, and expressing the gene(s) on the foreign chromosome(s) or fragment(s) thereof, and recovering the biologically active substance as the expression product.

**[0042]** The present invention also provides a vector comprising a foreign chromosome(s) for use in gene transfer into a non-human animal and a non-human animal cell. The foreign chromosome(s) is preferably one from human,

more preferably a human chromosome #14 fragment. The non-human animal is preferably a mouse.

[0043] The term "allele" is used herein.

[0044] The term "homologous gene" means herein a gene encoding the same kind of protein or a protein having a similar property in the same or different species of a given organism.

[0045] As used herein, the term "foreign chromosome" means a chromosome which is exogenously introduced into a target cell. When producing a cell comprising a modified foreign chromosome(s) or a fragment(s) thereof, the introduced chromosome is a chromosome exogenous to the cell where chromosome modification occurs (for example, a cell with high homologous recombination efficiency, such as a chicken DT-40 cell) Further, when producing a chimeric non-human animal comprising a modified foreign chromosome(s) or fragment(s) thereof, the introduced chromosome is a chromosome exogenous to a pluripotent non-human animal cell (for example, an ES cell) to which a modified chromosome is transferred, in addition to a cell where chromosome modification occurs. Alternatively, when producing a non-human animal comprising a modified foreign chromosome(s) or fragment(s) thereof, the introduced chromosome is a chromosome exogenous to a cell derived from non human animals (for example, a culture cell derived from an embryo, a blastocyst, a fetus, or an adult, or a fetal fibroblast) to which a modified chromosome(s) is transferred, in addition to a cell where chromosome modification occurs.

[0046] The present invention allows a gene on the chromosome(s) or fragment(s) thereof to be expressed in a chimeric non-human animal or a non-human animal that is produced from a pluripotent cell, into which a foreign chromosome(s) or fragment(s) thereof is transferred. Species from which target cells are derived and species from which foreign chromosomes are derived are not specifically limited. Both species of a target cell and of a foreign chromosome be the same or may be different.

[0047] Further summary of the present invention is as follows.

1. A method for producing a cell comprising a modified foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

- (a) preparing a microcell comprising a foreign chromosome (s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment (s) thereof into a cell with high homologous recombination efficiency through its fusion with said microcell;
- (b) in said cell with high homologous recombination efficiency, inserting a targeting vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking the desired site; and
- (c) in said cell with high homologous recombination efficiency, causing deletion and/or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof.

2. The method of item 1, wherein a plurality of said cells with high homologous recombination efficiency are subjected to whole cell fusion after steps (a) and (b) and are subjected to step (c).

3. The method of item 2, wherein a plurality of said cells with high homologous recombination efficiency each comprises a distinct foreign chromosome(s) or a fragment(s) thereof.

4. The method of item 1, wherein said cell comprising a modified foreign chromosome(s) or a fragment(s) thereof is an animal cell.

5. The method of item 4, wherein said animal cell is a mammalian cell.

6. The method of item 4, wherein said animal cell is a non-human animal cell.

7. The method of item 1, wherein said targeting vector contains a telomere sequence which is introduced into a desired site by insertion of the targeting vector.

8. The method of item 2, wherein said deletion occurs at a site into which said telomere sequence has been introduced.

9. The method of item 1, wherein said targeting vector comprises a recognition sequence for a site-directed recombination enzyme, and said recognition sequence is introduced into a desired site by insertion of the targeting vector.

10. The method of item 9, wherein a vector, which is capable of expressing a site-directed recombination enzyme, is introduced into said cell with high homologous recombination efficiency simultaneously with or after insertion of said targeting vector comprising the recognition sequence for a site-directed recombination enzyme, so that an activity of said site-directed recombination enzyme is expressed, resulting in deletion and/or translocation of said foreign chromosome(s) or a fragment(s) thereof at a site in which said recognition sequence for a site-directed recombination enzyme is introduced.

11. The method of item 10, wherein said translocation occurs between a plurality of foreign chromosomes or fragments thereof.

12. The method of item 11, wherein a plurality of said foreign chromosomes are derived from the same species.

13. The method of item 12, wherein said same species is a human.

14. The method of item 11, wherein a plurality of said foreign chromosomes are derived from different species.

15. The method of item 14, wherein said species are a human and a mouse.

16. The method of item 10, wherein said translocation occurs between a foreign chromosome(s) or a fragment(s) thereof and a chromosome(s) derived from said cell with high homologous recombination efficiency.

17. The method of item 9, wherein said site-directed recombination enzyme is a Cre enzyme.

18. The method of item 9, wherein said recognition sequence for a site-directed recombination enzyme is a LoxP sequence.

19. The method of item 1, wherein said cell with high homologous recombination efficiency is an embryonic stem cell (or ES cell).

20. The method of item 1, wherein said cell with high homologous recombination efficiency is a chicken DT-40 cell.

21. The method of item 1, which further comprises a step of screening a cell comprising said foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred.

22. The method of item 21, wherein said screening is based on expression of a marker gene.

23. The method of item 22, wherein said marker gene is a drug-resistance gene.

24. The method of item 22, said marker gene is a green fluorescent protein-encoding gene derived from the jellyfish *Aequorea victoria* or a modified gene thereof.

25. The method of item 1, wherein said foreign chromosome(s) or a fragment(s) thereof is derived from a human.

26. A method for producing a chimeric non-human animal comprising a modified foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

(a) preparing a microcell comprising a foreign chromosome(s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell with high homologous recombination efficiency through its fusion with said microcell;

(b) in said cell with high homologous recombination efficiency, inserting a vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof, and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking said desired site;

(c) in said cell with high homologous recombination efficiency, causing deletion and/or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof; and

(d) preparing a microcell comprising said foreign chromosome(s) or a fragment(s) thereof in which deletion or translocation has occurred, and transferring said foreign chromosome(s) or a fragment(s) thereof into a pluripotent non-human animal cell through its fusion with said microcell.

27. The method of item 26, wherein a plurality of said cells with high homologous recombination efficiency are subjected to whole cell fusion after steps (a) and (b) and are subjected to step (c).

28. The method of item 27, wherein a plurality of said cells with high homologous recombination efficiency each comprise a distinct foreign chromosome(s) or a fragment(s) thereof.

29. The method of item 26, wherein said targeting vector comprises a telomere sequence which is introduced into a desired site by insertion of the targeting vector.

30. The method of item 29, wherein said deletion occurs at a site where said telomere sequence has been introduced.

31. The method of item 26, wherein said targeting vector comprises a recognition sequence for a site-directed recombination enzyme, and said recognition sequence is introduced into a desired site by insertion of the targeting vector.

32. The method of item 31, wherein a vector, which is capable of expressing a site-directed recombination enzyme, is introduced into said cell with high homologous recombination efficiency simultaneously with or after insertion of said targeting vector comprising said recognition sequence for a site-directed recombination enzyme, so that an activity of said site-directed recombination enzyme is expressed, resulting in deletion and/or translocation of said foreign chromosome(s) or a fragment(s) thereof at a site into which said recognition sequence is introduced.

33. The method of item 32, wherein said translocation occurs between a plurality of foreign chromosomes or fragments thereof.

34. The method of item 32, wherein said translocation occurs between said foreign chromosome(s) or a fragment(s) thereof and said chromosome(s) derived from said cell with high homologous recombination efficiency.

35. The method of item 31, wherein said site-directed recombination enzyme is a Cre enzyme.

36. The method of item 31, wherein said recognition sequence for site-directed recombination enzyme is a LoxP sequence.

37. The method of item 26, wherein said cell with high homologous recombination efficiency is an embryonic stem cell (or ES cell).

38. The method of item 26, wherein said cell with high homologous recombination efficiency is a chicken DT-40 cell.

39. The method of item 26, which further comprises a step of screening cells comprising a foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred.

40. The method of item 39, wherein said screening is based on expression of a marker gene.

41. The method of item 40, wherein said marker gene is a drug-resistance gene.

42. The method of item 40, the marker gene is a green fluorescent protein-encoding gene derived from the jellyfish *Aequorea victoria* or a modified gene thereof.

43. The method of item 26, wherein in the step (d), a microcell is produced from said cell with high homologous recombination efficiency; said foreign chromosome(s) or a fragment(s) thereof, in which deletion and/or translocation has occurred is transferred into a CHO cell through its fusion with said microcell; a microcell is produced from the CHO cell; and then said foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred is transferred into a pluripotent cell through its fusion with said microcell.

44. The method of item 26, said pluripotent cell is an embryonic stem cell (or ES cell).

45. The method of item 26, said foreign chromosome(s) or a fragment(s) thereof is derived from a human.



46. A method for producing a non-human animal comprising a modified foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

(a) preparing a microcell comprising a foreign chromosome(s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell with high homologous recombination efficiency through its fusion with said microcell;

(b) in said cell with high homologous recombination efficiency, inserting a vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof, and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking said desired site;

(c) in said cell with high homologous recombination efficiency, causing deletion and/or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof;

(d) preparing a microcell comprising said foreign chromosome(s) or a fragment(s) thereof, in which deletion and/or translocation has occurred, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell derived from a non-human animal through its fusion with said microcell; and

(e) transplanting the nucleus of said cell derived from the non-human animal into an enucleated unfertilized egg derived from a homologous non-human animal of the same species.

47. The method of item 46, wherein a plurality of said cells with high homologous recombination efficiency are subjected to whole cell fusion after steps (a) and (b) and are subjected to the step (c).

48. The method of item 47, wherein a plurality of said cells with high homologous recombination efficiency comprise a distinct foreign chromosome(s) or a fragment(s) thereof.

49. The method of item 46, wherein said targeting vector comprises a telomere sequence, which is introduced into a desired site by insertion of the targeting vector.

50. The method of item 49, wherein said deletion occurs at a site into which a telomere sequence has been introduced.

51. The method of item 46, wherein said targeting vector comprises a recognition sequence for site-directed recombination enzyme, and said recognition sequence is introduced into a desired site by insertion of the targeting vector.

52. The method of item 51, wherein a vector, which is capable of expressing a site-directed recombination enzyme, is introduced into said cell with high homologous recombination efficiency simultaneously with or after insertion of said targeting vector comprising said recognition sequence for a site-directed recombination enzyme, so that an activity of said site-directed recombination enzyme is expressed, resulting in deletion and/or a translocation of said foreign chromosome(s) or fragment(s) thereof at a site into which said recognition sequence is introduced.

53. The method of item 52, wherein said translocation occurs between a plurality of foreign chromosomes or fragments thereof.

54. The method of item 52, wherein said translocation occurs between said foreign chromosome(s) or a fragment(s) thereof and said chromosome derived from a cell with high homologous recombination efficiency.

55. The method of item 51, wherein said site-directed recombination enzyme is a Cre enzyme.

56. The method of item 51, wherein said recognition sequence for a site-directed recombination enzyme is a LoxP sequence.

57. The method of item 46, wherein said cell with high homologous recombination efficiency is an embryonic stem cell (or ES cell).

58. The method of item 46, wherein said cell with high homologous recombination efficiency is a chicken DT-40 cell.

59. The method of item 46, which further comprises a step of screening cells containing a foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred.

60. The method of item 59, wherein said screening is based on expression of a marker gene.

61. The method of item 60, wherein said marker gene is a drug-resistant gene.

62. The method of item 60, wherein said marker gene is a green fluorescent protein-encoding gene derived from the jellyfish *Aequorea victoria* or a modified gene thereof.

63. The method of item 46, wherein, in the step (d), a microcell is produced from said cell with high homologous recombination efficiency; said foreign chromosome(s) or fragment(s) thereof, in which deletion and/or translocation have/has occurred, is/are transferred into a CHO cell through its fusion with the microcell; a microcell is produced from the CHO cell; and then said foreign chromosome(s) or a fragment(s) thereof, in which deletion and/or translocation has occurred, is transferred into a cell derived from a non-human animal through its fusion with the microcell.

64. The method of item 46, said cell derived from a non-human animal is a culture cell derived from an embryo or a blastocyst.

65. The method of item 46, said cell derived from a non-human animal is a culture cell derived from a fetus or an adult.

66. The method of item 46, said cell derived from a non-human animal is a fibroblast cell derived from fetus.

67. The method of item 46, said foreign chromosome(s) or a fragment(s) thereof is derived from a human.

68. A non-human animal, which retains a chromosomal fragment(s) obtained by deletion of a foreign chromosome(s) or a fragment(s) thereof.

69. The non-human animal of item 68, wherein said chromosomal fragment(s) comprises:

- (i) a marker gene and a telomere sequence, and/or
- (ii) a recognition sequence for a site-directed recombination enzyme.

70. A non-human animal, comprising a recombinant foreign chromosome(s) obtained by translocation between a plurality of foreign chromosomes or fragments thereof.

71. The non-human animal of item 70, wherein said recombinant chromosomal fragment(s) comprises:

- (i) a marker gene and a telomere sequence; and/or
- (ii) a recognition sequence for a site-directed recombination enzyme.

72. The non-human animal of item 70, wherein said recombinant foreign chromosome(s) is independently maintained in the nucleus of the non-human animal cell.

73. The non-human animal of item 70, wherein said recombinant foreign chromosome(s) is derived from a human.

74. The non-human animal of item 70, wherein the recombinant foreign chromosome(s) is derived from human chromosomes #14 and #2.

75. The non-human animal of item 70, wherein said recombinant foreign chromosome(s) is derived from human chromosomes #14 and #22

76. The non-human animal of item 70, wherein said recombinant foreign chromosome(s) comprises genes for a heavy-chain and a light-chain A of a human antibody.

77. The non-human animal of item 70, wherein said recombinant foreign chromosome(s) comprises genes for a heavy-chain and a light-chain  $\kappa$  gene of a human antibody.

78. The non-human animal of item 70, which is a mouse.

79. The non-human animal of item 70, which is an ungulate.

80. The non-human animal of item 70, which is a bovine.

81. The non-human animal of item 70, which is an ovine.

82. The non-human animal of item 70, which is an avian.

83. The non-human animal of item 70, which is a chicken.

84. A cell, comprising a recombinant chromosome(s) or a fragment(s) thereof obtained by deletion and/or translocation of a chromosome(s) or a fragment(s) thereof, which comprises at least a part of a human chromosome and into which

- (i) a marker gene and telomere sequence, and/or
- (ii) a recognition sequence for site-directed recombination enzyme is/are introduced.

85. A method for modifying a foreign chromosome(s) or a fragment(s) thereof in a cell, which comprises the steps of:

- (a) preparing a microcell containing a foreign chromosome(s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell with high homologous recombination efficiency through its fusion with the microcell;
- (b) in said cell with high homologous recombination efficiency, inserting a targeting vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking said desired site; and
- (c) in said cell with high homologous recombination efficiency, causing deletion or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof.

86. An artificial chromosome vector, which comprises a centromere sequence derived from a human chromosome #14 or #21, and a recognition sequence for a site-directed recombination enzyme.

87. The artificial chromosome vector of item 86, wherein said recognition sequence for a site-directed recombination enzyme is a LoxP sequence.

88. A recombinant chromosome or a fragment thereof obtained by deletion and/or translocation of a chromosome(s) or a fragment(s) thereof, into which

- (i) a marker gene and a telomere sequence, and/or
- (ii) a recognition sequence for a site-directed recombination enzyme is/are introduced, and which comprises at least a part of a human chromosome.

89. The recombinant chromosome or a fragment thereof of item 88, which comprises fragments of human chromosomes #14 and #22.

90. The recombinant chromosomes or a fragment thereof of item 88, which comprises fragments of human chromosomes #14 and #2.

91. The recombinant chromosomes or a fragment thereof of item 88, which comprises genes for a heavy-chain and a light-chain  $\lambda$  of a human antibody.

92. The recombinant chromosomes or a fragment thereof of item 88, which comprises genes for a heavy-chain and a light-chain  $\kappa$  of a human antibody.

[0048] The present specification includes the contents as disclosed in the specification and/or drawings of Japanese Patent Application No. 10-236169, which is a priority document of the present application.

## Brief Description of Drawings

[0049]

Fig.1 shows the results of PCR analysis of an A9 cell retaining human chromosome #2 (fragment).  
 Fig.2 shows that human chromosome #22 (fragment) is retained in an E14 drug resistant cell (PCR analysis).  
 Fig. 3 is a photograph of electrophoresis patterns showing that human L1 sequence is retained in a chimeric mouse produced from a human chromosome #22-transferred ES cell (Southern analysis).  
 Fig. 4 is a photograph of electrophoresis patterns showing the presence of a human chromosome in organs of a human chromosome #22 transferred chimeric mouse (PCR analysis).  
 Fig. 5 is a photograph of electrophoresis patterns showing the results of the expression of human genes in a human chromosome #22 transferred chimeric mouse (RT-PCR).  
 Fig. 6 is a photograph of electrophoresis patterns showing the results of the expression of human genes in organs of a human chromosome #22 transferred chimeric mouse (RT-PCR).  
 Fig. 7 shows that human chromosome #4 (fragment) is retained in an E14 drug resistant cell (PCR analysis).  
 Fig. 8 is a photograph of electrophoresis patterns showing the detection of human L1 sequence in a human chromosome #4-transferred E14 cell clone (Southern analysis).  
 Fig. 9 is a photograph of electrophoresis patterns showing that human L1 sequence is retained in a chimeric mouse produced from a human chromosome #4-transferred ES cell (Southern analysis).  
 Fig. 10 shows that human chromosome #14 (fragment) is retained in a TT2 drug resistant cell (PCR analysis).  
 Fig. 11 is a photograph of electrophoresis patterns showing the presence of a human chromosome in organs of a chimeric mouse produced from a human chromosome #14 transferred ES cell (PCR analysis).  
 Fig. 12 shows the results of a test on a tail-derived fibroblast cell for resistance to G418.  
 Fig. 13 shows the concentration of human antibody IgM in a serum of a human serum albumin (hereinafter referred to as "HSA")-immunized chimeric mouse (ELISA).  
 Fig. 14 shows the concentration of human antibody IgG in a serum of an HSA-immunized chimeric mouse (ELISA).  
 Fig. 15 shows the results of ELISA of hybridoma clone H4B7 capable of producing human IgM.  
 Fig. 16 is a photograph of the results of FISH analysis of a mouse ES cell clone (TT2 cell clone PG15) retaining partial fragments of human chromosomes #2 and 14.  
 Fig. 17 shows that the antibody titer of anti-HSA human IgG is increased in a serum of an HSA-immunized chimeric mouse.  
 Fig. 18 shows that the antibody titer of anti-HSA human Ig  $\kappa$  is increased in a serum of an HSA-immunized chimeric mouse.  
 Fig. 19 is a photograph of electrophoresis patterns showing the detection of human L1 sequence in a human chromosome #22-transferred TT2 cell clone (Southern analysis).  
 Fig. 20 shows that the antibody titer of anti-HSA human Ig  $\lambda$  is increased in a serum of an HSA-immunized chimeric mouse.  
 Fig. 21 is a photograph showing that a partial fragment of human chromosome #2 is retained in a progeny of a chimeric mouse into which a partial fragment of a human chromosome #2 was transferred (PCR analysis).  
 Fig. 22 shows the presence of a cell expressing human  $\mu$  chain on the cell surface in a spleen of a human chromosome #14-transferred chimeric mouse (flow cytometry analysis).  
 Fig. 23 shows the structure of LoxP-pstNEO plasmid DNA.  
 Fig. 24 shows the structure of genomic DNA carrying a mouse antibody heavy chain C $\mu$  gene.  
 Fig. 25 shows the structure of genomic DNA carrying a mouse antibody light-chain  $\kappa$  gene.  
 Fig. 26 shows the structures of a mouse antibody heavy-chain targeting vector and a probe for Southern blotting, as well as a DNA fragment to be detected in homologous recombinants.  
 Fig. 27 shows the structures of a mouse antibody light-chain  $\kappa$  targeting vector and a probe for Southern blotting, as well as a DNA fragment to be detected in homologous recombinants.  
 Fig. 28 is a photograph of electrophoresis patterns showing the results of Southern blot analysis of mouse antibody heavy-chain homologous recombinants and high concentration G418 resistant clones derived therefrom.  
 Fig. 29 shows a photograph of electrophoresis patterns showing the results of Southern blot analysis of mouse antibody light-chain homologous recombinants.  
 Fig. 30 shows the structure of pLoxP-PGKPuro plasmid DNA.  
 Fig. 31 shows a mouse antibody light-chain  $\kappa$  targeting vector, a probe for use in the southern blot analysis of genomic DNA from transformant TT2F cells, and DNA fragments to be detected in homologous recombinants.  
 Fig. 32 shows a photograph of electrophoresis patterns showing the results of Southern blot analysis of high concentration G418 resistant cell clones derived from mouse antibody light-chain homologous recombinants.  
 Fig. 33 shows that the antibody titers of anti-HSA human IgH antibodies are increased in a serum of an HSA-

immunized chimeric mouse.

Fig. 34 is a photograph of the result of FISH analysis of an antibody heavy- and light-chains deficient mouse ES cell clone retaining partial fragments of human chromosomes #2 and #14.

Fig. 35 shows that the antibody titers of anti-HSA human Ig antibodies are increased in a serum of an HSA-immunized chimeric mouse.

Fig. 36 is a photograph showing the result of FISH analysis of a mouse A9 cell containing human chromosome #14 (human centromere sequence probe).

Fig. 37 is a photograph showing the result of FISH analysis of a mouse A9 cell containing human chromosome #14 (human chromosome-specific probe).

Fig. 38 shows the results of a test for stability of human chromosome fragments (#14: SC20, #2:W23) in a mouse ES cell.

Fig. 39 shows the results of analysis for stability of human chromosome #14 fragments in a mouse.

Fig. 40 shows the results of PCR analysis of a G418 resistant hybrid cells retaining human chromosome #22 (fragment).

Fig. 41 is a photograph showing the results of FISH analysis of an A9 cell retaining fragmented human chromosome #22.

Fig. 42 shows the results of complete human antibody-producing mouse strains established by mating.

Fig. 43 shows the results of the determination of the concentration of human antibody  $\kappa$  chain in a serum of a mouse retaining a human chromosome #2 fragment, W23.

Fig. 44 shows the results of the determination of the concentration of human antibody  $\kappa$  and  $\lambda$  chains in a serum of a mouse.

Fig. 45 shows the structure of pBS-TEL/LIFPuro.

Fig. 46 shows that human chromosome #22 is retained in a chicken DT40 cell clone.

Fig. 47 shows the identification of homologous recombinant in LIF locus.

Fig. 48 shows the fragmentation of human chromosome #22 in a DT40/#22neo cell clone.

Fig. 49 is a photograph showing a chicken DT40 cell clone retaining full length or fragmented human #22 chromosome.

Fig. 50 shows an increase in the anti-TNF- $\alpha$  human Ig antibody titer in the sera of TNF- $\alpha$ -immunized chimeric mice.

Fig. 51 shows the response to ASIALO-GM1 in chimeric mice.

Fig. 52 shows the response to GM2 in chimeric mice.

Fig. 53 shows the results of FACS analysis for single Tc/KO mouse peripheral blood nuclear cells.

Fig. 54 shows the results of measurements of human Ig concentration in the sera of single Tc/KO mice.

Fig. 55 shows the results of measurements of human Ig subclass concentration in the sera of single Tc/KO mice.

Fig. 56 shows a rise in the anti-HSA human Ig antibody titer in the sera of HSA-immunized double Tc/KO mice.

Fig. 57 shows a rise in the anti-HSA human Ig $\kappa$  antibody titer in the sera of HSA-immunized double Tc/KO mice.

Fig. 58 shows the outline of the production of human artificial chromosomes  $\lambda$  - HAC and  $\kappa$  - HAC.

Fig. 59 shows the outline of production of human artificial chromosomes  $\lambda$  - HAC and  $\kappa$  - HAC - introduced mice.

Fig. 60 shows the structure of cassette vectors ploxPHYg and ploxPbsr.

Fig. 61 shows the structure of a targeting vector pHCF2loxPHYg(F) and a method for identifying a homologous recombinant.

Fig. 62 shows the structure of a targeting vector pHCF2loxPHYg(R) and a method for identifying a homologous recombinant.

Fig. 63 shows the structure of a targeting vector pRNR2loxPbsr and a method for identifying a homologous recombinant.

Fig. 64 shows the structure of a targeting vector pYHZloxPHYg(F) and a method for identifying a homologous recombinant.

Fig. 65 shows the structure of a cassette vector pTELPuro.

Fig. 66 shows the structure of a targeting vector pTELPuroCD8A(F) and a method for identifying a homologous recombinant.

Fig. 67 shows the structure of a targeting vector pTELPuroCD8A(R) and a method for identifying a homologous recombinant.

Fig. 68 shows the presence of a genetic marker on human chromosome 2 and the presence of a genetic marker on human chromosome 2 in a clone CD10 in which telomere was truncated at CD8A locus.

Fig. 69 shows the results of FISH for the clone CD10 using a pGKPuro probe.

Fig. 70 shows the structure of a vector, pBS185hisD, for stable expression of Cre recombination enzyme.

Fig. 71 shows the structure of an artificial chromosome after translocation between loxPs of RHF clone and of RHR clone.

Fig. 72 shows PCR identification for confirming the structure of a genomic region after translocation between loxP

in RHF clone and confirming the translocation.

Fig. 73 shows the results of PCR for confirming translocation.

Fig. 74 shows the results of FISH for RHF clone using a pGKPuro probe and a 14qter specific probe, and results of FISH for the same clone using a chromosome 22-specific probe and a chromosome 14-specific probe.

Fig. 75 shows a rise in anti-HSA human IgY antibody titer and in Igλ antibody titer in the sera of HAS-immunized chimeric mice C-10.

#### Best Mode for Carrying Out the Invention

**[0050]** The present invention will now be described in detail.

**[0051]** A mouse that retains a human chromosome(s) or a fragment(s) thereof and which expresses the gene on the chromosome(s) or fragment(s) thereof can be produced by

- (1) preparing a chromosome donor cell which retains a labeled human chromosome or a fragment thereof;
- (2) transferring the human chromosome or fragment thereof into a murine pluripotent cell by microcell fusion;
- (3) producing a chimeric mouse from the cell; and
- (4) confirming that the human chromosome is retained in the chimeric mouse and that a human gene is expressed.

**[0052]** In this procedure, a mouse is used as a non-human animal that retains a human chromosome or a fragment thereof and which expresses the gene on the chromosome or fragment thereof (the mouse is hereinafter referred to as a "human chromosome transferred mouse").

**[0053]** The term "human chromosome" means a naturally occurring complex which consists of nucleic acids and proteins that are derived from human cells. There are 46 normal human chromosomes of 23 kinds (24 kinds in male), each of which contains DNAs of about 50-300 Mb in size. In the present invention, the human chromosome includes not only partial fragments which can be stably replicated and segregated as independent chromosomes but also fragments that are translocated on mouse chromosomes and which are retained stably. The size of the DNA is usually at least 1 Mb and in some cases, it is smaller than 1 Mb. The feature of the present invention resides in that a mouse can retain and express the foreign gene on a foreign chromosome as a mediator without treatments such as cloning in an E. coli or yeast cell, or extraction of the DNA from a cell.

**[0054]** The term "human chromosome transferred mouse" means a mouse retaining a human chromosome(s) or a fragment(s) thereof in all or part of its normal somatic cells. The mouse expresses the gene(s) on a human chromosome(s) or a fragment(s) thereof in all or part of its normal somatic cells.

- (1) Preparation of a chromosome donor cell which retains a labeled human chromosome or a fragment thereof

**[0055]** A desired chromosome donor cell 1) retains a human chromosome (s) labeled with a marker also available for selection of recipient cells; 2) does not contain other human chromosomes; and 3) has a higher ability to form a microcell.

**[0056]** Any human-derived cell lines, cancer cells and primary culture cells can be used as materials for providing human chromosomes. Among them, normal fibroblast cells are suitable because they have a low possibility of abnormality such as deletion and amplification of chromosomes and can be readily cultured.

**[0057]** As for 1), human cells can be transformed with vectors that express genes for markers such as drug-resistance (e.g., G418-, puromycin-, hygromycin- or blasticidin-resistance). Promoters operating efficiently not only in human cells but also in recipient cells such as mouse ES cells are desirably used to regulate the expression of the marker used. For this purpose, herpes simplex virus thymidine kinase promoter linked with SV 40 enhancer (Kato et al., Cell Struct. Funct., 12:575, 1987), mouse PGK-1 promoter (Soriano et al., Cell, 64:693, 1991) and the like can be used. A library of human cell transformants in which the introduced marker genes have been inserted into 46 human chromosomes of 23 kinds at random can be prepared by transformation through electroporation (Ishida et al., "Cell Technology Experiment Manual", published by Kodansha, 1992) and the like and subsequent selection of transformants.

**[0058]** As for 3), since many human normal cells have a very low ability to form microcells, the whole cell of the transformant may be fused with a cell having a high ability to form microcells such as mouse A9 cell (Oshimura, M., Environ. Health Perspect., 93:57, 1991) so as to provide the transformed cell with an ability to form microcells. It is known that in mouse-human hybrid cells, human chromosomes selectively disappear. The fused cell selected by the marker can retain stably the marked human chromosome.

**[0059]** In order to meet the condition of 2), it is desired to obtain a microcell from the fused cell and fuse it again with a mouse A9 cell. In this case, too, most of the cells selected by the marker will meet the three conditions 1), 2) and 3) above. The marked human chromosomes can be identified in the finally obtained mouse-human monochromosomal hybrid cells by PCR (Polymerase Chain Reaction, Saiki et al., Science, 239:487, 1988), Southern blot analysis (Ausubel

et al., Current protocols in molecular biology, John Wiley & Sons, Inc., 1994), FISH analysis (Fluorescence In Situ Hybridization, Lawrence et al., Cell, 52:51, 1988) and the like. If the transfer of a specified chromosome is desired, the above procedures are applied to each of many human cell transformant clones to select a clone in which a chromosome of interest is marked. Alternatively, the above procedures are applied to a mixture of human cell transformant clones and the identification of human chromosomes is carried out on a large number of the resulting mouse-human mono-chromosome hybrid cells.

[0060] In addition, a marker gene can be inserted into a desired site by homologous recombination of a specific DNA sequence on the chromosome which is to be transferred (Thomas et al., Cell, 51:503, 1987).

[0061] A microcell prepared from the mouse-human hybrid cell may be irradiated with  $\gamma$ -rays such that the marked human chromosome is fragmented and transferred into a mouse A9 cell. Even if the microcell is not irradiated with  $\gamma$ -rays, a partially fragmented human chromosome may be transferred at a certain frequency. In these cases, the resulting microcell fused clones retain partial fragments of the marked human chromosomes. These clones can be used when it is desired to transfer the partial fragments into recipient cells.

[0062] Human chromosomes to be introduced into ES cells may be modified by deletion, translocation, substitution and the like. Specific procedures for these modifications are as follows:

1) In each of the steps of preparing the aforementioned mouse-human hybrid cell, inducing a microcell from the mouse-human hybrid cell, further fusing the microcell with a mouse A9 cell, inducing a microcell from the further fused cell and fusing the latter microcell with a mouse ES cell, deletion and/or translocation of human chromosomes may occasionally occur. Cells retaining such mutated chromosomes are selected under the microscopic observation of chromosomes or by use of PCR, Southern analysis, or the like. A clone retaining a desired mutant chromosome can be selected from a mouse A9 library retaining various human chromosomes. A clone retaining a desired mutant chromosome can be selected from A9 or ES cell fused with a microcell induced from a mouse A9 cell retaining a certain human chromosome. The frequency of fragmentation of chromosomes can be raised by  $\gamma$ -ray irradiation (Koi et al., Science, 260:361, 1993).

2) A targeting vector retaining a loxp sequence that is recognized by Cre enzyme is constructed. A clone into which a loxp sequence has been inserted at a desired site on a chromosome is obtained by homologous recombination in a cell retaining a human chromosome. Subsequently, Cre enzyme is expressed in the cell of the clone to select a mutant having chromosomal deletion and/or translocation caused by site-directed recombination. See WO97/49804 and Smith et al., Nature Genetics, 9:376, 1995. As a host into which a targeting vector is to be introduced, a cell allowing for high-frequency homologous recombination such as a chicken DT40 cell (Dieken et al., Nature Genetics, 12:174, 1996) may also be used.

3) A targeting vector retaining a human telomere sequence is constructed and the telomere sequence is inserted in the cell at a desired site on a chromosome by homologous recombination in a cell retaining a human chromosome. After a clone into which the telomere sequence has been inserted is obtained, a mutant having deletion caused by the telomere truncation is obtained. See Itzhaki et al., Nature Genet., 2, 283-287, 1992 and Brown et al., P. N. A. S., 93:7125, 1996. As a host into which a targeting vector is to be introduced, a cell allowing for high-frequency homologous recombination such as a chicken DT40 cell (Dieken et al., supra) may also be used. Telomere truncation of human chromosomes in a chicken DT40 cell is first disclosed in the present invention. Brown (supra) discloses that a vector was inserted into a repeat sequence on a chromosome. However, no specific site can be targeted. Itzhaki et al. discloses that tumor cells, i.e., 12000 cells of cell line HT1080 into which a telomere sequence was introduced were analyzed and 8 homologous recombinants were obtained. They found that out of the 8 cells, only one caused deletion by insertion of the telomere sequence. For some kinds of cells, results were reported that no mutant having truncation was obtained by insertion of a telomere sequence into some kinds of cells (Barnett et al., Nucleic Acids Res., 21:27, 1993). In spite of this report, the inventors believed that it was necessary to increase the absolute number of homologous recombinants in order to obtain mutants having truncation and made an attempt to perform telomere truncation using a chicken DT40 cell as a host. As a result, it was surprisingly found that truncation occurred in all of the 8 homologous recombinants obtained.

[0063] As mentioned above, a gene that should not be expressed in a human chromosome-transferred mouse can be removed by modification of a introduced chromosome. If the size of a chromosome to be transferred is shortened by fragmentation, the chromosome fragment to be transferred can be transmitted to progenies of the chromosome-transferred mice. In addition, using chromosome translocation and substitution techniques, genes derived from a plurality of chromosomes can be expressed on the same chromosome fragment and portions of a plurality of genes on the chromosome fragments can be replaced with different genes. In other words, foreign chromosome fragments can be used as vectors for transferring genes into individual mice and their cells.

A more detailed description of a method for artificially modifying human chromosomes will now be given.

[0064] To date, an accidentally fragmented human chromosome or a full length human chromosome has been in-

roduced into a mouse (Tomizuka et al., Nature Genet., 16: 133, 1997). In this case, some of the introduced human chromosomes may be unstable in a mouse, or may affect development of the mouse so that a human chromosome-introduced mouse itself cannot be generated. Furthermore, it is assumed that an introduced human chromosome cannot be transmitted to its progeny since the presence of an abnormal chromosome in a human or a mouse is thought to inhibit meiosis (Tomizuka et al., Nature Genet., 16: 133, 1997). In fact, Tomizuka et al. reported that male chimeric mice retaining a fragment which contains a region approximately half a human chromosome 14 (about 100 Mb) became infertile, and this fragment was not transmitted to progeny (Nature Genet., 16: 133, 1997). On the other hand, small chromosomal fragments, such as a fragment W23 of chromosome 2, (it is thought to be 5 to 20 Mb; Rastan, Nature Genet., 16: 113, 1997) as described in Tomizuka et al.'s report and a fragment SC20 of chromosome 14, (somewhat larger than W23) as shown in Examples of this specification, could be transmitted to progeny. That is, the use of smaller chromosomal fragments may increase the possibility for them to be transmitted to progeny.

**[0065]** In order to produce a mouse, which contains a target region of a human chromosome, stably retains it, and then transmits it to the progeny, there is a need to develop techniques which make it possible to freely process a chromosome. That is, techniques that do not depend on accidentally obtained chromosomal fragments, but enables us to cleave a human chromosome at a desired site or to combine only a desired chromosomal fragment with another stable chromosome. These techniques are called chromosome engineering, and have been used to date for generating a mutant mouse having a modified chromosome, by site-directed cleavage of an endogenous mouse chromosome mainly in a mouse ES cell (WO 98/54348), or by causing recombination (translocation) between homologous chromosomes so as to cause deletion, inversion, or multiplication of a specific genetic region (R. Ramirez-Solis et al., Nature 378: 720, 1995). There have been no reports on production of a non-human animal, such as a mouse, having a human chromosome(s) artificially modified in these various ways. Recently, it has been reported that an artificial chromosome vector (Mammalian Artificial Chromosome, MAC), which is stably retained in a mammalian cell, has been developed (for example, W. Mills et al., Human Molecular Genetics 8: 751, 1999). However, there has been no report on a universal construction of an artificial chromosome, which allows cloning of a genomic region exceeding the cloning size of YAC vector.

**[0066]** This invention provides a universal system for constructing human artificial chromosomes, which will allow cloning of any chromosomal fragments showing production of a human artificial chromosome (HAC) into which a human antibody gene cluster is cloned as an example.

The production of human artificial chromosomes  $\lambda$ -HAC and  $\kappa$ -HAC having human antibody  $\lambda$  light-chain and  $\kappa$  light-chain gene clusters introduced thereto will be described as follows. It will include the introduction of the produced HAC into a mouse, expression of human antibody genes contained in HAC in a mouse, and transmission of HAC to the progeny of chimeric mice.

#### A. Modification of human chromosome 22

**[0067]** A human antibody  $\lambda$  light-chain gene cluster is present at 22q11.2 on chromosome 22 (for example, J.E. Collins et al., Nature 377: 367, 1995). To use only the periphery of this region of the chromosome for production of an artificial chromosome, first, a human telomere sequence is inserted by a homologous recombination into LIF locus closely linked to a  $\lambda$  light-chain gene cluster in the telomere side resulting in telomere-directed truncation to cleave the chromosome (for example, Kuroiwa et al., Nucleic Acid Research, 26: 3447, 1998). Next, a recognition sequence, loxP sequence, of a recombination enzyme, Cre, is inserted by homologous recombination into the HCF2 locus closely linked to the  $\lambda$  light-chain gene cluster in the centromere side. Here, only HCF2-Ig $\lambda$ -LIF fragments are used for the production of artificial chromosomes. By cloning only fragments of chromosome 22, causative genetic regions on the chromosome 22 of Cats Eye syndrome (for example, Johnson, A et al., Genomics 57: 306, 1999) and Digeeorge syndrome (for example, H. Yamagishi et al., SCIENCE 283: 1158, 1999), which may affect the development of a mouse, can be eliminated.

#### B. Modification of human chromosome 2

**[0068]** A human antibody  $\kappa$  light-chain gene cluster is present at 2p11.2 on chromosome 2 (for example, Gottfried M. et al., Genomics 16: 512, 1993). To use only the periphery of this region of the chromosome for production of an artificial chromosome, first, a human telomere sequence is inserted by homologous recombination into CD8A locus (for example, Gottfried M. et al., Genomics 16: 512, 1993) closely linked to the  $\kappa$  light-chain gene cluster in the telomere side. This insertion results in telomere-directed truncation to cleave the chromosome. Next, a recognition sequence, loxP sequence, of a recombination enzyme, Cre, is inserted by homologous recombination into the cosYHZ304 genomic region closely linked to the  $\kappa$  light-chain gene cluster in the centromere side. Here, only cosYHZ304-Ig $\kappa$ -CD8A fragment is used for production of artificial chromosomes.



## C. Modification of human chromosome 14 fragment SC20

[0069] It has been shown that human chromosome 14 fragment SC20, is retained almost 100% in a mouse and can be transmitted to progeny. Therefore, in this invention, SC20 is used as a human artificial chromosome vector. That is, the translocation of HCF2-Ig $\lambda$ -LIF fragment of human chromosome 22 and cosYHZ304-Ig $\kappa$ -CD8A fragment of human chromosome 2 as described above in A and B to SC20 resulting in construction of human artificial chromosomes  $\lambda$  and  $\kappa$ -HAC having the fragments of the human chromosomes 22 and 2 cloned thereto.

First, a loxP sequence is inserted by homologous recombination into RNR2 locus located at 14p12 of chromosome 14. The Cre-loxP system (for example, R. Ramirez-Solis et al., Nature 378:720, 1995) is applied for cloning the HCF2-Ig $\lambda$ -LIF fragment of human chromosome 22 and cosYHZ304-Ig $\kappa$ -CD8A fragment of human chromosome 2 into the RNR2 locus on SC20 by translocation.

[0070] For efficient telomere-directed truncation or homologous recombination as described in A, B, and C above, a chicken DT-40 cell can be used as a host cell (for example, Kuroiwa et al., nucleic Acid Research 26: 3447, 1998, and Dieken et al., Nature Genet., 12: 174, 1996).

[0071] Furthermore, it was found that human chromosome 21 (fragment) screened from a monochromosome hybrid cell library that has been constructed by the methods described in Tomizuka et al.'s report (Nature Genet., 16: 133, 1997) was also very stable in a chimeric mouse. That is, a chimeric mouse was produced from mouse ES cells, which retain chromosomal fragments containing most of human chromosome 21 (chimerism: 95 % or more). The retention rate of the chromosome in a brain cell, a hepatic cell, and a fibroblast derived from the tail of this chimeric mouse were all 95% or more. Hence, it is concluded that human chromosome 21 and fragments thereof can be used as a human artificial chromosome vector, as in the above SC20.

D. Translocation of HCF2-Ig $\lambda$ -LIF fragment of human chromosome 22 and cosYHZ304-Ig $\kappa$ -CD8A fragment of human chromosome 2 onto SC20 by Cre-loxP system

[0072] As described in A, B and C above, a chicken DT-40 cell, which retains either a fragment of the modified human chromosome 22 or 2, or a modified SC20, is produced. To translocate a HCF2-Ig $\lambda$ -LIF fragment of human chromosome 22 or cosYHZ304-Ig $\kappa$ -CD8A fragment of human chromosome 2 onto SC20 by the Cre-loxP system, chicken DT-40 cells, each retaining either one modified human chromosomal fragment, are fused to each other. A chicken DT-40 hybrid cell retaining two modified human chromosomal fragments is produced. In this way, all the materials required for translocation are prepared.

Next, the Cre recombination enzyme is expressed in the chicken DT-40 hybrid cell to cause translocation. The recombination (translocation) frequency between two non-homologous chromosomes is reported to be very low (for example, A.J. Smith et al., Nature Genet., 9:376, 1995). Moreover in this invention, recombination efficiency may be lower since the translocation occurs between exogenous human chromosomes, not between endogenous chromosomes. Accordingly, a positive selection system, which can select a cell in which recombination occurs between loxPs as expected, must be devised.

[0073] In recent years, A. J. H. Smith et al. (Nature Genet., 9:376, 1995) succeeded in causing translocation to occur between chromosomes 12 and 15 in a mouse ES cell using the Cre-loxP system. They inserted a portion on the 5' side (including loxP) and a portion on the 3' side (including loxP) of the Hprt gene into specific regions of chromosomes 12 and 15 in a Hprt (hypoxanthine - guanine - phosphoribosyl transferase) gene-deficient ES cell, respectively, by homologous recombination. Since a Hprt gene is re-constructed only when recombination occurs between loxPs after the transient expression of Cre, the ES cells can grow even in a medium (HAT medium) lacking hypoxanthine. However, this selection method can be used only for Hprt gene-deficient cells. That is, the method cannot be used for all cells. As Qin, M. et al. (PNAS 91: 1706, 1994) reported, there is a selection method, by which, instead of Hprt genes, appropriate drug-resistant genes are reconstructed as a result of translocation, and target cells grow in a medium containing the drug. In this invention, the inventors have devised the following system that can be applied for any type of cell and allows rapid concentration of target cells.

[0074] In recent years, a GFP (Green Fluorescent Protein) gene (for example, Prasher D. C. et al., Gene, 111: 229, 1992) and a modified gene thereof (for example, Heim et al., Nature, 373: 663, 1995) have been developed as reporter genes for introduction of a gene into an animal cell. Since emission of GFP requires no substrate and GFP can be detected with fluorescence, live cells can be monitored in a short time. Even when the expression amount of GFP is low, GFP is gradually accumulated to a detectable level in a cell because of its stability. Furthermore, the use of FACS allows detection of very weak fluorescence. Therefore, in this invention, a GFP gene is used as a positive selection marker for loxP recombinant. For example, a GFP gene containing no promoter is inserted together with loxP into RNR2 locus on SC20, and then a promoter required for the expression of GFP is inserted together with loxP into HCF2 locus on human chromosome 22, or into a cosYHZ304 genomic region on chromosome 2. When recombination between loxP sequences occurs by Cre, the promoter and GFP gene are joined together so that GFP is expressed. The

recombinants can be sorted by FACS because they emit fluorescence. Since expression of GFP can be detected in a short time as described above, repetition of FACS sorting enables concentration of GFP positive cells faster than selection using a drug.

To increase the recombination frequency, it is assumed that the Cre recombination enzyme is expressed stably, not transiently. In this case, a chromosome, in which translocation occurs, may go back to the initial state by re-recombination (or re-translocation) but with a low frequency, because translocation between two chromosomes occurs from each other. Hence, such an experiment generally needs a transient expression of Cre recombination enzyme, or strict control over the expression of Cre recombination enzyme. However, in this invention, a target human artificial chromosome, in which translocation occurs, is transferred to a Chinese hamster CHO cell immediately after translocation in a DT40 hybrid cell by the microcell-mediated chromosome transfer (herein after referred to as "MMCT."), so that the target chromosome in the CHO cell can avoid the effect of Cre recombination enzyme. Therefore, no more strict expression control is required and Cre recombination enzyme can simply be expressed stably. As described below, the present invention is the first one that has succeeded in translocation between two non-homologous human chromosomes in a non-human cell (a chicken DT-40 cell in this invention).

**[0075]** As disclosed in A, B, C and D above, the use of a series of telomere-directed truncations and translocations between chromosomes by the Cre-loxP system in a chicken DT-40 cell enables the cloning of human chromosomal fragments with any size (exceeding the cloning size of YAC vector) into loxP site (14p12) on stable SC20 chromosome vector. Thus the inventors can announce a human artificial chromosome-constructing system. The human artificial chromosome-constructing system as disclosed herein is performed preferably using chicken DT-40 cells because the frequency of homologous recombination is very low in normal cultured animal cells. On the other hand, a mouse ES cell is known as another animal cell with a high frequency of homologous recombination other than chicken DT-40 cells (Shinichi Aizawa, Bio Manual Series 8, Gene Targeting, Yo-do sha, 1995). Thus mouse ES cells can also be used in constructing human artificial chromosomes according to this invention. However, to culture mouse ES cells, which have an ability to differentiate into various tissues, while maintaining them in an undifferentiated state, complex procedures like culture of vegetative cells, and other skills are required (Shinichi Aizawa, supra). Furthermore, mouse ES cells can differentiate depending on the types of chromosomes to be introduced (W097/07671). Accordingly, chicken DT-40 cells are preferred for the construction of human artificial chromosomes because mouse ES cells have the problems as described above.

**[0076]** In addition to recombination between human chromosomes as illustrated in examples, the translocation of a desired region (or a fragment) of a human chromosome to a mouse chromosome derived from a mouse ES cell is also possible. This can be done by the procedures (1) to (4) as shown below.

(1) A recognition sequence, for example a loxP sequence, of a site-directed recombination enzyme is inserted by homologous recombination into a chromosome of a mouse ES cell. A targeting vector comprising the loxP sequence contains a promoter sequence to express a marker, such as the following GFP. A preferable insertion site for the loxP sequence is, for example a subtelomere region of a mouse chromosome in order to avoid an effect on a gene of the mouse.

(2) The following procedures A and B are performed in chicken DT-40 cells which retain human chromosomes containing desired regions.

(2)-A: A telomere sequence is inserted on the telomere side of a desired region on the human chromosome to shorten this chromosome. This procedure may not be required when said desired region is located near the original telomere of said human chromosome.

(2)-B: A recognition sequence, for example, a loxP sequence, is inserted on the centromere side of the desired region on the human chromosome. A targeting vector comprising the loxP sequence comprises, for example a GFP gene.

(3) The human chromosome (fragment) as produced in (2) above is transferred to a CHO cell by MMCT. Then the human chromosomal fragments are transferred from the CHO cell to the mouse ES cell prepared in (1) above.

(4) Site-directed recombination enzyme is expressed in the mouse ES cell comprising the human chromosome (or a fragment) as prepared in (3) above. In a cell wherein site-directed recombination occurs between the loxP sequences, the promoter sequence and GFP gene are joined together, and GFP is expressed. Thus these cells can be selected.

The above procedure enables mouse ES cells that have a chromosome comprising a desired human chromosomal region (or a fragment) to be obtained through translocation. From the mouse ES cells, a chimeric mouse or its progeny, stably retaining the human chromosomal region (fragment), can be produced.

**[0077]** The human artificial chromosome produced by the above human artificial chromosome-constructing system is preferably introduced into a Chinese hamster CHO cell before its introduction into mouse ES cells. When Dieken et al. (Nature, Genet., 12:174, 1996) transferred modified human chromosomes from chicken DT-40 cells to mouse MEL

cells (a kind of cancer cell), most were transferred in a fragmented state. When the inventors tried to transfer human chromosomes from chicken DT-40 cells into mouse A9 cells and the like, however, fragmentation was observed for all the human chromosomes and no intact chromosomes could be transferred. However, upon transfer into Chinese hamster CHO cells, the inventors found for the first time that intact human chromosomes can be transferred. Therefore,

the human artificial chromosomes prepared by this human artificial chromosome-constructing system, are temporarily transferred to CHO cells.

CHO cells are known to form microcells efficiently as mouse A9 cells do (for example, M. Koi et al., SCIENCE 260: 361, 1993). It should enable us to transfer human artificial chromosomes from CHO cells to mouse ES cells and to produce chimeric mice retaining the human artificial chromosomes. Furthermore, it is known that SC20 fragments themselves can be transferred to progeny. Thus it is naturally expected that human artificial chromosomes ( $\lambda$ ,  $\kappa$  - HAC) produced by this system using these chromosomal fragments can be transferred to progeny.

**[0078]** In the afore-mentioned report of Tomizuka et al. (Nature Genet., 16: 133, 1997), the human chromosome introduced into a mouse was derived from a human fibroblast. After introduction into a mouse ES cell via a mouse A9 cell, it was functional in both a chimeric mouse and its progeny. Expression of the human gene on the introduced chromosome at the transcription level and the protein level was confirmed for, for example, a human antibody gene (Tomizuka et al., Nature Genet., 16: 133, 1997). To date, it is still unknown if human chromosomes introduced into mouse cells via the cells of Aves, which is evolutionally distant from mammals, can keep their ability to function in mouse cells. Moreover, apart from being derived from birds, chicken DT-40 cells have a very high homologous recombination efficiency (Takeda et al., Proc. Natl. Acad. Sci. USA, 89: 023, 1992). That is, a human gene on an introduced human chromosome may be inactivated by gene conversion between the human gene and homologous bird gene on a bird chromosome. Concerning this problem, there is only one report (Dieken et al., Nature Genet., 12: 174, 1996) that when human chromosome 11 was transferred to a mouse MEL cell via a chicken DT-40 cell, transcripts of the human  $\beta$ -globin gene were detected. There is no report on the expression of a human gene at the protein level and the function of expressed human proteins.

This invention shows that a human gene on a human chromosome transferred via a chicken DT-40 cell, and a protein encoded by this gene are functional in a mouse. Human immunoglobulin  $\lambda$  chain protein was detected in the sera of chimeric mice retaining human chromosome 22 fragment (comprising a human immunoglobulin  $\lambda$  chain gene) that had been genetically engineered in chicken DT-40 cells. Furthermore, in response to immunization with human serum albumin (HSA), increased antibody titer including antigen-specific human  $\lambda$  chain was confirmed. That is, the expressed human immunoglobulin  $\lambda$  chain protein is present as a constituent of a functional antibody molecule.

## (2) Transfer of the human chromosome or fragment thereof into a mouse pluripotent cell

**[0079]** It has been reported to date that an embryonic carcinoma cell (EC cell, Hanaoka et al., Differentiation, 48:83, 1991), an embryonic stem cell (ES cell, Evans, Nature, 292:154, 1981) or an embryonic germ cell (EG cell, Matsui et al., Cell, 70:841, 1992) that are derived from various strains of mice contribute to the normal somatic cells in mice, or are capable of the production of chimeric mice, by injection into or coculturing with a mouse early embryo. ES and EG cells have a very high ability in this respect and in many cases, they also contribute to germ cells thereby making it possible to produce progenies derived from the cells. EC cells can be obtained predominantly from teratocarcinoma; ES cells from the inner cell masses of blastocysts; and EG cells from primordial germ cells appearing at the early stage of embryogeny. These cell lines and their mutants, and any undifferentiated cells that are capable of differentiation into all or part of the normal somatic cells in mice can be used as recipient cells for the transfer of human chromosome in the present invention. In these recipient cells, for the purpose of achieving advantageous expression of a human gene to be introduced, a gene or genes such as a mouse gene homologous to the human gene can be disrupted in a chimeric mouse or a chimeric-mouse derived tissue or cell by using homologous recombination in gene targeting (Joyner et al., Gene Targeting, 1993, IRL PRESS) or other techniques.

**[0080]** The microcells prepared from the human chromosome donor cells obtained in item (1) or the microcells irradiated with  $\gamma$ -rays can be used as materials for the transfer of human chromosomes into the recipient cells. The human chromosome can be transferred into the recipient cell through fusion of the recipient cell with the microcell by the method described in Motoyuki Shimizu, "Cell Technology Handbook", published by Yodosha, 1992. The microcell donor cells retain markers by which human chromosomes or fragments thereof can be selected in the recipient cells. The clone containing a gene, a chromosome or a fragment of interest can be selected by PCR, Southern blot analysis, FISH method or the like in the same manner as in (1), thus all kinds of human chromosomes or fragments thereof can be transferred. Moreover, if several chromosomes or fragments thereof which contain different selection markers are transferred sequentially, a recipient cell retaining these chromosomes or fragments at the same time can be obtained. In addition, clones having an increased number of the transferred chromosome can be selected from the clones into which the human chromosome has been transferred. Such selection can be accomplished by increasing the concentration of a selection drug to be added to a culture medium.

[0081] In order to determine whether the recipient cell selected by the marker (e.g., G418 resistance) on the human chromosome retains the whole or part of the chromosome retained by the donor cell, the following confirmative techniques may be employed: Southern blot analysis using the genomic DNA extracted from the selected recipient cell, with a human specific repeated sequence (L1, Alu, etc.: Korenberg et al., Cell, 53:391, 1988) or a human gene used as a probe; and chromosome analysis such as PCR method using a human gene specific primer or FISH method using a human chromosome specific probe (Lichter et al., Human Genetics, 80:224, 1988).

(3) Production of a chimeric mouse from the human chromosome transferred ES cell

[0082] The method described in Shinichi Aizawa, "Biotechnology Manual Series 8, Gene Targeting", published by Yodosha, 1995 may be used to produce chimeric mice from the ES cell clone obtained in (2). In selecting factors for efficient production of chimeric mice, such as the developmental stage of the host embryo and its strain, it is desired to employ the conditions already reviewed for the respective ES cell clones. For example, 8-cell stage embryos derived from Balb/c (albino, CREA JAPAN, INC.) or ICR (albino, CREA JAPAN, INC.) are desirably used for CBAXC57BL/6 F1-derived TT2 cell (agouti, Yagi et al., Analytical Biochemistry, 214:70, 1993).

(4) Confirmation of the retention of the human chromosome in the chimeric mice and the expression of a human gene

[0083] The contribution of the ES cells in mice produced from the embryos into which ES cells were injected can be roughly judged by the color of their coat. However, it should be noted that the total absence of contribution to the coat color does not always lead to the conclusion that there is no contribution to other tissues. The detailed information on the retention of the human chromosome in various tissues of the chimeric mice can be obtained by Southern blot analysis using the genomic DNA extracted from various tissues, by PCR or the like.

[0084] The expression of the gene on the transferred human chromosome can be confirmed by the following methods. The expression of mRNA transcribed from the human chromosome can be detected by RT-PCR method or northern blotting (Ausubel et al., supra) using RNAs derived from various tissues (Kawasaki et al., P.N.A.S., 85:5698, 1988). The expression at the protein level can be detected by enzyme immunoassay using an anti-human protein antibody that is rendered minimal in its ability to enter into a cross reaction with mouse homologous proteins (ELISA, Toyama and Ando,

"Monoclonal Antibody Experiment Manual", published by Kodansha Scientific, 1987; Ishikawa, "Enzyme immunoassay with Superhigh Sensitivity", published by Gakkai Shuppan Center, 1993), western blotting (Ausubel et al., supra), isozyme analysis utilizing the difference in electrophoretic mobility (Koi et al., Jpn. J. Cancer Res., 80:413, 1989) or the like. The retention of the human chromosome in the chimeric mice and the expression of the gene on the human chromosome can be confirmed by the appearance of the cells expressing a drug resistance marker gene in primary culture cells derived from the chimeric mice.

[0085] For example, human IgM, IgG, IgA and the like in sera of the chimeric mice which are produced from ES cells retaining human chromosome #14 on which a gene for human immunoglobulin heavy chain exists can be detected by enzyme immunoassay using an anti-human Ig antibody that is rendered minimal in its ability to enter into cross reaction with mouse antibody. Hybridomas capable of producing a human immunoglobulin heavy chain can be obtained by ELISA screening of hybridomas prepared by immunizing the chimeric mouse with a human-derived antigen (e.g., HSA) and fusing the spleen cells of the immunized mice with mouse myeloma cells (Toyama and Ando, "Monoclonal Antibody Experiment Manual", published by Kodansha Scientific, 1987).

[0086] The method for producing a chimeric non-human animal of the present invention has been explained above with reference to the case of a mouse retaining a human chromosome(s) or a fragment(s) thereof and expressing the gene(s) on the chromosome(s) or fragment(s). In the present invention, chromosomes or fragments thereof to be transferred into chimeric non-human animals are not limited to those derived from humans but include any foreign chromosomes and fragments thereof. The term "foreign chromosome" means a chromosome which is transferred into a pluripotent cell and, subsequently, the gene on which (or a fragment thereof) is expressed in a chimeric non-human animal. The organism species from which the foreign chromosome is derived is not particularly limited. Other kinds of chimeric animals such as chimeric rat and pig can be produced by the method of the present invention. ES cells or ES-like cells derived from animals other than mouse were established with rat (Iannaccone et al., Dev. Biol., 163, 288-, 1994), pig (Wheeler et al., Reprod. Fertil. Dev., 6, 563-, 1994) and bovine (Sims et al., Proc. Natl. Acad. Sci. USA, 91, 6143-6147, 1994) and attempts have been made on cyprinodont, chicken and the like ("Transgenic Animal", Protein Nucleic Acid Enzyme, October, 1995, Special Issue, published by Kyoritsu Shuppan). It is known that sheep is developed normally from an unfertilized egg transplanted with the nucleus from ES-like cell (ED cell) or epithelial-like cell obtained by subcultivation of the ES-like cell through at least 10 generations (Campbell et al., Nature, 380, 64-, 1996). These ES cells and ES-like cells can be used as recipient cells in the transfer of foreign chromosomes to produce chimeric non-human animals retaining the foreign chromosomes or fragments thereof and expressing the genes on

the chromosomes or fragments thereof in the same manner as in the case of mouse.

[0087] Recently, rapid progression of nuclear transplantation technology allows us to use the somatic cells from a fetus or adult, more differentiated than the above ES or ES-like cells, as donors for nuclear transplantation. There have been reports that viable progeny could be obtained by nuclear transplantation into unfertilized eggs using mammary glandular cells of an adult sheep (Wilmut et al., Nature, 385: 810, 1997), the fibroblasts of a fetal calf (Schnieke et al., Science, 278:2130, 1997), or the cumulus cells of an adult mouse (Wakayama et al., Nature, 394: 369, 1998) as donors. Moreover, transgenic sheep (Schnieke et al., supra) and transgenic bovine (Cibelli et al., Science 280: 1256, 1998) can also be generated using somatic cells, into which cloned foreign DNA has been introduced, as donors.

[0088] Therefore, it is concluded that nuclear transplantation using somatic cells, into which foreign chromosomes or foreign recombinant chromosomes have been introduced, as donors enables the generation of an animal that retains and expresses the foreign chromosomes or the foreign recombinant chromosomes. This can be done, for example by the following procedures (1) to (5).

(1) The following steps A and B are performed in a chicken DT-40 cell that retains human chromosome 22 comprising an Ig $\lambda$  region.

(1)-A: A telomere sequence is inserted on the telomere side of the Ig $\lambda$  region on human chromosome 22 to shorten the chromosome.

(1)-B: A recognition sequence of a site-directed recombination enzyme, for example a loxP sequence, is inserted on the centromere side (for example HCF locus) of the Ig $\lambda$  region on human chromosome 22. A targeting vector comprising the loxP sequence comprises a promoter sequence for the expression of a marker, such as a GFP gene as shown below.

(2) A chicken DT-40 cell retaining human chromosome 14 fragment SC20 in which a loxP sequence and GFP gene are inserted on RNR2 locus, and the chicken DT-40 cell produced in (1) above is subjected to the whole cell fusion, thereby obtaining chicken DT-40 cells retaining two types of human chromosomal fragments at the same time.

(3) The site-directed recombination enzyme is expressed in a mouse ES cell retaining the human chromosome (fragment) produced in (2) above. The promoter sequence and the GFP gene are joined together in the cell, wherein site-directed recombination occurs between the loxP sequences, so that the GFP is expressed. That is, chicken DT-40 cells comprising recombinant human chromosomes resulting from translocation can be selected.

(4) The recombinant human chromosome (fragment) as produced in (3) above is transferred to a CHO cell by MMCT. A microcell is induced from the CHO cell retaining the obtained recombinant human chromosome (fragment), and then the microcell is fused with, for example a fibroblast derived from a fetal calf. These fibroblasts, retaining the recombinant human chromosomal fragments, can be selected by, for example the expression of a drug-resistant marker, present on the recombinant human chromosome.

(5) Nuclear transplantation using the fibroblast derived from a fetal calf, retaining the recombinant human chromosomal fragments obtained in (4) above as a donor, and using a bovine unfertilized egg as a recipient, allows the generation of a bovine and offspring retaining and expressing the recombinant human chromosomal fragments (Cibelli et al., Science 280: 1256, 1998).

[0089] In the present invention, pluripotent cells into which a foreign chromosome(s) or a fragment(s) thereof are transferred are not limited to the ES cells, EC cells and EG cells mentioned above. For example, it is possible to transfer a foreign chromosome(s) or a fragment(s) thereof into bone marrow stem cells. If these bone marrow stem cells are transplanted into a living organism, hereditary diseases, etc. may be treated.

[0090] If an ES cell retaining a foreign chromosome(s) or a fragment(s) thereof is differentiated to a germ cell in the chimeric non-human animal, reproduced progenies will retain the transferred chromosome(s) or fragment(s) thereof and express the gene(s) on the chromosome(s) or fragment(s) thereof.

[0091] The chimeric non-human animals or their progenies can be used to express the gene on the foreign chromosome or fragment thereof and to recover the expression product, thereby producing a biologically active substance. More specifically, the chimeric non-human animals or their progenies can be bred under the conditions for expressing the gene on the foreign chromosome or fragment thereof to recover the expression product from the blood, ascites and the like of the animals. Alternatively, the tissues or cells of the chimeric non-human animal, or immortalized cells derived therefrom (e.g., hybridomas immortalized by fusion with myeloma cells) can be cultured under the conditions for expressing the gene on the foreign chromosome or fragment thereof and the expression product is thereafter recovered from the culture. Furthermore, a foreign chromosome(s) or a fragment(s) thereof which was extracted from tissues or cells of these chimeric non-human animals or their progenies, or from immortalized cells derived therefrom; the DNA which is a component of said foreign chromosome(s) or fragment(s) thereof; or cDNA derived from the foreign chromosome(s) or fragment(s) thereof retained in tissues or cells of the chimeric non-human animals or their progenies, or in immortalized cells derived therefrom may be used to transform animal cells or insect cells (e.g., CHO cells, BHK

cells, hepatoma cells, myeloma cells, SF9 cells) and the transformed cells may be cultured under the conditions for expressing the gene on the foreign chromosome(s) or fragment(s) thereof to recover the expression product (e.g., an antibody protein specific to a particular antigen) from the culture. The expression product can be collected by known techniques such as centrifugation and purified by known techniques such as ammonium sulfate fractionation, partition chromatography, gel filtration chromatography, adsorption chromatography, preparative thin-layer chromatography and the like. The biologically active substance includes any kinds of substances encoded on foreign chromosomes, for example, antibodies, particularly human antibodies. For example, the human antibody gene on the foreign chromosome can be cloned from spleen cells of the chimeric non-human animal or immortalized cells such as hybridomas derived therefrom and transferred into Chinese hamster ovary cells (CHO), myeloma cells or the like to produce a human antibody (Lynette et al., *Biotechnology*, 10:1121-, 1992; Bebbington et al., *Biotechnology*, 10:169-, 1992).

**[0092]** The chimeric mice or their progenies that retain human chromosomes #2, 14 and/or 22 (or fragments thereof) which can be produced by the method of the present invention can retain the greater part of the functional sequences of respective genes for human antibody heavy chain on chromosome #14, light chain  $\kappa$  on chromosome #2 and light chain  $\lambda$  on chromosome #22. Hence, they can produce a wide repertoire of antibodies which are more similar to human antibody repertoire, compared with known transgenic mice into which parts of human antibody gene have been transferred by using yeast artificial chromosomes and the like (Green et al., *Nature Genetics*, 7, 13-, 1994; Lonberg et al., *Nature*, 368, 856-, 1994). Also, the chimeric mice and their progenies retaining two human chromosomes (or fragments) of #2+#14, #22+#14 or other combination and the mice and their progenies retaining three human chromosomes (or fragments) of #2+#14+#22 or other combination which are obtainable by mating said chimeric mice and their progenies retaining two human chromosomes (or fragments), as produced by the method of the invention, can produce complete human antibodies both heavy- and light-chains of which are derived from human. These mice can recognize human-derived antigens as foreign substances to cause an immunoreaction with the antigens, thereby producing antigen-specific human antibodies. These properties can be utilized to produce human monoclonal and polyclonal antibodies for therapeutic treatments (Green et al, supra; Lonberg et al., supra). On the other hand, in order to obtain a human antibody having high affinity for a particular antigen more efficiently, it is desirable to produce a mouse which produces a human antibody but not a mouse antibody (Green et al., supra; Lonberg et al., supra). In the present invention, this is achieved typically by the following Method A or B using known techniques.

Method A: a method using a mouse antibody-deficient ES cell and a mouse antibody-deficient host embryo for chimera production.

Method B: a method in which a progeny retaining a human chromosome is obtained from a human chromosome-transferred chimeric mouse, followed by mating said progeny with a mouse in a strain deficient in a mouse antibody gene.

**[0093]** A typical example for each of Methods A and B will be described below specifically.

Specific procedures for Method A

1. One allele of a mouse antibody heavy-chain gene present in two copies in a mouse ES cell is disrupted by homologous recombination in gene targeting (Joyner et al., "Gene Targeting", published by IRL PRESS, 1993). A marker gene, such as a G 418 resistance gene, sandwiched with two copies of a sequence which can be removed later by site-directed recombination [for example, loxP sequence (see recombination with Cre recombinase in Sauer et al., supra; and see also the use of FLP recombinase-FRT sequence in O'Gorman, *Science*, 251:1351-, 1991)] is inserted at the site where the targeted gene is disrupted.

2. The resultant drug-resistant mouse ES cells in which one allele of an antibody heavy-chain gene was disrupted is cultured in the presence of the drug at a high concentration. Then, those clones which became high concentration drug-resistant are selected. By screening these clones, clones in which both antibody heavy-chain genes were disrupted can be obtained (Shinichi Aizawa, supra).

Alternatively, the other allele of a target gene in the drug-resistant mouse ES cell in which one allele of the antibody heavy-chain gene has been disrupted is also disrupted by homologous recombination. The same procedure may be repeated using a marker gene other than the precedingly inserted marker gene. For example, homologous recombination is performed using a G418-resistance gene, followed by another homologous recombination using a puromycin-resistance gene to obtain clones in which both alleles of the antibody heavy-chain gene have been disrupted. When the same marker as the precedingly inserted marker is used, an enzyme gene that can cause site-directed recombination between recombinant sequences inserted at the both ends of the drug-resistance gene of item 1 is transiently introduced. Subsequently, drug-sensitive clones are selected that are free of the drug-resistance gene that has been inserted in the target gene. Then, a marker gene is inserted again by homologous recombination in gene targeting to obtain clones in which both alleles of the target gene have been disrupted (Seishi Takatsu et al., *Experimental Medicine*, supplement, *Basic Techniques for Immunological Study*, p. 255-, 1995, Yodosha).

3. An enzyme gene (e.g., a Cre recombinase gene (Sauer et al., supra)) which causes a site-directed recombination

between the recombination sequences inserted at both the ends of the drug-resistance gene in step 1 above is transiently transferred into the mouse ES cells from step 2 above in which both antibody heavy-chain genes were disrupted. Then, drug-sensitive clones are selected in which the drug-resistance genes inserted at the sites of both heavy-chain genes were deleted as a result of recombination between the loxP sequences [Seiji Takatsu et al., "Experimental Medicine (extra number): Basic Technologies in Immunological Researches", p. 255-, published by Yodosha, 1995].

4. The same procedures in steps 1-3 above are repeated for the mouse antibody light-chain  $\kappa$  gene to finally obtain drug-sensitive clones which are completely deficient in antibody heavy-chain and light-chain  $\kappa$ .

5. Human chromosome #14 (fragment) containing a human antibody heavy-chain gene and marked with a drug-resistance gene (e.g., G418 resistance gene) is transferred into the clone from step 4 above (antibody heavy-chain and light-chain  $\kappa$ -deficient mouse ES cell) by microcell fusion.

6. Human chromosome #2 (fragment) or #22 (fragment) or both containing a human antibody light-chain gene(s) and marked with a drug-resistance gene different from the one used in step 5 above (e.g., puromycin resistance gene) are transferred into the clone obtained in step 5 above by microcell fusion.

7. Chimeric mice are produced from the ES cells obtained in step 6 above by using embryos obtained from a mouse in a strain having no ability to produce its own antibody (e.g., RAG-2 knockout mouse, Shinkai et al., Cell, 68:855-, 1992; membrane-type  $\mu$  chain knockout mouse, Kitamura et al., Nature, 350:423-, 1991) as host embryos.

8. Most of the functional B lymphocytes in the resultant chimeric mice are derived from the ES cells [Seiji Takatsu et al., "Experimental Medicine (extra number): Basic Technologies in Immunological Researches", p. 234-, published by Yodosha, 1995]. Since those B lymphocytes are deficient in mouse heavy-chain and light-chain  $\kappa$ , they produce human antibodies alone mainly as a result of the expression of the functional human antibody genes on the transferred chromosomes.

#### Specific procedures for Method B

##### [0094]

1. Chimeric mice retaining a human chromosome or a fragment thereof containing human antibody heavy-chain, light-chain  $\kappa$  or light-chain  $\lambda$  are used to produce a progeny which stably retains the human chromosome or fragment thereof and which can transmit it to the next generation.

2. A mouse in a strain which is homozygous regarding the deficiency in mouse antibody heavy-chain and light-chain  $\kappa$  and which retains human chromosomes containing human antibody heavy-chain (#14) + light-chain  $\kappa$  (#2), heavy-chain (#14) + light-chain  $\lambda$  (#22) or heavy-chain (#14) + light-chain  $\kappa$  (#2) + light-chain  $\lambda$  (#22) is obtained by mating the mouse in a strain expressing human antibody heavy-chain or light-chain from step 1 above or a mouse in a strain expressing both human antibody heavy and light-chains obtained by mating the mice from step 1, with a mouse in a strain deficient in its own antibody genes (e.g., the membrane-type  $\mu$  chain knockout mouse mentioned above; light-chain  $\kappa$  knockout mouse, Chen et al., EMBO J., 3:821-, 1993). Since mice in the resultant strain are deficient in mouse antibody heavy-chain and light-chain  $\kappa$  genes, they produce human antibodies alone mainly as a result of the expression of the functional human antibody genes on the transferred chromosomes.

[0095] Both Method A and Method B may be used not only to yield human antibodies but also to yield products of any genes located on a foreign chromosome efficiently.

[0096] The present invention will now be explained in greater detail with reference to the following examples, which do not limit the scope of the present invention.

#### Example 1

##### Production of chromosome donor cell retaining human chromosome (fragment) labeled with G418 resistance

[0097] Plasmid pSTneoB containing a G418 resistance gene (Katoh et al., Cell Struct. Funct., 12:575, 1987; Japanese Collection of Research Biologicals (JCRB), Deposit Number: VE 039) was linearized with restriction enzyme Sall (TAKARA SHUZO CO., LTD.) and introduced into human normal fibroblast cell HFL-1 (obtained from RIKEN Cell Bank, RCB0251). The HFL-1 cells were treated with trypsin and suspended in Dulbecco's phosphate-buffered saline (PBS) at a concentration of  $5 \times 10^6$  cells/ml, followed by electroporation using a Gene Pulser (Bio-Rad Laboratories, Inc.) in the presence of  $10 \mu\text{g}$  of DNA (Ishida et al., "Cell Technology Experiment Procedure Manual", published by Kodansha, 1992). A voltage of 1000 V was applied at a capacitance of  $25 \mu\text{F}$  with an Electroporation Cell of 4 mm in length (165-2088, Bio-Rad Laboratories, Inc.) at room temperature. The electroporated cells were inoculated into an Eagle's F12 medium (hereinafter referred to as "F12") supplemented with 15% fetal bovine serum (FBS) in 3-6 tissue culture

plastic plates (Corning) of 100 mm  $\phi$ . After one day, the medium was replaced with a F12 supplemented with 15% FBS and containing 200  $\mu$ g/ml of G418 (GENENTICIN, Sigma). The colonies formed after 2-3 weeks were collected in 52 groups each consisting of about 100 colonies. The colonies of each group were inoculated again into a plate of 100 mm  $\phi$  and cultured.

**[0098]** Mouse A9 cells (Oshimura, Environ. Health Perspect., 93:57, 1991; JCRB 0211) were cultured in Dulbecco's modified Eagle's medium (hereinafter referred to as "DMEM") supplemented with 10% FBS in plates of 100 mm $\phi$ . The G418 resistant HFL-1 cells of 52 groups were cultured in F12 supplemented with 15% FBS and 200  $\mu$ g/ml of G418 in plates of 100 mm $\phi$ . The mouse A9 cells and HFL-1 cells were treated with trypsin and one fourth to one half of both cells were mixed. The mixed cells were inoculated into a plate of 100 mm $\phi$  and cultured in a mixture of equal amounts of DMEM containing 10% FBS and F12 containing 15% PBS for a period ranging from a half day to one day. Cell fusion was carried out in accordance with the method described in Shimizu et al., "Cell Technology Handbook", published by Yodosha, p. 127-, 1992. The cell surface was washed twice with DMEM and then treated sequentially with 2 ml of a PEG (1:1.4) solution for 1 minute and with 2 ml of PEG (1:3) for 1 minute. After the PEG solution was sucked up, and the cells were washed three times with a serum-free DMEM, followed by cultivation in DMEM supplemented with 10% FBS for 1 day. The cells were dispersed by treatment with trypsin and suspended in a double selective medium (10% FBS supplemented DMEM) containing ouabain ( $1 \times 10^{-5}$  M, Sigma) and G418 (800  $\mu$ g/ml), followed by inoculation in 3 plates of 100 mm $\phi$ . After about 3 weeks cultivation, the colonies formed were treated with trypsin to disperse the cells, which were cultured in a selective medium (10% FBS supplemented DMEM) containing G418 (800  $\mu$ g/ml).

**[0099]** The cells were dispersed by treatment with trypsin and two groups of the cells were collected, followed by cultivation in 6 centrifuge flasks (Coaster, 3025) of 25 cm<sup>2</sup> until the cell density reached 70-80% confluence. The medium was replaced with a medium (20% FBS supplemented DMEM) containing Colcemid (0.05  $\mu$ g/ml, Demecolcine, Wako Pure Chemicals Co., Ltd) and the cells were cultured for 2 days to form microcells. After the culture medium was removed, a cytochalasin B (10  $\mu$ g/ml, Sigma) solution preliminarily warmed at 37°C was filled in the 25 cm<sup>2</sup> centrifuge flask, which were inserted into an acryl centrifuge container, followed by centrifugation at 34°C at 8,000 rpm for 1 hour. The microcells were suspended in a serum-free medium and purified by passage through a filter. To the mouse A9 cells cultured to 80% confluence in the flask of 25 cm<sup>2</sup>, the purified microcells were added and the two kinds of cells were fused with a PEG solution. The fused cells were cultured in a G418 containing selective medium and colonies formed were isolated. Human chromosomes #2, 4, 14 and 22 retained in the respective clones were identified by the methods described in (1)-(3) below. All other experimental conditions such as operating procedures and reagents were in accordance with Shimizu et al., "Cell Technology Handbook", published by Yodosha, p127-.

#### (1) PCR analysis

**[0100]** The isolated cells were cultured and genomic DNA was extracted from the cells with a Puregene DNA Isolation kit (Gentra System Co.). PCR was performed using the genomic DNA as a template with human chromosome specific primers to select the clones retaining human chromosome #2, 4, 14 or 22. The PCR amplification was conducted with about 0.1  $\mu$ g of the genomic DNA as a template, using a thermal cycler (GeneAmp 9600, Perkin-Elmer Corp.) in accordance with the method described in Innis et al., "PCR Experiment Manual", published by HBJ Publication Office, 1991. Taq polymerase was purchased from Perkin-Elmer Corp. and the reaction was performed in a cycle of 94°C, 5 minutes and 35 cycles of denaturing at 94°C, 15 seconds, annealing at 54-57°C, 15 seconds (variable with the primers) and extension at 72°C, 20 seconds. The gene on each chromosome (O'Brien, Genetic Maps, 6th edition, Book 5, Cold Spring Harbor Laboratory Press, 1993) and polymorphic markers (Polymorphic STS Primer Pair, BIOS Laboratories, Inc.; Weissenbach et al., Nature 359:794, 1992; Walter et al., Nature Genetics, 7:22, 1994) were used as primers. The primers for the genes were prepared on the basis of nucleotide sequences obtained from data bases such as GenBank, EMBL and the like. The names of the polymorphic primers and the sequences of the primers for the genes will be shown for the respective chromosomes in the following examples (#2, Example 1; #4, Example 6, #14, Example 9; #22, Example 2). The following genetic markers and polymorphic makers (Polymorphic STS Primer Pairs: D2S207, D2S177, D2S156 and D2S159, BIOS Laboratories, Inc.) were used to identify chromosome #2.

C $\kappa$  (immunoglobulin kappa constant) : 5' -

TGGAAGGTGGATAACGCCCT (SEQ ID NO:1), 5' -

TCATTCTCCTCCAACATTAGCA (SEQ ID NO:2)



FABP1 (fatty acid binding protein-1 liver) : 5'-  
 GCAATCGGTCTGCCGGAAGA (SEQ ID NO:3), 5'-  
 5 TTGGATCACTTTGGACCCAG (SEQ ID NO:4 )

Vk3-2 (immunoglobulin kappa variable) : 5'-  
 10 CTCTCCTGCAGGGCCAGTCA (SEQ ID NO:5), 5'-  
 TGCTGATGGTGAGAGTGAAGTC (SEQ ID NO:6)

Vk1-2 (immunoglobulin kappa variable) : 5'-  
 15 AGTCAGGGCATTAGCAGTGC (SEQ ID NO:7), 5'-  
 20 GCTGCTGATGGTGAGAGTGA (SEQ ID NO:8)

## (2) Fluorescence in situ hybridization (FISH)

25 [0101] FISH analysis was conducted with probes specific to human chromosomes #2, 4, 14 and 22 (CHROMOSOME PAINTING SYSTEM, Cambio Ltd.) in accordance with the method described in Matsubara et al., "FISH Experiment Protocol", published by Shujunsha, 1994.

[0102] For example, at least one clone retaining chromosome #2 was obtained in 10 groups out of 26 groups (745 clones). Among them, only 5 clones were positive to all the used primers specific to chromosome #2. FISH analysis was conducted with these clones. FISH analysis was conducted with probes specific to human chromosomes #2 (CHROMOSOME PAINTING SYSTEM, Cambio Ltd.) in accordance with the method described in Matsubara et al., "FISH Experiment Protocol", published by Shujunsha, 1994. In the cells positive to all the primers, an intact form of human chromosome #2 was observed. In some of the clones positive to part of the primers, an independent chromosome smaller than human chromosome #2 was observed or a cell having a chromosome in a form fusing with chromosomes other than human chromosome #2 was observed (Fig. 1). In Fig. 1, the names of the clones are shown in the horizontal line and the primers used in the PCR are shown in the left longitudinal line. ● shows positive clones and X shows negative clones. The forms of human chromosome #2 observed by FISH are shown in the bottom line. No description means no performance of experiment.

[0103] A9 cells retaining human chromosomes #4, 14 and 22 were obtained by the same procedure.

## Example 2

### Transfer of human chromosome #22 into mouse ES cells by microcell fusion

45 [0104] The mouse A9 cell clones retaining human chromosome #22 (hereinafter referred to as "A9/#22") from Example 1 were used as chromosome donor cells. Mouse ES cell line E14 (obtained from Martin L. Hooper; Hooper et al., Nature, 326:292, 1987) was used as a chromosome recipient cell.

E14 cells were cultured in accordance with the method described in Aizawa Shinichi, "Biomannual Series 8, Gene Targeting", published by Yodosha, 1995 and G418 resistant STO cell line (obtained from Prof. Kondo Hisato, Osaka University) treated with mitomycin C (Sigma) was used as a feeder cell. In the first step, microcells were prepared from about 10<sup>8</sup> cells of A9/#22 in accordance with the method reported by Shimizu et al. "Cell Technology Handbook", published by Yodosha, 1992. The total amount of the resulting microcells were suspended in 5 ml of DMEM. About 10<sup>7</sup> cells of E14 were dispersed with trypsin and washed three times with DMEM and suspended in 5 ml of DMEM. The cells were then mixed with the microcells and the mixture was centrifuged at 1,250 rpm for 10 minutes to remove the supernatant. The precipitate was dispersed by tapping and 0.5 ml of a PEG solution (1:1.4) [5 g of PEG 1000 (Wako Pure Chemicals Co., Ltd.) and 1 ml of DMSO (Sigma) as dissolved in 6 ml of DMEM] was added. The mixture was left to stand at room temperature for 1 minute and 30 seconds and 10 ml of DMEM was added slowly. Immediately thereafter, the resulting mixture was centrifuged at 1,250 rpm for 10 minutes to remove the supernatant. The precipitate

was suspended in 30 ml of a medium for ES cells and inoculated into 3 tissue culture plastic plates (Corning) of 100 mm in diameter into which feeder cells were inoculated. After 24 hours, the medium was replaced with a medium supplemented with 300 µg/ml of G418 (GENETICIN, Sigma) and medium replacements were thereafter conducted daily. Drug resistant colonies appeared in 1 week to 10 days. The frequency of appearance was 0-5 per 10<sup>7</sup> of E14 cells. The colonies were picked up and grown. The cells were suspended in a storage medium (a medium for ES cells + 10% DMSO (Sigma)) at a concentration of 5 x 10<sup>6</sup> cells per ml and stored frozen at -80°C. At the same time, genomic DNA was prepared from 10<sup>6</sup>-10<sup>7</sup> cells of each drug resistant clone with a Puregene DNA Isolation Kit (Gentra System Co.).

[0105] Human chromosome #22 was fragmented by irradiating the microcells with Y rays (Koi et al., Science, 260: 361, 1993). The microcells obtained from about 10<sup>8</sup> cells of A9/#22 were suspended in 5 ml of DMEM and irradiated with Y rays of 60 Gy on ice with a Gammacell 40 (Canadian Atomic Energy Public Corporation) at 1.2 Gy/min for 50 minutes. The fusion of γ ray-irradiated microcells and the selection of drug resistant clones were conducted by the same procedure as in the case of the unirradiated microcells. As a result, the frequency of the appearance of the drug resistant clones was 1-7 per 10<sup>7</sup> of E14 cells. The drug resistant clones were stored frozen and DNA was prepared from the clones by the same procedure as in the case of the unirradiated microcells.

[0106] The retention of the transferred chromosomes in the unirradiated microcell-transferred drug resistant clones E14/#22 - 9 and E14/#22 - 10, and in the γ ray-irradiated microcell-transferred drug resistant clones E14/#22-14 and E14/#22-25 was confirmed by the methods described in (1)-(3) below.

#### (1) PCR analysis (Fig. 2)

[0107] The presence of the gene on human chromosome #22 (Genetic Maps, supra) and polymorphic markers (Polymorphic STS Primer Pairs: D22S315, D22S275, D22S278, D22S272 and D22S274, BIOS Laboratories, Inc.; Nature 359:794, 1992) was detected by a PCR method using the genomic DNA of the drug resistant clone as a template. The sequences of oligonucleotide primers for the genes prepared on the basis of nucleotide sequences obtained from data bases such as GenBank, EMBL and the like are described below.

PVALB (parvalbumin) :

5' - TGGTGGCTGAAAGCTAAGAA (SEQ ID NO:9) ,

5' - CCAGAAGAATGGTGTTCATTA (SEQ ID NO:10)

MB (myoglobin) :

5' - TCCAGGTTCTGCAGAGCAAG (SEQ ID NO:11) ,

5' - TGTAGTTGGAGGCCATGTCC (SEQ ID NO:12)

DIA1 (cytochrome b-5 reductase) :

5' - CCCCACCCATGATCCAGTAC (SEQ ID NO:13) ,

5' - GCCCTCAGAAGACGAAGCAG (SEQ ID NO:14)

Igλ (immunoglobulin lambda) :

5' - GAGAGTTGCAGAAGGGGTGACT (SEQ ID NO:15) ,

5' - GGAGACCACCAAACCTCCAAA (SEQ ID NO:16)

ARSA (arylsulfatase A):

5' - GGCTATGGGGACCTGGGCTG (SEQ ID NO:17),

5' - CAGAGACACAGGCACGTAAGAAG (SEQ ID NO:18)

[0108] PCR amplification (Innis et al., supra) was conducted by using about 0.1 µg of the genomic DNA as a template with the above 10 kinds of the primers. As a result, amplification products having expected lengths were detected with all the primers in the case of the two unirradiated clones and with part of the primers in the case of the γ ray-irradiated two clones. The results are shown in Fig. 2. In Fig. 2, a schematic chromosome map based on the G bands of human chromosome #22 and the location of some markers on bands are shown at the left side (O'Brien, GENETIC MAPS, 6th edition, BOOK 5, etc.). The arrangement of the genetic and polymorphic markers shows approximate positional relationships on the basis of the presently available information (Science, HUMAN GENETIC MAP, 1994; Nature Genetics, 7:22, 1994; Nature 359:794, 1992, etc.) and the order is not necessarily correct. With respect to four kinds of the G418 resistant E14 cell clones, the markers for which the expected amplification products were detected by PCR are shown by ■ and the markers for which the expected amplification products were not detected are shown by □. The results of the observation by FISH analysis are shown at the bottom side. A9/#22 is a chromosome donor cell.

## (2) Southern blot analysis

[0109] Southern blot analysis of about 2 µg of the genomic DNA digested with restriction enzyme BglII (TAKARA SHUZO CO., LTD.) was conducted by using human specific repeated sequence L1 ( $10^4$ - $10^5$  copies were present per haploid genome, obtained from RIKEN DNA Bank; Nucleic acids research, 13:7813, 1985; pUK19A derived EcoRI-BamHI fragment of 1.4 kb) as a probe in accordance with the method described in Ausubel et al., Current Protocols in Molecular Biology, John Wiley & Sons, Inc., 1994. As a result, a large number of bands hybridized with the human L1 sequence were detected in DNA of each drug resistant clone. With respect to the unirradiated 2 clones, their patterns and the quantitative ratio of human chromosomal DNA to mouse genomic DNA which could be presumed from the density of the respective bands were the same as those of A9/#22. The total signal intensity of the bands of the γ-ray irradiated clones correlated with the degree of the deletion confirmed by the PCR analysis, as compared with that of A9/#22.

## (3) Fluorescence in situ hybridization (FISH)

[0110] FISH analysis was conducted with probes specific to human chromosomes #22 (CHROMOSOME PAINTING SYSTEM, Cambio Ltd.) in accordance with the method described in Matsubara et al., "FISH Experiment Protocol", published by Shujunsha, 1994. As a result, in almost all of the observed metaphase spreads, human chromosome #22 was detected in the form of translocation to the mouse chromosome with respect to E14/#22-9 and in the form of an independent chromosome with respect to the three other clones.

[0111] The results of the above experiments demonstrate that the obtained G418 resistant clones E14/#22-9 and E14/#22-10 retained all or most part of human chromosome #22 whereas the clones E14/#22-14 and E14/#22-25 retained partial fragments of human chromosome #22.

## Example 3

### Production of chimeric mice from the ES cells retaining human chromosome #22

[0112] General procedures for obtaining mouse embryos, cultivation, injection of the ES cells into the embryos, transplantation to the uteri of foster mothers were carried out in accordance with the method described in Aizawa Shinichi, "Biomaterial Series 8, Gene Targeting", published by Yodosha, 1995. The cells in a frozen stock of the G418 resistant ES clone E14/#22-9 which was confirmed to retain human chromosome #22 were thawed, started to culture and injected into blastocyst-stage embryos obtained by mating a C57BL/6XC3H F1 female mouse (CREA JAPAN, INC.) with a C3H male mouse (CREA JAPAN, INC.); the injection rate was 10-15 cells per embryo. Two and half days after

a foster mother [ICR or MCH(ICR)] mouse (CREA JAPAN, INC) was subjected to a pseudopregnant treatment, about ten of the ES cell-injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 1.

Table 1.

Production of chimeric mice from the ES cells retaining human chromosome #22 (fragments)							
ES cell clone/ human chromosome	G418 resistant clone No.	Number of ES cell- injected blastocyst stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color		
					<-10%	10-30%	30%<
E14/#22	9	166	29	16	7	3	6

**[0113]** As a result of the transplantation of a total 166 of injected embryos, 29 offspring mice were born. Chimerism in the offsprings can be determined by the extent of E14 cell-derived pale gray coat color in the host embryo-derived agouti coat color (dark brown). Out of the 29 offsprings, 16 mice were recognized to have partial pale gray coat color, indicating the contribution of the E14 cells. The maximum contribution was about 40% in K22-22.

**[0114]** These results show that the mouse ES cell clone E14/#22 - 9 retaining human chromosome #22 maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse.

#### Example 4

Confirmation of retention of human chromosomal DNA in various tissues of the chimeric mice derived from the ES cells retaining human chromosome #22

**[0115]** In addition to the determination of coat color in Example 3, the retention of the transferred chromosome was confirmed by PCR analysis using a template genomic DNA prepared from the tail of the chimeric mouse. The tail was obtained from the chimeric mouse at least 3 weeks old in accordance with the method described in Motoya Katsuki, "Development Technology Experiment Manual", published by Kodansha Scientific, 1987. Genomic DNA was extracted from the tail with a Puregene DNA Isolation Kit. Out of the polymorphic primers used in Example 2, PVALB and D22S278 were used, with the extracted genomic DNA as a template, to confirm the amplification products. The analysis was conducted with 10 of the mice in which the contribution to coat color was observed. As a result, the products of amplification with at least either of the primers were detected in all the mice.

**[0116]** Southern blot analysis was conducted in the same manner as in Example 2 by using human L1 sequence as a probe with 2 µg of the genomic DNA derived from the tails of the 6 chimeric mice and one non-chimeric mouse. As a result, the presence of a large number of human L1 sequence was observed in all the chimeric mice and their patterns were similar to those of E14/#22-9. The quantitative ratio to mouse genome was about 10% at maximum (Fig. 3). In Fig. 3, 2 µg of genomic DNA digested with BglII was used in each lane. Human L1 sequence labeled with <sup>32</sup>P was used as a probe and signals were detected with Image Analyzer BAS2000 (Fuji Photo Film Co., Ltd.). The lanes represent the genomic DNA derived from the tails of the chimeric mice (K22-6, 7, 8, 9, 10, 11 and 12; 9 is the non-chimeric mouse) and control DNA (C which is a mixture of E14/#22-9 genomic DNA and E14 genomic DNA at a weight ratio of 1:9) as counted from the right. The DNA molecular weights are shown at the left side and chimerism in the chimeric mice at the right side (- : 0%, +: <10%, and ++: 10-30%).

**[0117]** With respect to the chimeric mouse (K22-7) having about 5% contribution to coat color, genomic DNA was obtained from the brain, liver, muscle, heart, spleen, thymus, ovary and kidney with an ISOGEN (Nippon Gene Co.). For each tissue, PCR analysis was conducted with MB and D1A1 selected from the primers for the genes used in Example 2. As a result, both primers gave expected amplification products in all the tissues. The results of PCR analysis using D1A1 primer are shown in Fig. 4. The PCR products were electrophoresed on a 2% agarose gel and stained with ethidium bromide for detection. The lanes in Fig. 4 represent the following from the left: B, brain; L, liver; SM, skeletal muscle; H, heart; Sp, Spleen; Th, thymus; Ov, ovary; K, kidney; nc, non-chimeric mouse tail-derived DNA (negative control); pc, human fibroblast cell (HFL-1) DNA (positive control).

**[0118]** These results show that E14/#22-9 contributed to various normal tissues in the mouse and that it retained human chromosome #22.

## Example 5

Expression of the human genes in the chimeric mouse derived from the ES cell retaining human chromosome #22

[0119] The tail of the mouse (K22-7) having about 5% contribution to coat color was frozen with liquid nitrogen and then disrupted for use as a sample for confirming the expression of the human genes. The sample was a mixture of tissues such as skin, bones, muscles, blood and the like. Total RNA was extracted from the sample with an ISOGEN (Nippon Gene Co.) and used in an RT-PCR method to detect mRNAs of human myoglobin (MB) and human cytochrome b5 reductase (D1A1). The RT-PCR was performed in accordance with the method described in Innis et al., "PCR Experiment Manual", published by HBJ Publication Office, 1991. Random hexamer oligonucleotides (final concentration: 100 pmol, TAKARA SHUZO CO., LTD.) were used as primers for reverse transcription and Super Script (BRL Co.) as reverse transcriptase. The following primers were used for amplification using cDNA as a template.

MB : 5' - TTAAGGGTCACCCAGAGACT (SEQ ID NO:19) ,

5' - TGTAGTTGGAGGCCATGTCC (SEQ ID NO:20)

D1A1 : 5' - CAAAAAGTCCAACCCTATCA (SEQ ID NO:21) ,

5' - GCCCTCAGAAGACGAAGCAG (SEQ ID NO:22)

[0120] As a result, amplification products specific to mRNAs of both genes were detected (Fig. 5). The RT-PCR products were electrophoresed on a 2% agarose gel and stained with ethidium bromide for detection. In Fig. 5, M is a marker (HindIII digested  $\lambda$  DNA + HaeIII digested  $\phi$  X174DNA, TAKARA SHUZO CO., LTD.); MB, human myoglobin; D1A1, human cytochrome b5 reductase; and WT, a wild-type C3H mouse.

[0121] With respect to the same individual (K22-7), total RNA was extracted from the brain, heart, thymus, liver, spleen, kidney, ovary and skeletal muscle with an ISOGEN and RT-PCR was performed on each organ with the above two primers. As a result, expected products of amplification with D1A1 were observed in all the organs and those with MB were observed only in the heart and skeletal muscle (Fig. 6). Myoglobin is known to be expressed specifically in muscle cells (Bassel-Duby et al., MCB, 12:5024, 1992). Hence, the above results show that the gene on the transferred human chromosome can be subjected to the normal tissue-specific regulation in the mouse. The PCR products were electrophoresed on a 2% agarose gel and stained with ethidium bromide for detection. In Fig. 6, the lanes represent the following from the left: B, brain; H, heart; Th, thymus; L, liver; Sp, spleen; K, kidney; Ov, ovary; SM, skeletal muscle; and M, marker (supra). The lower band observed in the results of MB are believed to represent non-specific products.

[0122] These results show that the transferred human chromosome #22 can function in normal tissues of the chimeric mice.

## Example 6

Transfer of human chromosome #4 or fragments thereof into ES cells

[0123] The mouse A9 cell clone retaining human chromosome #4 (hereinafter referred to as "A9/#4") from Example 1 was used as a chromosome donor cell. Mouse ES cell line E14 (see Example 2) was used as a chromosome recipient cell. The microcell fusion and the selection of G418 resistant clones were conducted by the same procedures as in Example 2. The frequency of the appearance of the drug resistant clones was 1-2 per  $10^7$  of E14 cells. The drug resistant clones were stored frozen and genomic DNA were prepared by the same procedures as in Example 2. The retention of the transferred human chromosome #4 or fragments thereof in the drug resistant clones E14/#4-4, E14/#4-7 and E14/#4-11 was confirmed by the methods described in (1) - (3) below:

## (1) PCR analysis (Fig. 7)

[0124] The presence of the gene on human chromosome #4 (O'Brien, Genetic Maps, 6th edition, Book 5, Cold Spring Harbor Laboratory Press, 1993) and polymorphic markers (Polymorphic STS Primer Pairs: D4S395, D4S412, D4S422, D4S413, D4S418, D4S426 and F11, BIOS Laboratories, Inc., Nature 359:794, 1992) was detected by a PCR method. The sequences of oligonucleotide primers for the genes prepared on the basis of nucleotide sequences obtained from data bases such as GenBank, EMBL and the like will be described below.

HD (huntington disease) : 5'-TCGTTTCCTGTCGAGGATGAA (SEQ ID NO:23), 5'-TCACTCCGAAGCTGCCTTTC (SEQ ID NO:24)

IL-2 (interleukin-2) : 5'-ATGTACAGGATGCAACTCCTG (SEQ ID NO:25), 5'-TCATCTGTAAATCCAGCAGT (SEQ ID NO:26)

KIT (c-kit) : 5'-GATCCCATCGCAGCTACCGC (SEQ ID NO:27),

5'-TTCGCCGAGTAGTCGCACGG (SEQ ID NO:28)

FABP2 (fatty acid binding protein 2, intestinal), 5'-GATGAACTAGTCCAGGTGAGTT (SEQ ID NO:29), 5'-CCTTTTGGCTTCTACTCCTTCA (SEQ ID NO:30)

[0125] PCR amplification was conducted with the above 11 kinds of the primers. As a result, the amplification products having expected lengths were detected with all or part of the primers in all the three clones. In the E14/#4 - 4 and E14/#4 - 7 clones, the deletion of partial regions was observed. The results are shown in Fig. 7. In Fig. 7, a schematic chromosome map based on the G bands of human chromosome #4 and the location of some markers on bands are shown at the left side (see Example 2). The arrangement of the genetic and polymorphic markers shows approximate positional relationships on the basis of the presently available information (see Example 2) and the order is not necessarily correct. With respect to the three kinds of the G418 resistant E14 cell clones, the markers for which the expected amplification products were detected are shown by ■ and the markers for which the expected amplification products were not detected are shown by □. The results of the observation by FISH analysis are shown at the lower side. A9/#4 is a chromosome donor cell.

## (2) Southern blot analysis (Fig. 8)

[0126] Southern blot analysis was conducted by the same procedure as in Example 2 using human L1 sequence as a probe with genomic DNAs obtained from E14/#4-4 and E14/#4-7. As a result, a large number of bands hybridized with the human L1 sequence were detected in DNAs of both drug resistant clones. The total signal intensity correlated with the degree of the deletion confirmed by the PCR analysis, as compared with that of A9/#4. In Fig. 8, 2 µg of genomic DNA digested with BglII was used in each lane. Human L1 sequence labeled with <sup>32</sup>P was used as a probe and the signals were detected with an Image Analyzer (BAS 2000, Fuji Photo Film Co., Ltd.). In Fig. 8, the lanes represent the following as counted from the left: 1, A9/#4 (chromosome donor cell); 2, A9/#4+A9 (1:2); 3, A9/#4+A9 (1:9); 4, A9; 5, E14/#4-7; and 6, E14/#4-4. Lanes 2 and 3 represent mixtures of two kinds of DNAs at the ratios shown in parentheses. The molecular weights of DNAs are shown at the left side.

## (3) Fluorescence in situ hybridization (FISH)

[0127] FISH analysis was conducted with probes specific to human chromosomes #4 (CHROMOSOME PAINTING SYSTEM, Cambio Ltd.) by the same procedure as in Example 2. As a result, in almost all of the observed metaphase spreads of the three clones used, human chromosome #4 or partial fragments thereof were detected in the form of translocation to the mouse chromosome with respect to E14/#4-4 and in the form of an independent chromosome with respect to the two other clones. The relative sizes of the observed human chromosome were consistent with those presumed from the results of the PCR analysis.

[0128] The results of the above experiments demonstrate that the obtained G418 resistant clones retained the whole human chromosome #4 or partial fragments thereof.

## Example 7

## Production of chimeric mice from the ES cells retaining human chromosome #4 fragments

[0129] The cells in frozen stocks of the G418 resistant ES cell clones E14/#4-4 and E14/#4-7 which were confirmed to retain partial fragments of human chromosome #4 were thawed, started to culture, and injected into blastocyst stage embryos obtained by the same method as in Example 3; the injection rate was 10-15 cells per embryo. Two and half days after a foster mother [ICR or MCH(ICR)] mouse (CREA JAPAN, INC.) was subjected to a pseudopregnant treatment, about ten of the ES cell-injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 2.

Table 2.

Production of chimeric mice from the E14 cell clones retaining human chromosome #4 (fragments)							
ES cell clone/ human chromosome	G418 resistant clone No.	Number of ES cell- injected blastocyst stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color		
					<10%	10-30%	30%<
E14/#4	4	160	8	5	5	-	-
	7	80	5	2	1	1	-

[0130] As a result of the transplantation of a total of 240 injected embryos, 13 offspring mice were born. Chimerism in the offsprings can be determined by the extent of E14 cell-derived pale gray coat color in the host embryo-derived agouti coat color (dark brown). Out of the 13 offsprings, 7 mice were recognized to have partial pale gray coat color, indicating the contribution of the E14 cells. The maximum contribution was about 15% in one individual derived from E14/#4-7.

[0131] These results show that the mouse ES cell clones E14/#4-4 and E14/#4-7 which retain fragments of human chromosome #4 maintain the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse.

## Example 8

## Confirmation of retention of human chromosomal DNA in the chimeric mice derived from the ES cells retaining partial fragments of human chromosome #4 and expression of the G418 resistance gene

## (1) PCR analysis

[0132] Using the chimeric mice produced in Example 7, genomic DNAs were prepared from the tails of one individual derived from E14/#4-7 (K#4-7-1: about 5% chimerism) and one individual derived from E14/#4-4 (K#4-4-41: about 5% chimerism) by the same procedure as in Example 4. These DNAs were used as templates to conduct PCR analysis using polymorphic marker F11 for chromosome #4 analysis (see Example 6) which was detected in E14/#4 - 7 and E14/#4 - 4. As a result, expected amplification products were detected in both mice.

## (2) Southern analysis (Fig. 9)

[0133] Southern analysis was conducted in the same manner as in Example 2 by using human L1 sequence as a probe with 2  $\mu$ g of the genomic DNA derived from the tail of one individual derived from E14/#4-7 (K#4-7-1: about 5% chimerism). As a result, the presence of a large number of human L1 sequence was observed and their patterns were similar to those of E14/#4-7. The quantitative ratio to mouse genome was about 10% of that of E14/#4-7 at maximum. In Fig. 9, 2  $\mu$ g of genomic DNA digested with BglII was used in each lane. Human L1 sequence labeled with  $^{32}$ P was used as a probe and signals were detected with Image Analyzer BAS2000 (Fuji Photo Film Co., Ltd.). The molecular weights of DNAs are shown at the left side. The lanes represent the following as counted from the left: 1, K#4 - 7 - 1; 2, blank; and 3, E14/#4 - 7.

## (3) Test on the tail-derived fibroblast cells for G418 resistance

[0134] Fibroblast cells were prepared from the tails of one individual derived from E14/#4-7 (K#4-7-1: about 5% chimerism) and one individual derived from E14/#4-4 (K#4-4-41: about 5% chimerism). In the same procedure as in Example 4, the tail of each mouse was cut at a length of 5-10 mm and washed several times with PBS/1mM EDTA, followed by notching of the tail with a knife. The outer skin layer was removed and the inner tissues were cut into fine pieces. The fine pieces of tissues were transferred into a tube containing 5 ml of PBS/1 mM EDTA and left to stand for 30 minutes to 1 hour at room temperature. Subsequently, the supernatant was removed leaving a 1 ml portion of the PBS/EDTA behind, and 1 ml of 0.25% trypsin/PBS was added. The tissues were dispersed thoroughly by tapping or pipetting at room temperature for 5-10 minutes. After centrifugation at 1,000 rpm for 10 minutes, the precipitate was suspended in 2 ml of DMEM (10% FCS) and inoculated into a 35 mm plate. After cultivation for 7-10 days, the cells were treated with trypsin and about  $10^4$  cells per plate were inoculated into two 35 mm plates. G418 was added to the medium in one plate at a final concentration of 400  $\mu$ g/ml. The cells were cultured for 5-7 days and the appearance of viable cells in each plate were examined. Under these conditions, 100% of the wild-type ICR mouse-derived fibroblast cells were killed in the presence of G418. As a result, G418 resistant fibroblast cells were present in both mice.

[0135] These results show that E14/#4 - 7 and E14/#4 - 4 contributed to various normal tissues in the mouse and that they retained partial fragments of human chromosome #4.

## Example 9

## Transfer of human chromosome #14 or fragments thereof into mouse ES cells

[0136] The mouse A9 cell clone retaining human chromosome #14 (hereinafter referred to as "A9/#14") from Example 1 was used as a chromosome donor cell. Mouse ES cell line TT2 (purchased from Lifetech Oriental Co., Yagi et al., Analytical Biochem., 214:70, 1993) was used as a chromosome recipient cell. The TT2 cells were cultured in accordance with the method described in Aizawa Shinichi, "Biomaterial Series 8, Gene Targeting", published by Yodosha, 1995 and G418 resistant primary culture cells (purchased from Lifetech Oriental Co.) treated with mitomycin C (Sigma) were used as feeder cells. The microcell fusion and the selection of G418 resistant clones were conducted by the same procedures as in Example 2. The frequency of the appearance of the drug resistant clones was 3-6 per  $10^7$  of TT2 cells. The drug resistant clones were stored frozen and genomic DNA was prepared by the same procedures as in

## Example 2.

[0137] Human chromosome #14 was fragmented by irradiating the microcells with  $\gamma$ -rays (Koi et al., Science, 260: 361, 1993). The microcells obtained from about  $10^6$  cells of A9/#14 were suspended in 5 ml of DMEM and irradiated with  $\gamma$ -rays of 30 Gy on ice with a Gammacell 40 (Canadian Atomic Energy Public Corporation) at 1.2 Gy/min for 25 minutes. The fusion of  $\gamma$  ray-irradiated microcells and the selection of drug resistant clones were conducted by the same procedure as in the case of the unirradiated microcells. As a result, the frequency of the appearance of the drug resistant clones was 3 per  $10^7$  of TT2 cells. The drug resistant clones were frozen stored and DNA was prepared by the same procedure as in Example 2.

[0138] The retention of human chromosome #14 or partial fragments thereof in the unirradiated microcell-transferred G418 resistant clones 1-4 and 1-5, and in the G418 resistant clones 3-1 and 3-2 (a total of 4 clones) into which the  $\gamma$ -ray-irradiated microcell was transferred was confirmed by the methods described in (1) and (2) below.

## (1) PCR analysis (Fig. 10)

[0139] The presence of the gene on human chromosome #14 (O'Brien, Genetic Maps, 6th edition, Book 5, Cold



Spring Harbor Laboratory Press, 1993) and polymorphic markers (Polymorphic STS Primer Pairs: D14S43, D14S51, D14S62, D14S65, D14S66, D14S67, D14S72, D14S75, D14S78, D14S81 and PCI, BIOS Laboratories, Inc.; Nature 359:794, 1992; Nature Genetics, 7:22, 1994) was detected by a PCR method using genomic DNA of the drug resistant clone as a template. The sequences of oligonucleotide primers for the genes prepared on the basis of nucleotide sequences obtained from data bases such as GenBank, EMBL and the like are described below.

NP (nucleoside phosphorylase) :

5' - ATAGAGGGTACCCACTCTGG (SEQ ID NO:31) ,

5' - AACCAGGTAGGTTGATATGG (SEQ ID NO:32)

TCRA (T-cell receptor alpha) :

5' - AAGTTCCTGTGATGTCAAGC (SEQ ID NO:33) ,

5' - TCATGAGCAGATTAAACCCG (SEQ ID NO:34)

MYH6 (myosin heavy chain cardiac) :

5' - TGTGAAGGAGGACCAGGTGT (SEQ ID NO:35) ,

5' - TGTAGGGGTTGACAGTGACA (SEQ ID NO:36)

IGA2 (immunoglobulin alpha-2 constant) :

5' - CTGAGAGATGCCTCTGGTGC (SEQ ID NO:37) ,

5' - GGCGGTTAGTGGGGTCTTCA (SEQ ID NO:38)

IGG1 (immunoglobulin gamma-1 constant) :

5' - GGTGTCGTGGAACCTCAGGCG (SEQ ID NO:39) ,

5' - CTGGTGCAGGACGGTGAGGA (SEQ ID NO:40)

IGM (immunoglobulin mu constant) :

5' - GCATCCTGACCGTGTCCGAA (SEQ ID NO:41) ,

5' - GGGTCAGTAGCAGGTGCCAG (SEQ ID NO:42)

IGVH3 (immunoglobulin heavy variable-3) :

5' - AGTGAGATAAGCAGTGGATG (SEQ ID NO:43) ,

5' - GTTGTGCTACTCCCATCACT (SEQ ID NO:44)

[0140] PCR amplification was conducted using the genomic DNAs of the 4 drug resistant clones as templates with the above 18 kinds of the primers by the same procedure as in Example 2. As a result, expected amplification products were detected with all or part of the primers. In the drug resistant clones 3-1 and 3-2 obtained by using the  $\gamma$  - ray irradiated microcells, a tendency for the deletion of partial regions of chromosome #14 was observed. In the case where the unirradiated microcells were used, deletion was observed as in the case of the 1-4 clone. The results are shown in Fig. 10. In Fig. 10, a schematic chromosome map based on the G bands of human chromosome #14 and the location of some markers on bands are shown at the left side (see Example 2). The arrangement of the genetic and polymorphic markers shows approximate positional relationships on the basis of the presently available information (see Example 2) and the order is not necessarily correct. With respect to four kinds of the G418 resistant TT2 cell clones, the markers for which the expected amplification products were detected are shown by ■ and the markers for which the expected amplification products were not detected are shown by □. A9/#14 is a chromosome donor cell. The results of Example 11 (1) are shown at the right side.

## (2) Fluorescence in situ hybridization (FISH)

[0141] FISH analysis was conducted with probes specific to human chromosomes #14 (CHROMOSOME PAINTING SYSTEM, Cambio Ltd.) in accordance with the method described in Matsubara et al., "FISH Experiment Protocol", published by Shujunsha, 1994. As a result, in almost all of the observed metaphase spreads of all the 4 clones, human chromosome #14 or partial fragments thereof were detected in the form of an independent chromosome. The relative sizes of the observed human chromosome were consistent with those presumed from the results of the PCR analysis.

[0142] The results of the above experiments demonstrate that the obtained G418 resistant clones 1-4, 1-5, 3-1 and 3-2 retained the whole or partial fragments of human chromosome #14.

## Example 10

### Production of chimeric mice from the ES cells retaining human chromosome #14 or fragments thereof

[0143] The cells in the frozen stocks of four G418 resistant ES cell clones (1-4, 3-1, 3-2 and 1-5) that were prepared in Example 9 and which were confirmed to retain human chromosome #14 or fragments thereof were thawed, started to culture and injected into embryos at the eight-cell stage obtained by mating [ICR or MCH(ICR)] male and female mice (CREA JAPAN, INC.); the injection rate was 8-10 cells per embryo. The embryos were cultured in an ES medium overnight to develop to blastocysts. Two and half days after a foster mother ICR mouse (CREA JAPAN, INC.) was subjected to a pseudopregnant treatment, about ten of the injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 3.

Table 3.

Production of chimeric mice from the TT2 cell clones retaining human chromosome #14 (fragments)							
ES cell clone/ human chromosome	G418 resistant clone No.	Number of ES cell- injected 8-cell stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color		
					<20%	20-50%	50-80%
TT2/#14	1-4	98	20	1	-	-	1
	1-5	110	14	2	1	-	1
	3-1	103	11	2	1	1	-

Table 3. (continued)

Production of chimeric mice from the TT2 cell clones retaining human chromosome #14 (fragments)							
ES cell clone/ human chromosome	G418 resistant clone No.	Number of ES cell- injected 8-cell stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color		
					<20%	20-50%	50-80%
	3-2	183	19	3	-	2	1

[0144] As a result of the transplantation of a total of 494 injected embryos, 64 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo-derived albino coat color. Out of the 64 produced offsprings, 8 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells. The maximum contribution was about 80% in one individual derived from 1-4.

[0145] These results show that the G418 resistant ES cell clones (1-4, 1-5, 3-1 and 3-2) retaining human chromosome #14 or fragments thereof maintain the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse.

#### Example 11

Confirmation of retention of human chromosome #14 fragment DNA in the chimeric mice derived from the ES cells retaining human chromosome #14 fragments

[0146] The retention of human chromosome #14 partial fragments in the chimeric mice obtained in Example 10 was confirmed by the methods described in (1)-(3) below.

#### (1) PCR analysis using DNAs derived from various tissues

[0147] Genomic DNA was extracted from the tail of one individual derived from 3-1 (K3-1-1: about 25% chimerism) by the same procedure as in Example 4. The DNA was used as a template to conduct PCR analysis using all of the 14 primers for chromosome #14 analysis which were detected in 3-1. As a result, expected amplification products were detected with all the 14 primers (Fig. 10).

[0148] With respect to the same individual (K3-1-1), genomic DNA was obtained from the brain, kidney, spleen, heart, liver and thymus with a Puregene DNA Isolation Kit. For each tissue, PCR analysis was conducted with IGM primers (see Example 9). As a result, expected amplification products were detected in all the tissues (Fig. 11). The PCR products were electrophoresed on a 2% agarose gel and stained with ethidium bromide for detection. In Fig. 11, the lanes represent the following as counted from the left: B, brain; K, kidney; Sp, Spleen; H, heart; L, liver; Th, thymus; pc, human fibroblast cell (HFL-1) DNA (positive control); nc, non-chimeric mouse tail DNA (negative control); and M, marker (HindIII digested  $\lambda$  DNA + HaeIII digested  $\phi$  X174 DNA, TAKARA SHUZO CO., LTD.).

#### (2) Test on the tail-derived fibroblast cells for G418 resistance

[0149] Fibroblast cells were prepared from the tails of two individuals derived from 3-2 (K3-2-1: about 25% chimerism, and K3-2-3: about 50% chimerism) and one individual derived from 1-4 (K1-4-1: about 80% chimerism). In the same procedure as in Example 4, the tail of each chimeric mouse of 3-6 weeks was cut at a length of 5-10 mm and washed several times with PBS/1mM EDTA, followed by notching of the tail with a knife. The outer layer was removed and the inner tissues were cut into fine pieces. The fine pieces of tissues were transferred into a tube containing 5 ml of PBS/1 mM EDTA and left to stand for 30 minutes to 1 hour at room temperature. Subsequently, the supernatant was removed leaving a 1 ml portion of the PBS/EDTA behind, and 1 ml of 0.25% trypsin/PBS was added. The tissues were dispersed thoroughly by tapping or pipetting at room temperature for 5-10 minutes. After centrifugation at 1,000 rpm for 10 minutes, the precipitate was suspended in 2 ml of DMEM (10% FCS) and inoculated into a 35 mm plate. After cultivation for 7-10 days, the cells were treated with trypsin and about 104 cells per plate were inoculated into four 35 mm plates. G418 was added to the medium in two of the plates at a final concentration of 400  $\mu$ g/ml. The cells were cultured for 5-7 days and the viable cells in each plate were counted. Under these conditions, 100% of the wild-type ICR mouse-derived fibroblast cells were killed in the presence of G418. Assuming the same growth rate of the G418 resistant fibroblast in the non-selective and selective media, the ratio of the viable cells in the selective medium to those in the

non-selective medium is believed to reflect the contribution in the fibroblast cell populations of the G418 resistant ES cell-driven fibroblast. As a result, the presence of G418 resistant fibroblast cells was observed in all the three individuals as shown in Fig. 12. In Fig. 12, % resistance is an average of 2 pairs of the selective/non-selective 35 mm plates for each mouse. ICR refers to the wild-type ICR mice.

### (3) FISH analysis of the tail-derived G418 resistant fibroblast cells

[0150] FISH analysis of the K3-2-3 and K1 - 4 - 1 derived G418 resistant fibroblast cells obtained in (2) was conducted by the same procedure as in Example 2. Total human DNA extracted from the HFL-1 cells (Example 1) was labeled with FITC so that it could be used as a probe (Matsubara et al., "FISH Experimental Protocol", published by Shujunsha, 1994). As a result, in almost all of the observed metaphase spreads of the both individuals, partial fragments of the human chromosome in independent forms were observed.

[0151] These results show that the TT2 cell clones retaining fragments of human chromosome #14 contributed to various normal tissues in the mouse individuals and that they retained partial fragments of human chromosome #14.

### Example 12

#### Transfer of partial fragments of human chromosome #2 into ES cells

[0152] The mouse A9 cell W23 retaining a human chromosome #2 fragment (hereinafter referred to as "A9/#2 W23") from Example 1 was used as a chromosome donor cell. Mouse ES cell line TT2 (see Example 9) was used as a chromosome recipient cell. The microcell fusion and the selection of G418 resistant clones were conducted by the same procedures as in Example 2. The frequency of the appearance of the drug resistant clones was 1-3 per 10<sup>7</sup> of TT2 cells. The drug resistant clones were stored frozen and genomic DNA was prepared by the same procedures as in Example 2. The retention of partial fragments of human chromosome #2 in drug resistant clones 5-1, 5-2 and 5-3 was confirmed by the methods described in (1) and (2) below.

#### (1) PCR analysis

[0153] The presence of C  $\kappa$  and FABP1 that are the genes on human chromosome #2 (Genetic Maps, supra) and which were detected in the chromosome donor cell A9/#2 W23 was detected by a PCR method.

[0154] As a result of PCR amplification using each primer, expected amplification products were detected with both primers in all of the 3 clones.

#### (2) Fluorescence in situ hybridization (FISH)

[0155] FISH analysis was conducted with probes specific to human chromosome #2 (CHROMOSOME PAINTING SYSTEM, Cambio Ltd.) by the same method as in Example 2. As a result, in almost all of the observed metaphase spreads of the 3 clones, partial fragments of human chromosome #2 in the form of independent chromosomes were detected. The sizes of the observed human chromosome were the same as those observed in A9/#2 W23.

[0156] The results of the above experiments demonstrate that the obtained G418 resistant clones retained partial fragments of human chromosome #2.

### Example 13

#### Production of chimeric mice from the ES cells retaining human chromosome #2

[0157] The cells in a frozen stock of the G418 resistant ES cell clone 5-1 that was obtained in Example 12 and which was confirmed to retain human chromosome #2 was thawed, started to culture and injected into 8-cell stage embryos obtained by mating ICR or MCH(ICR) male and female mice (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. The embryos were cultured in an ES medium (Example 9) overnight to develop to blastocysts. Two and half days after a foster mother ICR mouse (CREA JAPAN, INC.) was subjected to a pseudopregnant treatment, about ten of the injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 4.

Table 4.

Production of chimeric mice from the TT2 cell clone retaining human chromosome #2 (fragments)							
ES cell clone/ human chromosome	G418 resistant clone No.	Number of ES cell- injected 8 cell stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color		
					<20%	20-50%	50-80%
TT2/#2 (W23)	5-1	264	51	18	7	5	6

[0158] As a result of the transplantation of a total of 264 injected embryos, 51 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo-derived albino coat color. Out of the 51 produced offsprings, 18 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells. The maximum contribution was about 80%.

[0159] These results show that the G418 resistant ES cell clone (5-1) retaining a fragment of human chromosome #2 maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse individual.

#### Example 14

Detection of human antibody heavy chain in sera of the human chromosome #14 transferred chimeric mice

[0160] The concentrations of human antibody in the sera were determined by enzyme-linked immunosorbent assay (ELISA). The ELISA for human antibody was performed in accordance with the method described in Toyama and Ando, "Monoclonal Antibody Experiment Manual", published by Kodansha, 1987; Andou and Chiba, "Monoclonal Antibody Experiment Procedure Manual", published by Kodansha Scientific, 1991; Ishikawa, "Super High Sensitivity Enzyme Immuno Assay", published by Gakkai-syuppan center, 1993; Ed Harlow and David Lane, "Antibodies A Laboratory Manual", published by Cold Spring Harbor Laboratory, 1988 and A. Doyle and J. B. Griffiths, "Cell & Tissue Culture: Laboratory Procedures", published by John Wiley & Sons Ltd., 1996. In some assays, the condition of reaction were modified, for example, the reaction was performed at 4°C over night. Antibodies to human-immunoglobulin or antigen were diluted to about 0.5-10 µg/ml (100-5000 fold) and ELISA plates were coated with these solutions. PBS supplemented with 5% mouse serum (Sigma, M5905) was used for blocking and dilution of the samples and labeled antibodies. PBS was used for 20-fold dilution of the chimeric mouse sera. After washed, the coated plate was blocked over 1 hour. After plate was washed, sample was added and incubated over a half hour. After washed, Enzyme labeled anti-human immunoglobulin antibodies diluted 100-5000 folds were added to the plates and incubated over 1 hour, the plate was washed and then substrate was added. In some assays, the same procedure was applied except that a biotin-labeled antibody was used. After plate was washed, avidin-enzyme complex was added. After plate was washed, substrate was added. Absorbances were measured with a microplate reader (Bio-tek instrument, EL312e). The chimeric mice (Example 10, K3-1-2, K3-2-2 and K3-2-3) which were 29-35 days old were bled and assayed by ELISA. Anti-human IgM mouse monoclonal antibody (Sigma, I6385) was diluted with 50 mM carbonate - bicarbonate buffer (pH 9.6) and absorbed to the 96-well microtiter plates. The serum samples diluted with mouse serum (Sigma, M5905) supplemented PBS were added to the plates. Subsequently, peroxidase-labeled anti-human IgM goat antibody (Tago, 2392) was added and the plates were incubated. After ABTS substrate (Kirkegaard & Perry Laboratories Inc., 506200) was added, enzyme activity was determined by absorbance measurement at 405 nm. Purified human IgM antibody (CAPEL, 6001-1590) and IgG (Sigma, I4506) were used as standards. The standards were diluted stepwise with mouse serum-supplemented PBS. In the determination of human IgG concentration, anti-human IgG goat antibody (Sigma, I3382) was absorbed to the plate and the human IgG was detected with peroxidase-labeled anti-human IgG goat antibody (Sigma, A0170). The results are shown in Table 5. Both human IgM and IgG were detected.

Table 5.

Concentrations of Human Antibodies in Chimeric Mouse Sera (ELISA)		
Chimeric Mouse (mg/l)	IgG (mg/l)	IgM
K3-1-2	0.37	3.7
K3-2-2	0.33	5.9

Table 5. (continued)

Concentrations of Human Antibodies in Chimeric Mouse Sera (ELISA)		
Chimeric Mouse (mg/l)	IgG (mg/l)	IgM
K3-2-3	0.51	3.4

[0161] Two milliliters of human serum albumin (HSA, Sigma, A3782) dissolved in PBS was mixed with adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc.) to prepare an antigen solution at a concentration of 0.25 mg/ml. The chimeric mice retaining human chromosome #14 fragment (Example 10, K3-1-1 and K3-2-1) were immunized with 0.2 ml of the antigen solution 3 times at days 27, 34 and 41 after birth. The chimeric mouse sera were assayed by ELISA. The results are shown in Figs. 13 and 14. The human antibody concentration in the sera of the HSA-immunized chimeric mice was increased after the immunization. In the K3 - 1 - 1 mouse, 18  $\mu$ g/ml of human IgM and 2.6  $\mu$ g/ml of IgG were detected in the serum at day 17 after the immunization. In the serum of the control ICR mouse, the human antibody titer was not significant.

#### Example 15

Production of human antibody heavy chain-producing hybridomas from the human chromosome #14 transferred chimeric mouse

[0162] The spleen was removed from the human albumin-immunized chimeric mouse (K3-1-1, Example 14) at day 44 after birth. The spleen cell was fused with a myeloma cell to produce a hybridoma. The hybridoma was produced using a myeloma cell P3X63Ag8.653 (DAINIPPON PHARMACEUTICAL CO., LTD., 05-565) by the method described in Ando and Chiba, "Monoclonal Antibody Experimental Procedure Manual", published by Kodansha Scientific, 1991. The hybridomas were inoculated into ten 96-well plates and cultured for 1 week. The culture supernatant was analyzed by ELISA. The ELISA procedure was conducted by using anti-human IgM mouse monoclonal antibody (Sigma, I6385) immobilized on ELISA plate in the same manner as in Example 14 to give 6 positive clones. HSA (antigen) was dissolved in 50 mM carbonate-bicarbonate buffer (pH 9.6) at a concentration of 5  $\mu$ g/ml and the antigen solution was dispensed in 100  $\mu$ l portions into all the wells of the ELISA plates. After the addition of the supernatant, peroxidase-labeled anti-human IgA + IgG + IgM goat antibodies (Kierkegaard & Perry Laboratories Inc., 04-10-17) were used for detection of HSA-specific human antibody. One positive clone was confirmed in the ten plates. This clone was one of the 6 human IgM positive clones. The clone (H4B7) was further cultured and the culture supernatant was diluted, followed by ELISA analysis using HSA as an antigen with peroxidase-labeled anti-human IgM goat antibody (Tago, 2392) in the same manner as described above. As a result, the absorbance decreased with the increase in the dilution of the culture solution. Serial twofold dilutions of 2  $\mu$ g/ml human IgM (CAPEL, 6001-1590) showed low absorbance regardless of dilution ratios. This suggests that the antibody produced by hybridoma H4B7 had a specificity to HSA (Fig. 15). In Fig. 15, the dilution of the culture supernatant samples is plotted on the horizontal axis and the absorbance at 405 nm is plotted on the vertical axis.

#### Example 16

Re-marking of the G418 resistance-marked human chromosome #2 fragment with puromycin resistance

[0163] The A9 cells retaining the G418 resistance-marked human chromosome #2 fragment (W23) (see Example 1, Fig. 1) were cultured in a G418 (800  $\mu$ g/ml) containing selective medium (10% FBS, DMEM) in a 100 mm plate. Plasmid pPGKPuro (provided by Dr. Peter W. Laird (WHITEHEAD INSTITUTE)) containing puromycin resistance gene was linearized with restriction enzyme Sall (TAKARA SHUZO CO., LTD.) before transfection. The cells were treated with trypsin and suspended in Dulbecco's phosphate buffered saline (PBS) at a concentration of  $5 \times 10^6$  cells/ml, followed by electroporation using a Gene Pulser (Bio-Rad Laboratories, Inc.) in the presence of 10  $\mu$ g of DNA in the same manner as in Example 1. A voltage of 1000 V was applied at a capacitance of 25  $\mu$ F with an Electroporation Cell of 4 mm in length (Example 1) at room temperature. The electroporated cells were inoculated into media in 3-6 plates of 100 mm $\phi$ . After one day, the medium was replaced with a double-selective medium containing 10  $\mu$ g/ml of puromycin (Sigma, P-7255) and 800  $\mu$ g/ml of G418. The colonies formed after 2-3 weeks were collected in groups each consisting of about 200 colonies. The cells of each of the three groups were cultured in two or three 25 cm<sup>2</sup> flasks to form microcells. The mouse A9 cells were cultured in a 25 cm<sup>2</sup> flask and fused with the microcells by the same procedure as in Example 1. The fused cells were transferred into two 100 mm plates and cultured in the double-selective medium containing G418 and puromycin. One of the three groups gave two double-drug resistant clones. In these clones, it

was most likely that puromycin resistance marker had been introduced into human chromosome #2 fragment.

#### Example 17

5 Duplication of transferred human chromosome in the human chromosome transferred ES cells

[0164] The ES cell clone retaining the G418 resistance marked human chromosome #14 fragment (E14/#14 - 36) was cultured in a medium containing G418 at a high concentration to give ES cell clones in which the human chromosome was duplicated ( "Biomanual Series 8, Gene Targeting", published by Yodosha, 1995). G418 resistant mouse primary cells (purchased from Lifetech Oriental) were inoculated into a 100 mm plate without treating with mitomycin C and used as feeder cells. The E14/#14-36 cells were inoculated into the 100 mm plate and after half a day, the medium was replaced with a medium containing G418 at a concentration of 16 mg/ml. The medium was replaced every 1-2 days. The G418 concentration was changed to 10 mg/ml one week later and the cultivation was continued. Among the colonies formed, 15 were picked up and cultured, followed by FISH analysis of chromosome using human chromosome #14 specific probes (see Example 9). As a result, human chromosome #14 fragment was found to have duplicated in the 8 clones.

#### Example 18

20 Preparation of Mouse ES cells retaining both human chromosome #2 partial fragments and human chromosome #14 partial fragments.

[0165] In a microcell transfer experiment using the double-drug resistant clone PG-1 from Example 16 as a microcell donor cell and a wild-type A9 cell as a recipient cell, it was confirmed that the human chromosome #2 partial fragment retained in PG - 1 was marked with a puromycin resistance gene. The preparation of microcells and the fusion with the A9 cells was carried out by the same methods as in Example 1. As a result, 10 days after the microcell fusion, a total of fifty nine G418 resistant colonies appeared. After the medium for these colonies was changed to one containing 8 µg/ml puromycin, the colonies were cultured for 3 days to give 45 viable colonies (76%). In many cases of microcell fusion, only one or few chromosomes are transferred into a recipient cell. Hence, cotransfer of both the resistance genes at a high frequency shows that the G418 resistance-labeled chromosome #2 partial fragment retained in the PG1 clone was also marked with the puromycin resistance gene. In addition, for the detection of the respective marker genes on the human chromosome #2 partial fragment, FISH analysis was conducted by using pSTneoB (see Example 1) as a probe in the case of the A9/#2 W23 clone having only G418 resistance (see Example 16) and by using pPGKPuro (see Example 16) as a probe in the case of the PG1 clone in accordance with the method described in Matsubara et al., "FISH Experiment Protocol", published by Shujunsha, 1994. As a result, in the case of the A9/#2 W23 clone, one signal was observed in each of the sister chromatids of the human chromosome #2 partial fragment observed in Example 12 (2 signals in total). This indicated the insertion of pSTneoB into the human chromosome #2 partial fragment at one site. In the case of the PG1 clone, a total of 4 signals were observed on a chromosome fragment of the same size as in A9/#2 W23. Since pSTneoB and pPGKPuro had identical sequences in their vector portions, the pSTneoB could be detected by the pPGKPuro probe. Hence, it is believed that out of the four signals observed in the PG1 clone, two were from the pSTneoB and the other two were from the pPGKPuro. These results show that the human chromosome #2 partial fragment retained in the PG1 was marked with both the G418 and puromycin resistances.

[0166] The PG1 cell clone was used as a chromosome donor cell to prepare a mouse ES cell retaining both a human chromosome #2 partial fragment and a human chromosome #14 partial fragment. The G418 resistant TT2 cell clone 1-4 already retaining the human chromosome #14 partial fragment (see Example 9) was used as a chromosome recipient cell. The microcell fusion and the selection of puromycin resistant cells were carried out by the same methods as in the selection of the G418 resistant clones in Example 9 except that the concentration of puromycin was 0.75 µg/ml. The frequency of the appearance of the resulting puromycin resistant clones was 3-7 per 10<sup>7</sup> of 1-4 cells. The presence of G418 resistance in these puromycin resistant clones was confirmed from the fact that they were grown in the presence of 300 µg/ml of G418. The double-drug resistant clones were stored frozen and genomic DNA was prepared by the same methods as in Example 2. The retention of the human chromosome #2 partial fragment and human chromosome #14 partial fragment was confirmed by the method described in (1) in the case of double-drug resistant clones PG5, PG15 and PG16 and by the method described in (2) in the case of the clone PG15.

55 (1) PCR analysis

[0167] Genomic DNAs of the double-drug resistant clones were used as templates in the PCR amplifications. Among the markers on human chromosomes #2 and #14 (Genetic Maps, supra), the primers whose presence in the A9/#2

W23 clone was confirmed in Example 12 and those whose presence in the TT2/#14 1-4 clone was confirmed in Example 9 were used. All the primers gave expected amplification products in all the three clones.

## (2) Fluorescence in situ hybridization (FISH)

[0168] FISH analysis was conducted by using FITC-labeled human total DNA as a probe in the same manner as in Example 11. As a result, in almost all of the metaphase spreads, two (large and small) human chromosome fragments were detected. The large fragment had the same size as that of the partial fragment detected by using the human chromosome #14 specific probes in the case of the TT2/#14 1-4 clone in Example 9 and the small fragment had the same size as that of the partial fragment detected by using the human chromosome #2 specific probes in the case of the TT2/#2 5-1 in Example 12. The results are shown in Fig. 16. In Fig. 16, the less bright chromosome was derived from the mouse. The two (large and small) chromosome fragments of high brightness due to FITC fluorescence as shown by arrows were derived from the human, which are believed to correspond to the human chromosome #14 and #2 partial fragments.

[0169] These results show that the obtained double-drug resistant ES clones retained both the human chromosome #2 partial fragment and the human chromosome #14 partial fragment.

## Example 19

Production of chimeric mice from the mouse ES cell clones retaining both human chromosome #2 partial fragments and human chromosome #14 partial fragments.

[0170] The cells in frozen stocks of the G418 and puromycin double-resistant TT2 cell clones PG5, PG15 and PG16 from Example 18 which were confirmed to retain human chromosome #2 partial fragments and human chromosome #14 partial fragments were thawed, started to culture and injected into 8-cell stage embryos obtained by mating ICR or MCH(ICR) male and female mice (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. The embryos were cultured in a medium for ES cells (see Example 9) overnight to develop to blastocysts. Two and a half day after a foster mother ICR mouse was subjected to a pseudopregnant treatment, about ten of the injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 6.

Table 6.

Production of chimeric mice from the mouse ES cell clones retaining both human chromosome #2 partial fragments and human chromosome #14 partial fragments							
ES cell clone/ human chromosome	Double-drug resistant clone No.	Number of ES cell- injected 8-cell stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color		
					< 10%	10 - 50%	50%<
TT2/ #14+#2	PG5	160	26	8	7	1	-
	PG15	168	15	3	1	2	-
	PG16	223	32	12	3	6	3

[0171] As a result of the transplantation of a total of 551 injected embryos, 73 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo-derived albino coat color. Out of the 73 produced offsprings, 23 mice were recognized to have a partial agouti coat color, indicating the contribution of the ES cells.

[0172] These results show that the ES cell clones PG5, PG15 and PG16 retaining human chromosome #2 partial fragments and human chromosome #14 partial fragments maintain the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse.



## Example 20

Detection of human antibody in sera of the chimeric mice derived from the ES cells retaining both human chromosome #2 partial fragments and human chromosome #14 partial fragments

[0173] The two KPG-15 (9 weeks old; derived from the PG-5 clone, 10% chimerism) and KPG-18 (5 weeks old; derived from the PG-5 clone, 10% chimerism) chimeric mice from Example 19 were immunized with 0.2 ml of a solution of human serum albumin (HSA, Sigma, A3782) and adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc.) at a HSA concentration of 0.25 mg/ml. The chimeric mice were bled just before the immunization and 8 days after that and the concentrations of human antibody  $\mu$  and  $\kappa$  chains in the sera were determined by ELISA (see Example 14). Ninety six-well microtiter plates were coated with anti-human antibody  $\kappa$  chain goat antibody (VECTOR LABORATORIES INC., AI-3060) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) and then a serum sample diluted with mouse serum (Sigma, M5905) - containing PBS was added. Subsequently, biotin-labeled anti-human antibody  $\kappa$  chain goat antibody (VECTOR LABORATORIES INC., BA-3060) was added to the plates and incubated. A complex of biotinylated horseradish peroxidase and avidin DH (VECTOR LABORATORIES, INC., Vectastain ABC Kit, PK4000) was added and incubated. After 3,3',5,5'-tetramethylbenzidine (TMBZ, Sumitomo Bakelite, ML-1120T) was added as a peroxidase substrate, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgG antibody having  $\kappa$  chain (Sigma, I-3889) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS. In the case of  $\mu$  chain, 96-well microtiter plates were coated with anti-human antibody  $\mu$  chain mouse monoclonal antibody (Sigma, I-6385) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) and then a serum sample was added. Subsequently, peroxidase-labeled anti-human antibody  $\mu$  chain mouse antibody (The Binding Site Limited, MP008) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgM antibody having  $\mu$  chain (CAPPEL, 6001-1590) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS. As a result, both the human antibody  $\mu$  and  $\kappa$  chains were detected in both individuals. The concentrations of these human antibodies in the sera increased after the immunization (Tables 7 and 8).

Table 7.

Concentrations of Human Antibodies in Chimeric Mouse KPG15 (ELISA)		
	IgM (mg/l)	Ig $\kappa$ (mg/l)
Before Immunization	0.19	1.6
8 Days After Immunization	0.75	1.7

Table 8.

Concentrations of Human Antibodies in Chimeric Mouse KPG18 (ELISA)		
	IgM (mg/l)	Ig $\kappa$ (mg/l)
Before Immunization	0.29	0.57
8 Days After Immunization	3.4	0.87

[0174] These results show that human antibody heavy and light chain genes can function in the chimeric mice derived from the ES cells retaining both human chromosome #2 partial fragments and human chromosome #14 partial fragments.

## Example 21

Detection of anti-HSA human antibody  $\gamma$  chain in sera of the human chromosome #14 fragments transferred chimeric mice

[0175] The chimeric mice retaining human chromosome #14 fragments which were produced by the same method as in Example 10 (K9 and K11: both were derived from the TT2 cell clone 3-2, with chimerisms of 50% and 30%, respectively) were immunized with HSA either 4 times at days 79, 93, 107 and 133 after birth (K9) or 3 times at days 74, 88 and 111 after birth (K11) by the same method as in Example 20. Antibodies including human  $\gamma$  chain against human serum albumin in the sera of the chimeric mice were detected by ELISA. Ninety six-well microtiter plates were coated with HSA (Sigma, A 3782) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) and then a sample diluted

with PBS was added. Subsequently, peroxidase-labeled anti-human IgG mouse antibody (Pharmingen, 08007E) was added to the plates and incubated. After O-phenylenediamine (OPD, Sumitomo Bakelite, ML-11300) was added as a peroxidase substrate, enzyme activity was determined by absorbance measurement at 490 nm. The titer of the anti-HSA human IgG in the sera of the chimeric mice immunized with HSA increased after the immunization. On the other hand, control ICR mouse gave a background level of the anti-HSA human IgG titer after the immunization with HSA. The results are shown in Fig. 17. In Fig. 17, the number of days after the first immunization of the chimeric mice with HSA is plotted on the horizontal axis and the absorbance at 490 nm is plotted on the vertical axis. These results show that the antibody titer of the antigen specific human IgG was increased by stimulation with the HSA antigen in the chimeric mice retaining human chromosome #14 fragments.

#### Example 22

Detection of human antibody  $\lambda$  chain in a serum of the human chromosome #22 fragment transferred chimeric mouse

[0176] The chimeric mouse K22-7 from Example 3 (9 weeks old; 10% chimerism) was bled and human antibody  $\lambda$  chain in the serum was detected by ELISA (see Example 14). Ninety six-well microtiter plates were coated with anti-human antibody  $\lambda$  chain goat antibody (VECTOR LABORATORIES INC., AI-3070) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) and then a serum sample was added. Subsequently, biotin-labeled anti-human antibody  $\lambda$  chain goat antibody (VECTOR LABORATORIES INC., BA-3070) was added to the plates and incubated. A complex of biotinylated horseradish peroxidase and avidin DH (VECTOR LABORATORIES, INC., Vectastain ABC Kit) was added and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added as a peroxidase substrate, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgG antibody having  $\lambda$  chain (Sigma, I-4014) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS. As a result, human antibody  $\lambda$  chain was detected in the chimeric mouse at a concentration corresponding to 180 ng/ml of human IgG. These results show that human antibody  $\lambda$  chain gene can function in the chimeric mouse retaining a human chromosome #22 fragment.

#### Example 23

Detection of human antibody  $\kappa$  chain in sera of the human chromosome #2 fragment transferred chimeric mice

[0177] The chimeric mouse K2-8 from Example 13 (5 weeks old; 70% chimerism) and the chimeric mice K2-3, K2-4 and K2-12 from Example 13 (9 weeks old; chimerisms was 50%, 20% and 80%, respectively) were bled and human antibody  $\kappa$  chain in the sera was detected by ELISA (see Example 14). Ninety six-well microtiter plates were coated with anti-human antibody  $\kappa$  chain goat antibody (VECTOR LABORATORIES INC., AI - 3060) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) and then a serum sample was added. Subsequently, biotin-labeled anti-human antibody  $\kappa$  chain goat antibody (VECTOR LABORATORIES INC., BA-3060) was added to the plates and incubated. A complex of biotinylated horseradish peroxidase and avidin DH (VECTOR LABORATORIES, INC., Vectastain ABC Kit) was added and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgG antibody having  $\kappa$  chain (Sigma, I-3889) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS. The results are shown in Table 9.

Table 9.

Concentration of Human Antibody $\kappa$ Chain in Chimeric Mouse (ELISA)	
Chimeric Mouse	Ig $\kappa$ (mg/l)
K2 - 3	124
K2 - 4	85
K2 - 8	25
K2 - 12	56

[0178] The chimeric mice K2-3 and K2-4 retaining human chromosome #2 fragments from Example 13 were immunized with HSA, 3 times at days 66, 80 and 102 after birth by the same method as in Example 20. The chimeric mouse K2-12 was immunized with HSA, 4 times at days 63, 77, 91 and 116 after birth by the same method as in Example 20. Human antibody  $\kappa$  chain against HSA in the sera of the chimeric mice was detected by ELISA (see Example 14). Ninety six-well microtiter plates were coated with HSA (Sigma, A 3782) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) and then a sample was added. Subsequently, biotin-labeled anti-human antibody  $\kappa$  chain goat antibody (VECTOR

LABORATORIES, INC., BA - 3060) was added to the plates and incubated. A complex of biotinylated horseradish peroxidase and avidin DH (VECTOR LABORATORIES, INC., Vectastain ABC Kit) was added and incubated. After OPD (Sumitomo Bakelite, ML-11300) was added as a peroxidase substrate, enzyme activity was determined by absorbance measurement at 490 nm. The titer of the anti-HSA human  $\kappa$  chain in the sera of the chimeric mice immunized with HSA increased after the immunization. On the other hand, control ICR mouse gave a background level of the anti-HSA human  $\kappa$  chain titer after the immunization. The results are shown in Fig. 18. In Fig. 18, the number of days after the first immunization of the chimeric mice with HSA is plotted on the horizontal axis and the absorbance at 490 nm is plotted on the vertical axis. These results show that human antibody  $\kappa$  chain gene can function in the chimeric mice retaining human chromosome #2 fragments and that the antibody titer of the antigen specific human Ig  $\kappa$  was increased by stimulation with the HSA antigen in the chimeric mice.

#### Example 24

Preparation of human antibody heavy chain ( $\mu$  chain or  $\gamma$  chain)-producing hybridomas from the human chromosome #14 transferred chimeric mouse

[0179] The spleen was removed from the HSA-immunized chimeric mouse K9 (see Example 21) at day 136 after birth. A spleen cell was fused with a myeloma cell to produce a hybridoma. The hybridoma was produced using a myeloma cell Sp-2/0-Ag14 (Dainippon Pharmaceutical Co., Ltd., 05-554) by the method described in Toyama and Ando, "Monoclonal Antibody Experiment Procedure Manual", published by Kodansha Scientific, 1991. The cells were inoculated into a medium containing 10% ORIGEN Hybridoma Cloning Factor (HCF, Bokusui Brown) in eight 96-well plates and G418 was added after 3 days at a concentration of 1 mg/ml, followed by cultivation for 1-3 weeks. The culture supernatant was analyzed by ELISA. Ninety six-well microtiter plates were coated with anti-human  $\mu$  chain mouse monoclonal antibody (Sigma, I-6385) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) and a sample diluted with PBS was added. Subsequently, peroxidase-labeled anti-human  $\mu$  chain mouse antibody (The Binding Site LIMITED, MP008) was added to the plates and incubated. 2,2'-Azino-di- (3-ethyl-benzothiazoline-6-sulfonate) diammonium salt (ABTS, Kirkegaard & Perry Laboratories Inc., 04-10-17) was used as a substrate to detect seven positive clones. In the detection of  $\gamma$  chain-producing clones, 96-well microtiter plates were coated with anti-human  $\gamma$  chain mouse monoclonal antibody (Sigma, I-6260) and a sample diluted with PBS was added. Subsequently, peroxidase-labeled anti-human  $\gamma$  chain mouse antibody (Pharmingen, 08007E) was added to the plates and incubated. ABTS (Kirkegaard & Perry Laboratories Inc., 04-10-17) was used as a substrate and two human antibody  $\gamma$  chain-positive clones were obtained.

#### Example 25

Preparation of human antibody light chain-producing hybridomas from the human chromosome #2 transferred chimeric mouse

[0180] The spleen was removed from the HSA-immunized chimeric mouse K2-3 (see Example 23) at day 105 after birth. A spleen cell was fused with a myeloma cell to produce a hybridoma. The hybridoma was produced using a myeloma cell P3X63Ag8.653 (Dainippon Pharmaceutical Co., Ltd., 05-565) by the method described in Toyama and Ando, "Monoclonal Antibody Experiment Procedure Manual", published by Kodansha Scientific, 1991. The cells were inoculated into a medium containing 10% HCF (Bokusui Brown) in ten 96-well plates and G418 was added after 3 days at a concentration of 1 mg/ml, followed by cultivation for 1-3 weeks. The culture supernatant was assayed by ELISA. The ELISA analysis was conducted by the same method as in Example 23 and two human antibody  $\kappa$  chain-positive clones were obtained.

#### Example 26

Re-marking of the G418 resistance-marked human chromosome #22 with puromycin resistance

[0181] The A9 cells retaining the G418 resistance-marked human chromosome #22 (A9/#22  $\gamma$  2) from Example 1 were re-marked with puromycin resistance by the same method as in Example 16. About 200 colonies of double-drug resistant clones obtained by electroporation of the  $\gamma$  2 cells with pPGKPuro were collected as one group and three such groups (P1, P2 and P3) were used as donor cells to perform microcell transfer into wild-type mouse A9 cells. As a result, 6, 1 and 3 of double-drug resistant clones were obtained from the groups P1, P2 and P3, respectively. The clone 6-1 from group P3 was used as a microcell donor cell and a wild-type A9 cell as a recipient cell to perform a microcell transfer experiment (see Example 18). As a result, the human chromosome #22 was confirmed to have been

further marked with a puromycin resistance gene. The preparation of microcells and the fusion with A9 cells were conducted by the same methods as in Example 1. As a result, twenty eight G418 resistant colonies appeared 11 days after the microcell transfer. After the medium for these colonies was changed to one containing 8  $\mu$ g/ml puromycin, these colonies were cultured for 3 days to give 21 (75%) viable colonies. In many cases of microcell fusion, only one or few chromosomes are transferred into a recipient cell. Hence, cotransfer of both the resistance genes at a high frequency shows that the G418 resistance-labeled chromosome #22 retained in the 6-1 clone was marked with the puromycin resistance gene.

#### Example 27

Preparation and Sequencing of cDNA of a human antibody heavy chain variable region from the human antibody heavy chain-producing hybridoma

[0182] Among the human antibody heavy chain (IgM)-producing hybridomas obtained in Example 15, H4B7 (HSA-specific) and H8F9 (non-specific) hybridomas were selected. Total RNAs were obtained from these hybridomas using ISOGEN (Nippon Gene). The synthesis of cDNA from 5  $\mu$ g each of the total RNAs was conducted with a Ready-To-Go T-primed 1st strand Kit (Pharmacia Co.). Using the resulting cDNA and the following primers prepared with reference to Larrick et al., BIO/TECHNOLOGY, 7, 934-, 1989; Word et al., Int. Immunol., 1, 296-, 1989, PCR was performed to amplify a human antibody heavy chain variable region.

CM1 (human IgM constant region) : 5' -

TTGTATTTCCAGGAGAAAGTG (SEQ ID NO:45)

CM2 (ditto) : 5' - GGAGACGAGGGGGAAAAGGG (SEQ ID NO:46)

HS1 (human heavy chain variable region) : 5' -

ATGGACTGGACCTGGAGG(AG)TC(CT)TCT(GT)C (SEQ ID NO:47) (a mixture of 8 sequences)

HS2 (ditto) : 5' -

ATGGAG(CT)TTGGGCTGA(GC)CTGG(GC)TTT(CT)T (SEQ ID NO:48)

(a mixture of 16 sequences)

HS3 (ditto) : 5' -

ATG(AG)A(AC)(AC)(AT)ACT(GT)TG(GT)(AT)(GCT)C(AT)(CT)(GC)  
)CT(CT)CTG (SEQ ID NO:49) (a mixture of 6144 sequences)

\* ( ) means that any one of the bases therein should be selected.

[0183] In both cases of the H4B7 and H8F9 hybridomas, the first run of PCR was performed by using three kinds of primer combinations of HS1xCM1, HS2xCM1 and HS3xCM1 in 40 cycles at 94 °C for 1 minute, 50 °C for 2 minutes and 72 °C for 3 minutes with a Thermal Cycler 140 (Perkin-Elmer Corp.). The PCR products were amplified again under the same temperature conditions in 30 cycles using HS1xCM2, HS2xCM2 and HS3xCM2 primers, respectively.

The amplification products were electrophoresed on a 1.5% agarose gel and detected by staining with ethidium bromide. As a result, an amplification product of about 490 bp was detected with the HS3xCM2 primer in the case of the H4B7 hybridoma. In the case of the H8F9 hybridoma, a slight band was detected at the same site with the HS3xCM2 primer. The band in the case of H8F9 was amplified again with the HS3xCM2 primer in 30 cycles under the same temperature conditions as above. As a result, the amplification product was detected as a very intensive signal. These PCR products were cloned into a pBlueScriptII SK+ (Stratagene Ltd.) at a SmaI site in accordance with the method described in Ishida et al., "Gene Expression Experiment Manual", published by Kodansha Scientific, 1995. Among the amplification product-inserted plasmids, plasmids #2, #3, #4(H4B7), #11, #13 and #14 (H8F9) were selected and the nucleotide sequences of the amplification products were determined with a Fluorescence Autosequencer (Applied Biosystems Inc.). As a result of the comparison of the obtained nucleotide sequences or deduced amino acid sequences with those of known human antibody VH region (Marks et al., Eur. J. Immunol. 21, 985-, 1991) and JH region (Ravetch et al., Cell, 27, 583-, 1981), it was revealed that both the H4B7 and H8F9 hybridomas contained a combination of genes for VH4 family and JH2. These results show that the chimeric mouse retaining human chromosome #14 partial fragment produced a complete functional human antibody heavy chain protein.

#### Example 28

Preparation and Sequencing of cDNA of human antibody  $\kappa$  chain from the spleen of the human antibody  $\kappa$  chain-expressing chimeric mouse

[0184] In the same manner as in Example 5, cDNA was prepared from the spleen of the chimeric mouse K2-8 from Example 13 which was confirmed to express human antibody  $\kappa$  chain in Example 23. Using the resulting cDNA and the following primers prepared with reference to Larrick et al., BIO/TECHNOLOGY, 7, 934-, 1989; Whitehurst et al., Nucleic Acids Res., 20, 4929-, 1992, PCR was performed to amplify human antibody  $\kappa$  chain variable region. cDNA from the liver of the chimeric mouse K2-8 and cDNA from the spleen of the chimeric mouse K3-2-2 derived from the TT2/#14 3-2 clone (see Example 10) were used as negative controls.

KC2 (human Ig  $\kappa$  chain constant region) : 5' -

CAGAGGCAGTTCCAGATTTC (SEQ ID NO:50)

KC3 (ditto) : 5' - TGGGATAGAAGTTATTCAGC (SEQ ID NO:51)

KVMIX (human Ig  $\kappa$  chain variable region) : 5' -

ATGGACATG (AG) (AG) (AG) (AGT) (CT) CC (ACT) (ACG) G (CT) (GT) CA (CG) CTT (SEQ ID NO:52) (a mixture of 3456 sequences)

\* ( ) means that any one of the bases therein should be selected.

[0185] PCR was performed by using primer combinations of KVMIXxKC2 and KVMIXxKC3 in 40 cycles at 94 °C for 15 seconds, 55 °C for 15 seconds and 72 °C for 20 seconds with a Thermal Cycler 9600 (Perkin-Elmer Corp.). The amplification products were electrophoresed on a 1.5% agarose gel and detected by staining with ethidium bromide. As a result, expected amplification products of about 420 bp (KC2) and about 450 bp (KC3) were detected. In the case of the two negative controls, no specific amplification product was detected. These amplification products were cloned into a pBlueScriptII SK+ (Stratagene Ltd.) at a SmaI or EcoRI site in accordance with the method described in Ishida et al., "Gene Expression Experiment Manual", published by Kodansha Scientific, 1995. Among the amplification product-inserted plasmids, VK-#1 clone derived from the KVMIXxKC2 primers was selected and the nucleotide sequence of the amplification product was determined with a Fluorescence Autosequencer (Applied Biosystems Inc.). Since the obtained nucleotide sequence did not contain a termination codon at any site between an initiation codon and a constant region of human Ig  $\kappa$  chain, the cloned amplification products are believed to encode a variable region of functional human Ig  $\kappa$  chain. As a result of the comparison of the obtained nucleotide sequences with those of known human antibody V  $\kappa$  region (Klein et al., Eur. J. Immunol. 23, 3248-, 1993) and J  $\kappa$  region (Whitehurst et al., supra), it was revealed that the VK-#1 clone contained a combination of genes for V $\kappa$ 3 family and J $\kappa$ 4. These results show that the chimeric mouse retaining human chromosome #2 partial fragment produced a complete functional human antibody  $\kappa$

chain protein.

#### Example 29

- 5 Detection and quantitation of human antibody  $\gamma$  chain subclasses and  $\mu$  chain in sera of the chimeric mice retaining human chromosome #14 fragment

[0186] The chimeric mice K15A and K16A from Example 10 (derived from the 1-4 clone, with chimerism of 70% and 50%, respectively) of 11 weeks after birth were bled and human antibody  $\gamma$  chain subclasses and  $\mu$  chain in the sera were detected by the same ELISA method as in Example 14.

#### Quantative Determination of human IgG1

15 [0187] Ninety six-well microtiter plates were coated with anti-human IgG antibody (Sigma, I - 6260) diluted with PBS. A serum sample was added. Subsequently, peroxidase-labeled anti-human IgG1 antibody (Pharmingen, 08027E) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgG1 antibody (Sigma, I-3889) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS.

#### Quantative Determination of human IgG2

20 [0188] Ninety six-well microtiter plates were coated with anti-human IgG2 antibody (Sigma, I-9513) diluted with PBS. A serum sample was added. Subsequently, peroxidase-labeled anti-human IgG antibody (Sigma, A-0170) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgG2 antibody (Sigma, I-4139) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS.

#### Quantative Determination of human IgG3

30 [0189] Anti-human IgG3 antibody (Sigma, I-7260) was diluted with 100 mM glycine-HCl buffer (pH 2.5) and incubated for 5 minutes at room temperature, followed by 10-fold dilution with 100 mM phosphate buffer (pH 7.0). Ninety six-well microtiter plates were coated with the anti-human IgG3 antibody solution. A serum sample was added. Subsequently, peroxidase-labeled anti-human IgG antibody (Pharmingen, 08007E) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgG3 antibody (Sigma, I-4389) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS.

#### Quantative Determination of human IgG4

40 [0190] Anti-human IgG4 antibody (Sigma, I-7635) was diluted with 100 mM glycine-HCl buffer (pH 2.5) and incubated for 5 minutes at room temperature, followed by 10-fold dilution with 100 mM phosphate buffer (pH 7.0). Ninety six-well microtiter plates were coated with the anti-human IgG3 antibody solution. A serum sample was added. Subsequently, peroxidase-labeled anti-human IgG antibody (Pharmingen, 08007E) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgG4 antibody (Sigma, I-4639) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS.

#### Quantative Determination of human IgM

50 [0191] Ninety six-well microtiter plates were coated with anti - human  $\mu$  chain mouse monoclonal antibody (Sigma, I - 6385) diluted with PBS. A serum sample was added. Subsequently, peroxidase - labeled anti-human  $\mu$  chain mouse antibody (The Binding Site Limited, MP008) diluted with mouse serum (Sigma, M5905)-supplemented PBS was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added as a peroxidase substrate, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgM having  $\mu$  chain (CAPPEL, 6001-1590) was used as standard. The standard was diluted stepwise with mouse serum (Sigma, M5905)-supplemented PBS.

55 [0192] The results are shown in Table 10. All the subclasses IgG1, IgG2, IgG3 and IgG4, and IgM were detected in the two chimeric mice K15A and K16A.

Table 10.

Concentrations of Human antibody IgG Subclasses and IgM in the Chimeric Mice (ELISA)					
Chimeric mouse	IgG1	IgG2	IgG3	IgG4	IgM (mg/l)
K15A	2.25	1.96	0.17	0.43	7.09
K16A	0.30	0.69	0.10	0.07	0.87

### Example 30

Preparation of mouse ES cell clones (TT2) retaining human chromosome #22

[0193] The cell clone 6-1 (A9/#22, G418 and puromycin resistant) from Example 26 was used as a chromosome donor cell for the preparation of mouse ES cell (TT2) retaining human chromosome #22. A wild-type TT2 cell line (see Example 9) was used as a chromosome recipient cell. The microcell fusion and the selection of puromycin resistant clones were conducted by the same procedures as in the selection of G418 resistant clones in Example 9 except that the concentration of puromycin was 0.75  $\mu\text{g/ml}$ . The frequency of the appearance of the puromycin resistant clones was 1-2 per  $10^7$  of TT2 cells. The puromycin resistant clones were stored frozen and genomic DNA was prepared by the same methods as in Example 2. The retention of human chromosome #22 in the puromycin resistant clone PG22-1 was confirmed by the methods described in (1) and (2) below.

#### (1) PCR analysis

[0194] Genomic DNA of the puromycin resistant clone was used as a template in PCR amplification. Among the genes on human chromosome #22 (Genetic Maps, supra), ten primers whose presence in the A9/#22 clone was confirmed in Example 2 were used in the PCR amplification. All the markers which existed in the A9/#22 clone (see Example 2) were detected.

#### (2) Southern blot analysis

[0195] In accordance with the same method as described in Example 2 using human L1 sequence as a probe, Southern blot analysis was conducted with genomic DNAs obtained from wild-type TT2 (negative control), the chromosome donor cell 6-1 and the puromycin resistant TT2 cell clone PG22-1. The results are shown in Fig. 19. In Fig. 19, the molecular weights of DNAs are shown at the left side. The band pattern of the PG22-1 clone was equivalent to that of the 6-1 cell and the signal intensities were the same. Hence, it was confirmed that chromosome #22 in the 6-1 cell had been transferred certainly into the PG22-1 clone.

[0196] These experiments demonstrate that the puromycin resistant TT2 cell clone PG22-1 retained the whole or the most part of human chromosome #22.

### Example 31

Production of chimeric mice from the mouse ES cells (TT2) retaining human chromosome #22

[0197] The cells in a frozen stock of the puromycin resistant TT2 cell clone PG22-1 from Example 30 which was confirmed to retain human chromosome #22 were thawed, started to culture and injected into 8-cell stage embryos obtained by mating ICR or MCH(ICR) male and female mice (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. The embryos were cultured in a medium for ES cells (see Example 9) overnight to develop to blastocysts. Two and a half day after a foster mother ICR mouse was subjected to a pseudopregnant treatment, about ten of the injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 11.

Table 11.

Production of chimeric mice from the TT2 cell clone retaining human chromosome #22							
ES cell clone/ human chromosome	Puromycin resistant clone No.	Number of ES cell- injected 8-cell stage embryos	Number of offspring mice	Number chimeric mice	Contribution to coat color		
					<20%	20-50%	50-80%
TT2/#22	PG22-1	266	36	8	4	1	3

[0198] As a result of the transplantation of a total of 266 injected embryos, 36 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo-derived albino coat color. Out of the 36 produced offsprings, 8 mice were recognized to have a partial agouti coat color, indicating the contribution of the ES cells.

[0199] These results show that the ES cell clone (derived from TT2, PG22-1) retaining human chromosome #22 maintain the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse.

#### Example 32

Detection and quantitation of human antibody  $\lambda$  chain in sera of the chimeric mice retaining human chromosome #22

[0200] The concentration of human antibody  $\lambda$  in the sera of the chimeric mice KPG22-1, 2 and 3 from Example 31 was determined by ELISA in accordance with the same procedure as in Example 14. The chimeric mice of 2 months after birth were bled and human antibody  $\lambda$  chain in the sera was detected by ELISA. Ninety six-well microtiter plates were coated with anti-human immunoglobulin  $\lambda$  chain antibody (VECTOR LABORATORIES INC., IA-3070) diluted with PBS and then a serum sample was added. Subsequently, biotin-labeled anti-human immunoglobulin  $\lambda$  chain antibody (VECTOR LABORATORIES INC., BA-3070) was added to the plates and incubated. A complex of biotinylated horse-radish peroxidase and avidin DH (VECTOR LABORATORIES, INC., Vectastain ABC Kit) was added and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgM antibody having  $\lambda$  chain (Dainippon Pharmaceutical Co., Ltd., U13200) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS. The results are shown in Table 12. These results show that human antibody  $\lambda$  chain gene can function in the chimeric mice retaining human chromosome #22.

Table 12.

Concentration of Human Antibody $\lambda$ Chain in Chimeric Mice (ELISA)		
Chimeric Mouse	%Chimerism	Ig $\lambda$ (mg/l)
KPG22-1	50	12
KPG22-2	50	18
KPG22-3	20	24

#### Example 33

Detection of anti-human HSA human antibody  $\lambda$  chain in a serum of the human chromosome #22 transferred chimeric mouse

[0201] The chimeric mouse KPG22-3 from Example 31 was immunized with HSA, 3 times at days 79, 94 and 110 after birth by the same method as in Example 20. Human antibody  $\lambda$  chain in the serum of the chimeric mouse was detected by ELISA in accordance with the same procedure as in Example 14. Ninety six-well microtiter plates were coated with HSA (Sigma, A 3782) diluted with 50 mM carbonate-bicarbonate buffer (pH 9.6) to a concentration of 5  $\mu$ g/ml and a serum sample was added. Biotinylated anti-human Ig  $\lambda$  antibody (VECTOR LABORATORIES INC., BA-3070) was added. Subsequently, a complex of biotinylated-horseradish peroxidase and avidin DH (VECTOR LABORATORIES, INC., Vectastain ABC Kit) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-



1120T) was added, enzyme activity was determined by absorbance measurement at 450 nm. The titer of the anti-HSA human  $\lambda$  chain in the serum of the chimeric mouse increased after the immunization. On the other hand, control ICR mouse gave a background level of the anti-HSA human  $\lambda$  chain titer after the immunization with HSA. The results are shown in Fig. 20. In Fig. 20, the number of days after the first immunization of the chimeric mouse with HSA is plotted on the horizontal axis and the absorbance at 450 nm is plotted on the vertical axis. These results show that human antibody  $\lambda$  chain gene can function in the chimeric mouse retaining human chromosome #22 and that the antibody titer of the antigen specific human Ig  $\lambda$  was increased by stimulation with the HSA antigen.

#### Example 34

Preparation of human antibody light chain-producing hybridomas from the human chromosome #22 transferred chimeric mouse

[0202] The spleen was removed from the mouse KPG22-3 (see Example 33) at day 113 after birth by the same method as in Example 25. A spleen cell was fused with a myeloma cell to produce a hybridoma. The hybridoma was produced using a myeloma cell SP-2/0-Ag14 (Dainippon Pharmaceutical Co., Ltd., 05-554) by the method described in Toyama and Ando, "Monoclonal Antibody Experiment Manual", published by Kodansha Scientific, 1991. The cells were inoculated into a medium containing 10% HCF (Air Brown) in five 96-well plates and cultured for 1-3 weeks. The supernatant of the culture solution in colony-positive wells was analyzed by ELISA. The ELISA analysis was conducted by the same method as in Example 33 and four human antibody  $\lambda$  chain-positive clones were obtained.

#### Example 35

Preparation of mouse ES cell clones retaining both a human chromosome #22 partial fragment and a human chromosome #14 partial fragment

[0203] The 6-1 cell clone from Example 26 (A9/#22, G418 and puromycin resistant) was used as a chromosome donor cell for the preparation of mouse ES cells retaining both a human chromosome #22 partial fragment and a human chromosome #14 partial fragment. The G418 resistant TT2 cell clone 1-4 retaining a human chromosome #14 partial fragment from Example 9 was used as a chromosome recipient cell. The experiment of microcell fusion and the selection of puromycin resistant cells were carried out by the same methods as in the selection of the G418 resistant clones in Example 9 except that the concentration of puromycin was 0.75  $\mu$ g/ml. As a result, the frequency of the appearance of the puromycin resistant clones was 1-2 per  $10^7$  of 1-4 cells. The retention of G418 resistance in the puromycin resistant clones was confirmed from the fact that these clones were grown in the presence of 300  $\mu$ g/ml G418. The double-drug resistant clones were stored frozen and genomic DNAs were prepared by the same methods as in Example 2. The retention of human chromosome #22 and a human chromosome #14 partial fragment in the double-drug resistant clone PG22-5 was confirmed by PCR analysis. With genomic DNA of the double-drug resistant clone used as a template, PCR amplification was conducted using primers whose presence on chromosome #22 was confirmed in Example 2 (A9/#22) and primers whose presence on chromosome #14 was confirmed in Example 9 (TT2/#14 1-4); as a result, three markers (D22S275, D22S315 and Ig  $\lambda$ ) of the ten markers on chromosome #22 and all of the markers on chromosome #14 in the TT2/#14 1-4 clone were detected.

[0204] These experiments demonstrate that the obtained double-drug resistant TT2 cell clone retained both a human chromosome #22 partial fragment and a human chromosome #14 partial fragment.

#### Example 36

Production of the chimeric mouse from the mouse ES cell clone retaining both a human chromosome #22 partial fragment and a human chromosome #14 partial fragment

[0205] The cells in a frozen stock of the G418 and puromycin double-resistant TT2 cell clone PG22-5 from Example 35 which was confirmed to retain a human chromosome #22 partial fragment and a human chromosome #14 partial fragment were thawed, started to culture and injected into 8-cell stage embryos obtained by mating ICR or MCH(ICR) male and female mice (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. The embryos were cultured in a medium for ES cells (see Example 9) overnight to develop to blastocysts. Two and a half day after a foster mother ICR mouse was subjected to a pseudopregnant treatment, about ten of the injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 13.

Table 13.

Production of the chimeric mouse from the mouse ES cell clone retaining both a human chromosome #22 partial fragment and a human chromosome #14 partial fragment							
ES cell clone/ human chromosome	Double-drug resistant clone No.	Number of ES cell- injected 8-cell stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color		
					<20%	20-50%	50-80%
TT2/#22+#14	PG22-5	302	16	5	3	2	0

[0206] As a result of the transplantation of a total of 302 injected embryos, 16 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo-derived albino coat color. Out of the 16 produced offsprings, 5 mice were recognized to have a partial agouti coat color, indicating the contribution of the ES cell.

[0207] These results show that the ES cell clone PG22-5 retaining a human chromosome #22 partial fragment and a human chromosome #14 partial fragment maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse.

#### Example 37

Detection of human antibody  $\lambda$  chain and  $\mu$  chain in sera of the chimeric mice derived from the ES cells retaining both a human chromosome #22 partial fragment and a human chromosome #14 partial fragment

[0208] The chimeric mice KPG22-9, 10 and 12 from Example 36 were immunized with HSA. The chimeric mice KPG22-9 and 10 were immunized 11 weeks after birth and bled 2 weeks after the immunization. The chimeric mouse KPG22-12 was immunized twice at 7 and 11 weeks after birth and bled 2 weeks after the second immunization.

[0209] A serum human antibody  $\mu$  chain, a serum human antibody  $\lambda$  chain, and a serum antibody having both human antibody  $\lambda$  and  $\mu$  chains were detected by ELISA in accordance with Example 14.

[0210] For the detection of complete human antibody molecules, 96-well microtiter plates were coated with anti-human immunoglobulin  $\lambda$  chain antibody (Kirkegaard & Perry Laboratories Inc., 01-10-11) diluted with PBS and a serum sample was added. Subsequently, peroxidase-labeled anti-human immunoglobulin  $\mu$  chain antibody (The Binding Site Limited, MP008) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added as a peroxidase substrate, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgM antibody having  $\lambda$  chain (Dainippon Pharmaceutical Co., Ltd., U13200) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS. Human antibody  $\mu$  and  $\lambda$  chains were detected and determined quantitatively by ELISA in the same manner as in Examples 29 and 32. The results are shown in Table 14.

Table 14.

Concentrations of Human Antibodies in Chimeric Mice (ELISA)					
ES clone	Chimeric mouse	Chimerism (%)	IgM (mg/l)	Ig $\lambda$ (mg/l)	IgM, $\lambda$ (mg/l)
PG22-5	KPG22-9	30	2.54	9.9	0.043
PG22-5	KPG22-10	5	4.96	21.5	0.333
PG22-5	KPG22-12	40	3.71	7.0	0.048
3-2	K9	50	6.66	-	<0.003
PG22-1	KPG22-2	50	-	17.6	<0.003

[0211] Both  $\lambda$  and  $\mu$  chains were detected in the chimeric mice. An antibody molecule having both human antibody  $\mu$  and  $\lambda$  chains was detected. These results show: the human antibody  $\lambda$  chain gene and human antibody  $\mu$  chain gene can function at the same time in the chimeric mice derived from the ES cells retaining human chromosome #22 partial fragments and human chromosome #14 partial fragments; and a complete antibody containing both human heavy and light chains was produced in part of the B cells.

[0212] The control mice, that is, the chimeric mouse K9 retaining only human chromosome #14 from Example 10

and the chimeric mouse KG22-2 retaining only human chromosome #22 from Example 31, gave background levels of an antibody having both human antibody  $\lambda$  and  $\mu$  chains in the sera. It was confirmed that in these detection systems, only a complete antibody molecule having human  $\lambda$  and  $\mu$  chains was detected.

#### Example 38

Detection of human antibody having human  $\kappa$  and  $\mu$  chains in sera of the chimeric mice derived from the ES cells retaining both human chromosome #2 partial fragments and human chromosome #14 partial fragments

[0213] The chimeric mouse KPG-15 (derived from the TT2ES clone PG5, 10% chimerism) was immunized during 2-3 months after birth 3 times with 0.2 ml of a solution of human serum albumin (HSA, Sigma, A3782) and adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc.) in PBS at a HSA concentration of 0.25 mg/ml and bled (see Example 15). The chimeric mouse KPG-26 (derived from the TT2ES clone PG6, 40% chimerism) of 6 weeks after birth was bled. The concentration of a complete human antibody molecule in the sera was determined by ELISA in accordance with Example 14. Ninety six-well microtiter plates were coated with anti-human immunoglobulin  $\kappa$  chain antibody (Kirkegaard & Perry Laboratories Inc., 01-10-10) diluted with PBS, and a serum sample was added. Subsequently, peroxidase-labeled anti-human immunoglobulin  $\mu$  chain antibody (The Binding Site Limited, MP008) was added to the plates and incubated. After TMBZ (Sumitomo Bakelite, ML-1120T) was added as a peroxidase substrate, enzyme activity was determined by absorbance measurement at 450 nm. Purified human IgM antibody having  $\kappa$  chain (CAPPEL, 6001-1590) was used as standard. The standard was diluted stepwise with mouse serum-supplemented PBS. The concentrations of  $\kappa$  chain and  $\mu$  chain were determined by the same method as in Example 20. The results are shown in Table 15.

Table 15.

Concentrations of Human Antibodies in Chimeric Mice (ELISA)					
ES clone	Chimeric mouse	Chimerism (%)	IgM (mg/l)	Ig $\kappa$ (mg/l)	IgM, $\kappa$ (mg/l)
PG-5	KPG15	10	0.18	1.01	0.075
PG-6	KPG26	40	1.52	1.26	0.018
3-2	K9	50	6.66	-	<0.002
5-1	K2-9	40	-	135	<0.002

[0214] A antibody molecule having both human antibody  $\mu$  and  $\kappa$  chains was detected. The control mice, that is, the chimeric mouse K9 retaining only human chromosome #14 from Example 10 and the chimeric mouse K2-9 retaining only human chromosome #2 from Example 13, gave background levels (<0.002 mg/ml) of an antibody having human antibody  $\kappa$  and  $\mu$  chains in the sera. These results show: the human antibody  $\kappa$  chain gene and human antibody  $\mu$  chain gene can function at the same time in the chimeric mice derived from the ES cells retaining both human chromosome #2 partial fragments and human chromosome #14 partial fragments; and a complete antibody molecule containing both human heavy and light chains was produced in part of the B cells.

#### Example 39

Preparation of mouse ES cell clone (TT2F, XO) retaining a human chromosome #2 partial fragment

[0215] The cell clone PG1 from Example 16 was used as a chromosome donor cell for the preparation of a mouse ES cell (XO) retaining a human chromosome #2 partial fragment. A TT2F cell (purchased from Lifetec Oriental Co.) having a karyotype of (39, XO), which was reported to differentiate efficiently into an oocyte in chimeric mice (Shinichi Aizawa, "Biomaterial Series 8, Gene Targeting" published by Yodosha, 1995), was used as a chromosome recipient cell. The experiment of microcell fusion and the selection of puromycin resistant cells were carried out by the same methods as in the selection of the G418 resistant clones in Example 9 except that the concentration of puromycin was 0.75  $\mu$ g/ml. The frequency of the appearance of the puromycin resistant clones was 5 per  $10^7$  of TT2F cells. The puromycin resistant clones were stored frozen and genomic DNAs were prepared from the clones by the same methods as in Example 2. The retention of human chromosome #2 partial fragments in the drug resistant clones P-20 and P-21 was confirmed by PCR analysis. As a result of PCR amplification using genomic DNAs of the drug resistant clones as templates and three kinds of primers C  $\kappa$ , FABP1 and V  $\kappa$  1 - 2 whose presence in the A9/#2 W23 clone was confirmed in Example 1, all of the three primers gave expected amplification products in both of the two clones.

[0216] These experiments demonstrate that the obtained puromycin resistant ES cell clone (TT2F, XO) retained a

human chromosome #2 partial fragment.

#### Example 40

- 5 Production of the chimeric mice from the mouse ES cell clone (TT2F; XO) retaining a human chromosome #2 partial fragment

10 [0217] The cells in a frozen stock of the puromycin resistant TT2F cell clone P-21 from Example 39 which was confirmed to retain a human chromosome #2 partial fragment were thawed, started to culture and injected into 8-cell stage embryos obtained by mating ICR or MCH(ICR) male and female mice (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. The embryos were cultured in a medium for ES cells (see Example 9) overnight to develop to blastocysts. Two and a half day after a foster mother ICR mouse was subjected to a pseudopregnant treatment, about ten of the injected embryos were transplanted to each side of the uterus of the foster mother. The results are shown in Table 16.

Table 16.

Production of the chimeric mice from the TT2F cell clone retaining a human chromosome #2 partial fragment								
ES cell clone/ human chromosome	Puromycin resistant clone No.	Number of ES cell- injected 8-cell stage embryos	Number of offspring mice	Number of chimeric mice	Contribution to coat color			
					<20%	20-50%	50-90%	100%
TT2F/#2fg.	P-21	141	20	9	0	2	3	4

30 [0218] As a result of the transplantation of a total of 141 injected embryos, 20 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2F cell-derived agouti coat color (dark brown) in the host embryo-derived albino coat color. Out of the 20 produced offsprings, 9 mice were recognized to have a partial agouti coat color, indicating the contribution of the ES cell. Four of the 9 mice were chimeric mice having a full agouti coat color from the ES cells.

35 [0219] These results show that the ES cell clone P-21 retaining a human chromosome #2 partial fragment maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mouse.

#### Example 41

40 Detection and quantitative determination of human antibody  $\kappa$  chain in sera of the chimeric mice derived from the TT2F clone retaining a human chromosome #2 partial fragment

[0220] The chimeric mice K2-1F, 2F, 3F and 4F (derived from the P-21 clone, 100% chimerism) from Example 40 of about 1 month after birth were bled and the concentration of human antibody  $\kappa$  chain in the sera was determined quantitatively by ELISA in the same manner as in Example 20.

45 [0221] The results are shown in Table 17. It was confirmed that the human antibody  $\kappa$  chain gene could function in the chimeric mice when the TT2F was used as an ES cell.

Table 17

Concentration of Human Antibody $\kappa$ Chain in Chimeric Mice (ELISA)		
Chimeric mouse	%Chimerism	Ig $\kappa$ (mg/l)
K2-1F	100	66
K2-2F	100	156
K2-3F	100	99
K2-4F	100	20

## Example 42

Confirmation of the retention of human chromosome in progenies of the chimeric mice derived from the mouse ES cell (TT2F, XO) retaining a human chromosome #2 partial fragment

[0222] Examination was made as to whether ES cell-derived progenies would be reproduced by mating the female chimeric mice K2-1F and K2-4F (both were of 100% chimerism in coat color) from Example 40 with ICR male mice. In such a mating, offspring mice of an agouti coat color should be reproduced from the oocytes derived from the TT2F cell (agouti coat color, dominant) in the chimeric mice and offspring mice of an albino coat color should be reproduced from oocytes derived from ICR if the oocytes are fertilized with the sperms from ICR male mice (albino, recessive). All the viable offspring mice (K2-1F, 10 mice and K2-4F, 5 mice) obtained by one mating of the respective combinations had an agouti coat color which derived from the ES cells. The retention of human chromosome fragments in genomic DNAs prepared from the tails of the offspring mice was examined by a PCR method. As a result of the PCR amplification using three kinds of primers whose presence in the P-21 clone (see Example 39) was confirmed, the presence of these three markers was confirmed in 4 out of the ten mice from K2-1F and in 2 out of the five mice from K2-4F. The results of the PCR of these 15 offspring mice are shown in Fig. 21. In Fig. 21, markers ( $\phi$ X174/HaeIII fragment, Nippongene) and the DNA molecular weights of main bands are shown at the right side and the lengths of expected products of amplification with the respective primers are shown by arrows at the left side. At the right side, the results with tail-derived DNA of the mother chimeric mice K2-1F and K2-4F (positive controls) are shown. These results show that the TT2 cell clone P-21 differentiated into functional oocyte in the chimeric mice and that a human chromosome #2 partial fragment was transmitted to offsprings through oocyte.

## Example 43

Confirmation of the retention of human chromosome in progenies of the chimeric mice derived from the mouse ES cell (TT2, XY) retaining a human chromosome #2 partial fragment

[0223] Examination was made as to whether ES cell-contributed offspring mice would be produced by mating K2-18 (70% chimeric male mouse from Example 13) with K2-19 (60% chimeric female mouse of Example 13) or non-chimeric female littermates. Since TT2 cell has the karyotype of (40, XY), it may differentiate into a functional sperm in the male chimeric mouse K2-18. If this is the case, offspring mice of an agouti coat color should be reproduced from ICR (albino, recessive)-derived oocytes fertilized with sperms from TT2 cell (agouti color, dominant) in the chimeric mice. While a total of 110 viable offspring mice were obtained by the mating, ten had an agouti coat color which derived from the ES cells. The retention of human chromosome fragments in genomic DNAs prepared from the tails of 7 out of the ten offspring mice of an agouti coat color was examined by a PCR method. As a result of PCR amplification using two kinds of primers C $\kappa$  and FABP1 whose presence in the 5-1 clone (TT2/#2fg., Example 12) was confirmed and primer V $\kappa$ 1-2 which was shown in Example 1; the presence of all of the three markers was confirmed in 2 out of the seven mice. These results show that the TT2 cell clone 5-1 retaining a human chromosome #2 partial fragment differentiated into functional sperms in the chimeric mice and that the human chromosome #2 partial fragment was transmitted to offsprings through the sperms.

## Example 44

Detection and quantitative determination of human antibody  $\kappa$  chain in sera of offspring mice of the chimeric mice

[0224] The concentration of human antibody  $\kappa$  chain in the sera of the offspring mice K2-1F-1~10 and K2-4F-1~5 from Example 42 was determined quantitatively by ELISA. The mice of about 4-6 months after birth were bled and the concentration of human antibody  $\kappa$  chain in the sera was determined by ELISA in the same manner as in Example 20.

[0225] The results are shown in Table 18 together with the data obtained in Example 42 on the retention of chromosome. It was confirmed that the human antibody  $\kappa$  chain gene can function in the offspring mice reproduced from the chimeric mice.

Table 18.

Concentration of Human Antibody $\kappa$ Chain in Offspring Mice (ELISA)			
Mother mouse	Number of mouse	Presence of human chromosome #2 fragments	Ig $\kappa$ (mg/l)
K2-1F	#1	-	0.58

Table 18. (continued)

Concentration of Human Antibody $\kappa$ Chain in Offspring Mice (ELISA)			
Mother mouse	Number of mouse	Presence of human chromosome #2 fragments	Ig $\kappa$ (mg/l)
K2-1F	#2	+	84.1
K2-1F	#3	+	12.8
K2-1F	#4	+	15.1
K2-1F	#5	-	0.52
K2-1F	#6	-	0.58
K2-1F	#7	-	1.30
K2-1F	#8	-	0.90
K2-1F	#9	-	0.56
K2-1F	#10	+	28.8
K2-4F	#1	-	< 0.04
K2-4F	#2	+	23.3
K2-4F	#3	+	11.8
K2-4F	#4	-	0.08
K2-4F	#5	-	0.06

## Example 45

Analysis of spleen cells from the human chromosome #14 partial fragment transferred chimeric mice

[0226] Flow cytometry analysis was accordance with the method described in "New Biochemical Experiment Lecture 12, molecular immunology I-Immunocells - Cytokines-", edited by the Japanese Biochemical Society, 1989, published by Tokyo Kagaku Dojin; "Cell Technology Separated Volume 8, New Cell Technology Experiment Protocol", edited by the University of Tokyo, Medical Science Institute, Anti-cancer Laboratory, 1991, published by Shujunsha; and A.Doyle and J.B.Griffiths, "Cell & Tissue Culture: Laboratory Procedures", published by John Wiley & Sons Ltd., 1996. The spleen was removed from the chimeric mouse KPG06 (derived from the PG16 clone, 30% chimerism) from Example 19 of six months after birth and treated with an aqueous solution of ammonium chloride. The spleen cells were stained with fluorescein isothiocyanate (FITC)-labeled anti-mouse CD45R (B220) antibody (Pharmingen, 01124A) in PBS containing 1% rat serum. After being washed, the cells were reacted with 0.1  $\mu$ g of biotin-labeled anti-human IgM antibody (Pharmingen, 08072D) or a control biotin-labeled anti-human  $\lambda$  chain antibody (Pharmingen, 08152D) in PBS containing 5% mouse serum and stained with 0.1  $\mu$ g of streptavidin-phycoerythrin (Pharmingen, 13025D), followed by analysis with a flowcytometer (Becton Dickinson Immunocytometry Systems, FACSsort). An ICR mouse retaining no human chromosome was used as a control for analysis by the same method. The results are shown in Fig. 22. In Fig. 22, the human IgM is plotted on the horizontal axis and the CD45R (B220) is plotted on the vertical axis. A population of cells positive to both B cell marker CD45R (FITC) and human IgM (PE) increased by 4%, indicating that cells expressing human antibody  $\mu$  chain on the cell surfaces were present in the chimeric mice.

## Example 46

Cloning and sequencing of variable regions of human antibody genes from cDNA derived from the spleen of the chimeric mice expressing human antibody heavy chain,  $\kappa$  and  $\lambda$  chains, respectively

[0227] In the same manner as in Example 5, cDNAs were synthesized from RNAs extracted from the spleens of the chimeric mice K15A (derived from the 1-4 clone, prepared by the method described in Example 10), K2-8 prepared in Example 13 and KPG22-2 prepared in Example 31, all of which were confirmed to express human antibody heavy chain,  $\kappa$  and  $\lambda$  chains in Examples 29, 23 and 32, respectively. PCR was performed using the respective cDNAs and the following primers to amplify the variable regions of respective human antibody. cDNA derived from the spleen of a non-chimeric mouse ICR was used as a negative control. Those primers set forth below without indication of reference literature were designed on the basis of the nucleotide sequences obtained from data bases such as Genbank and the like.

· K15A (heavy chain)

[0228] For constant region:

HIGMEX1 - 2 : 5' - CCAAGCTTCAGGAGAAAGTGATGGAGTC (SEQ ID NO: 53)

HIGMEX1 - 1 : 5' - CCAAGCTTAGGCAGCCAACGGCCACGCT (used in 2nd PCR of VH3BACK) (SEQ ID NO: 54)

For variable region:

VH1/5BACK (59°C, 35 cycles, Marks et al., Eur. J. Immunol., 21, 985-, 1991),  
VH4BACK (59°C, 35 cycles, Marks et al., supra), and  
VH3BACK (1st PCR: 59°C, 35 cycles; 2nd PCR: 59°C, 35 cycles, Marks et al., supra)

· K2 - 8 (light chain κ)

[0229] For constant region:

KC2H : 5' - CCAAGCTTCAGAGGCAGTTCAGATTTC (SEQ ID NO: 55)

For variable region:

Vk1/4BACK (55°C, 40 cycles, Marks et al., Eur. J. Immunol., 21, 985-, 1991),  
Vk2BACK (55°C, 40 cycles, Marks et al., supra), and  
Vk3BACK (55°C, 40 cycles, Marks et al., supra)

· KPG22-2 (light chain λ)

[0230] For constant region:

C λ MIX (a mixture of the following three kinds of primers at an equal molar ratio)

IGL1 - CR : 5' - GGGAATTCGGGTAGAAGTCACTGATCAG (SEQ ID NO: 56)

IGL2 - CR : 5' - GGGAATTCGGGTAGAAGTCACTTATGAG (SEQ ID NO: 57)

IGL7 - CR : 5' - GGGAATTCGGGTAGAAGTCACTTACGAG (SEQ ID NO: 58)

For variable region:

Vλ 1LEA1 (55°C, 40 cycles, Williams et al., Eur. J. Immunol., 23, 1456-, 1993),  
Vλ 2MIX (55°C, 40 cycles, a mixture of Vλ 2 LEA1 and Vλ 2 JLEAD (Williams et al. (supra)) at an equal molar ratio)  
Vλ 3MIX (55°C, 40 cycles, a mixture of Vλ 3LEA1, Vλ 3JLEAD and Vλ 3BACK4, which were reported in Williams

et al. (supra) at an equal molar ratio.

[0231] The PCR was performed with combinations of the primers for constant regions with those for variable regions (3 primer pairs each for heavy chain,  $\kappa$  and  $\lambda$  chains) at 94 °C for 15 seconds, at the annealing temperatures shown with respect to the respective primers for variable region for 15 seconds, at 72 °C for 20 seconds in the cycle numbers shown with respect to the respective primers for variable region using a Thermal Cycler 9600 (Perkin-Elmer Corp.). In the second run of PCR using VH3BACK, the amplification products of the first run of PCR were amplified again with a combination of the two primers H1GMEX1-1 and VH3BACK. All of the amplification products were electrophoresed on a 1.5% agarose gel and stained with ethidium bromide for detection. As a result, the amplification products having expected lengths (heavy chain, about 470 bp; light chain  $\kappa$ , about 400 bp; and light chain  $\lambda$ , about 510 bp) were detected in all of the combinations. In the negative control, specific amplification product was not detected at the same position in any of the combinations. The obtained amplification products were extracted from the agarose gel using prep.A.gene (Bio-Rad Laboratories, Inc.), treated with restriction enzymes (heavy chain, HindIII and PstI; light chain  $\kappa$ , HindIII and PvuII; and light chain  $\lambda$ , HindIII and EcoRI), and cloned into pUC119 (TAKARA SHUZO CO., LTD.) at the sites of HindIII/PstI (heavy chain), HindIII/HincII ( $\kappa$  chain) and HindIII/EcoRI ( $\lambda$  chain). The nucleotide sequences of the products that were amplified with the following primers and which were cloned into the plasmids were determined with a Fluorescence Autosequencer (Applied Biosystems Inc.).

- HIGMEX1 - 2 × VH1/5BACK: 10 clones
- HIGMEX1 - 2 × VH4BACK: 8 clones
- HIGMEX1 - 2 (2nd PCR, HIGMEX1 - 1) × VH3BACK: 5 clones
- KC2H × v  $\kappa$  1/4BACK: 6 clones
- KC2H × v  $\kappa$  2BACK: 7 clones
- KC2H × v  $\kappa$  3BACK: 4 clones
- C  $\lambda$  MIX × V  $\lambda$  1LEA1 : 5 clones
- C  $\lambda$  MIX × V  $\lambda$  2MIX: 6 clones
- C  $\lambda$  MIX × V  $\lambda$  3MIX: 5 clones

[0232] The obtained nucleotide sequences were analyzed with DNASIS (Hitachi Software Engineering Co., Ltd.). The results show that all of the sequences were derived from human and that they were functional sequences which did not contain a termination codon at any site between an initiation codon and a constant region: this was true with all of the  $\kappa$  and  $\lambda$  chains and with 21 out of a total of 23 heavy chains. When the same sequences were removed from the determined sequences, unique variable region sequences were identified. as follows: 17 heavy chains, 11  $\kappa$  chains, and 12  $\lambda$  chains.

#### Example 47

Analysis of the nucleotide sequences of variable region of human antibody genes from cDNA derived from the spleen of the chimeric mouse expressing human antibody heavy chain,  $\kappa$  and  $\lambda$  chains, respectively

[0233] The nucleotide sequences determined in Example 46 (heavy chain, 17 clones;  $\kappa$  chain, 11 clones; and  $\lambda$  chain, 12 clones) were analyzed in the following points.

1. Identification of known germ line V gene segments used in the respective variable regions
2. Identification of known germ line J gene segments used in the respective variable regions
3. Identification of known germ line D gene segments used in the heavy chain variable regions
4. Identification of the addition of N region in the heavy chain variable regions on the basis of the results of 1, 2 and 3
5. Determination of the amino acid sequences deduced from the nucleotide sequences of the respective variable regions

[0234] The results are shown in Table 19. For the identification in points of 1 and 2, search for homology with germ line V and J segments registered in Genbank and the like was conducted with DNASIS. The VH segments, V  $\kappa$  segments and V  $\lambda$  segments are shown in Table 19 together with the family names of the respective V fragments in accordance with the conventions described in Cook et al., Nature genetics, 7, 162-, 1994 (VH fragments), Klein et al., Eur. J. Immunol, 23, 3248-, 1993 (V $\kappa$  fragments) and Williams et al. (supra) (V $\lambda$  fragments), respectively. For the identification in point 3, search for homology with germ line D fragments reported in Ichihara et al., The EMBO J., 7, 13, 4141-, 1988 was conducted with DNASIS. Assignment was based on at least 8 bp identity and the results are shown in Table 19. DN1\* is believed to be the new DN family segment reported in Green et al., Nature Genetics, 7, 13-, 1994. For the identification in point 4, the nucleotide sequences which did not appear in any germline sequences were determined



to be N regions on the basis of the results for 1(V), 2(J) and 3 (D). As a result, N region was observed in 11 of the 13 sequences in which D segment was identified and its average length was 8.7 bp. For the determination in point 5, the respective sequences were converted by DNASIS to amino acid sequences which were expressed with one letter symbols. In Table 19, only CDR3 region is shown. At the right side of Table 19, the names of the primers used in cloning of the respective variable regions and the names of clones are shown.

Table 19

V family	V segment	CDR3	J (D)	V primer	Clone
K15A					
VH1	VH1-8	VRSSSWY E Y Y Y Y GMDV	J6 (DN1)	VH4BACK	H4-10
	VH1-18	GGITMVRGLIITDWYFDL	J2 (DXP'1)	VH1/5BACK	H1-7
	VH1-24	APYSGRFDY	J4 (DK1)	VH1/5BACK	H1-6
	VH1-46	ERYYGSGSYQDY Y Y Y Y GMDV	J6 (DXP1)	VH1/5BACK	H1-2
	VH1-46	GGYSGYEDY Y Y Y Y GMDV	J6 (DK1)	VH1/53ACK	H1-10
VH2	VH2-5	SYFDWPDFDY	J4 (DXP1)	VH4BACK	H4-14
VH3	VH3-21	EGCSGGSCLPGY Y Y Y Y GMDV	J6 (DLR2)	VH1/5BACK	H1-4
	VH3-23	AHGDPYFDY	J4	VH1/5BACK	H1-3
	VH3-23	DADAFDI	J3	VH1/5BACK	H1-8
	VH3-23	SGWDY	J4 (DN1")	VH3BACK	H3-3
	VH3-23	TGFDL	J2	VH4BACK	H4-4
	VH3-33	EGGYGSGVDY Y Y Y Y GMDV	J6 (DXP'1)	VH1/5BACK	H1-9
	"VH3-33	GGYSYGYDY Y Y Y Y GMDV	J6 (DXP'1)	VH3BACK	H3-5
	VE3-33	GYSSGWYDY	J4 (DN1")	VH4BACK	H4-9
VH4	VH4-34	RYSSGWY Y Y FDY	J4 (DN1")	VH4BACK	H4-15
	VH4-59	GRIAVASFDY	J4 (DN1")	VH4BACK	H4-2
	VH4-59	GSGSYFHFDY	J4	VH4BACK	H4-6
K2-8					
V κ 1	O18-8	QQHDNLPFT	J3	V κ 1BACK	K1-1
	O18-8	QQYDNLPIIT	J5	V κ 1BACK	K1-3
	O18-8	QQHDNLPFA	J3	V κ 2BACK	K2-2
	L1	QQYNSYPLT	J4	V κ 1BACK	K1-6
V κ 2	A17	MQGTHLLT	J4	V κ 2BACK	K2-1
	A17	MQGTHWIT	J5	V κ 2BACK	K2-5
V κ 3	A27	QQYGSSPTWT	J1	V κ 3BACK	K3-1
	A27	QQYGSSPFT	J3	V κ 3BACK	K3-4
	A27	QQYGSSPLWT	J1	V κ 3BACK	K3-5
	A27	QQYGSSPPWT-	J1	V κ 3BACK	K3-6
V κ 6	A26-10	HQSSSLPQT	J1	V κ 1BACK	K2-4
KPG22-2					
V λ 1	DPL3	AAWDDSLDVV	JC3	V λ 1LEA1	L1-3
	DPL5	GTWDSSLSAGV	JC2	V λ 1LEA1	L1-4
	DPL5	GTWDSSLSAGVV	JC3	V λ 1LEA1	L1-6
	DPLS	GTWDSSLSAVV	JC2	V λ 1LEA1	L1-9
	DPL8	QSYDSSLSGVV	JC3	V λ 1LEA1	L1-8
V λ 2	DPL10	CSYAGSSTLV	JC2	V λ 2MIX	L2-4
	DPL11	SSYTSSSTVV	JC2	V λ 2MIX	L2-1
	DPL11	SSYTSSSTLV	JC2	V λ 2MIX	L2-3
	DPL11	CSYTSSSTFV	JC2	V λ 2MIX	L2-7
	DPL12	SSYAGSNNLV	JC3	V λ 2MIX	L2-5
	DAL12	SSYAGSNNFVV	JC3	V λ 2MIX	L2-6
V λ 3	DPL16	NSRDSSGNLV	JC2	V λ 3MIX	L3-1

## Example 48

Preparation of a targeting vector for knocking out antibody genes (heavy-chain and light-chain  $\kappa$  genes) in TT2F ES cells

[0235] It becomes possible to transfer a human chromosome #14 fragment marked with a G418 resistance gene (Example 9) and human chromosome #2 (Example 18) or #22 (Example 35) marked with a puromycin resistance gene into TT2 (or TT2F) cells in which mouse antibody genes (heavy-chain, light-chain  $\kappa$ ) are disrupted. Those chimeric mice which are produced from these human chromosomes #14+#2 or #14+#22-transferred, mouse antibody genes (heavy-chain, light-chain  $\kappa$ )-disrupted TT2 (or TT2F) ES cells according to the method of Example 19 (heavy-chain +  $\kappa$  chain) or Example 36 (heavy-chain +  $\lambda$  chain) are expected to produce antibodies both heavy- and light-chains of which are mainly derived from humans. The abbreviations of the restriction enzymes, etc. appearing in Figures 23-27 are as follows.

Restriction enzymes: Kp: KpnI, B: BamHI, Bg2: BglII, RI: EcoRI, RV: EcoRV, N: Not, S1: Sall, Sca: ScaI, Sfi: SfiI, Sm: SmaI, X: XhoI, (X): XhoI restriction site from  $\lambda$  vector

dK: deletion of KpnI restriction site, dX: deletion of XhoI restriction site,

(Sm/SI): The Sall restriction site was changed to a blunt end and subsequent ligation to a SmaI restriction site,

(S/RV): The Sall restriction site was changed to a blunt end and subsequent ligation to an EcoRV restriction site, Dotted portion: pBluescript SKII(+) or pUC18 plasmid DNA

: LoxP sequence

1. Preparation of plasmid pLoxP-STneo in which LoxP sequence is inserted at both the ends of a G418 resistance gene

[0236] For the deletion of a G418 resistance gene after knocking out an antibody gene of TT2F cells, it is necessary to insert LoxP sequence (Sauer et al., Proc. Natl. Acad. Sci. USA, 85, 5166-, 1988) which is the recognition sequence of Cre recombinase (Sauer et al., supra) at both the ends of the G418 resistance gene (Example 1) in the same direction. Briefly, a G418 resistance cassette (STneo) was cut out from pSTneoB plasmid DNA (Example 1, Katoh et al., Cell Struct. Funct., 12:575, 1987; Japanese Collection of Research Biologicals (JCRB), Deposit Number: VE039) with restriction enzyme XhoI. The DNA fragment was purified by agarose gel electrophoresis and then blunted with T4-DNA polymerase (Blunting End Kit from Takara Shuzo). LoxP sequence-containing plasmid DNA pBS246 (Plasmid pBS246, loxP2 Cassette Vector, U.S. Patent 4959317) was purchased from GIBCO BRL. XhoI linker DNAs were inserted into the EcoRI and SpeI restriction sites of this plasmid to change the sequences of these sites to a XhoI recognition sequence. The STneo DNA fragment described above was inserted into the EcoRV restriction site of the thus modified pBS246 to give plasmid pLoxP-STneo (Fig. 23).

2. Isolation of genomic DNA clones containing C57BL/6-derived antibody heavy-chain  $C\mu$  (IgM constant region) or light-chain  $J\kappa$  -  $C\kappa$  (Ig $\kappa$  joint region and constant region)

[0237] Since TT2 (or TT2F) cells were derived from F1 mice between C56BL/6 mice and CBA mice, the inventors have decided to prepare vectors for antibody gene knockout using genomic DNA clones derived from a C57BL/6 mouse. As a genomic DNA library, an adult C57BL/6N male liver-derived  $\lambda$  DNA library from Clontech was used. As a probe for screening, the following synthetic DNA sequences (60 mers) were used.

Heavy-chain  $C\mu$  probe: 5'-ACC TTC ATC GTC CTC TTC CTC  
CTG AGC CTC TTC TAC AGC ACC ACC GTC ACC CTG TTC AAG-  
3' (SEQ ID NO: 59)

Light-chain  $\kappa$  probe: 5'-TGA TGC TGC ACC AAC TGT ATC CAT  
CTT CCC ACC ATC CAG TGA GCA GTT AAC ATC TGG AGG-3' (SEQ  
ID NO: 60)

[0238] The  $\lambda$  clones were isolated and analyzed to subclone those DNA fragments containing heavy-chain  $C\mu$  or

light-chain J  $\kappa$  - C  $\kappa$  into plasmid pBluescript SKII(+) (Stratagene) (heavy-chain C $\mu$  : Fig. 24; light-chain J  $\kappa$  - C  $\kappa$  : Fig. 25). These DNA fragments were used to prepare targeting vectors for disrupting mouse antibody genes in TT2 (or TT2F) cells as described below.

### 3. Preparation of a vector plasmid for disrupting a mouse antibody heavy-chain gene

[0239] In the C $\mu$  - encoding region in the genomic DNA fragment containing a mouse antibody heavy-chain constant region which was prepared in 2 above, a DNA fragment containing the 2nd to 4th exons (BamHI-XhoI) was replaced with the LoxP-STneo gene prepared in 1 above (Fig. 26). The direction of transcription of STneo was the same as that of the antibody heavy-chain gene. Further, a DT-A cassette A (Proc. Natl. Acad. Sci. USA, 87, 9918-9922, 1990, made by Oriental Yeast Co.) (hereinafter referred to as "DT") modified by changing the Apal and Sall sites to NotI sites was inserted into the NotI site of this plasmid. A plasmid DNA containing a DT gene having the same direction of transcription as that of the heavy-chain gene was selected. This plasmid DNA was amplified using E.coli DH5 and purified by cesium chloride equilibrium centrifugation ("Introduction to Cell Technology Experimental Operations", published by Kodansha, 1992). The purified plasmid DNA was cleaved at one site with restriction enzyme SacII and used for transfection of TT2F ES cells. As a probe for Southern blot analysis of transformant genomic DNA to detect from transformant TT2F ES cells those clones in which homologous recombination has taken place in the antibody heavy-chain portion with the targeting vector, a DNA fragment (about 500 bp) of the switch region located upstream of C $\mu$ -encoding region. This DNA fragment was obtained by amplifying 129 mouse genomic DNAs by PCR under the following conditions.

Sense primer: 5' - CTG GGG TGA GCC GGA TGT TTT G - 3' (SEQ ID NO: 61)

Antisense primer: 5' - CCA ACC CAG CTC AGC CCA GTT C - 3' (SEQ ID NO: 62)

Template DNA: 1  $\mu$ g of EcoRI-digested 129 mouse genomic DNAs

The reaction buffer, deoxynucleotide mix and Taq DNA polymerase used were from Takara Shuzo.

Reaction conditions: 94°C, 3 min, 1 cycle  $\rightarrow$  94°C, 1 min; 55°C, 2 min; 72°C, 2 min; 3 cycles  $\rightarrow$  94°C, 45 sec; 55°C, 1 min; 72°C, 1 min; 36 cycles

[0240] After it was confirmed that amplified DNA fragment can be cleaved at one site with restriction enzyme HindIII as indicated in the Genbank database, this DNA fragment was subcloned into the EcoRV restriction site of plasmid pBluescript. This plasmid DNA (S8) was cleaved with restriction enzymes BamHI and XhoI. A PCR fragment (about 550 bp) was purified by agarose gel electrophoresis to give a probe. Genomic DNA from those TT2F ES cells transformed with the targeting vector was digested with restriction enzymes EcoRI and XhoI, and separated by agarose gel electrophoresis. Then, Southern blotting was performed using the above probe.

### 4. Preparation of a vector for disrupting the mouse antibody light-chain $\kappa$ gene

[0241] The genomic DNA fragment prepared in 2 above contains the J region and constant region of mouse antibody light-chain  $\kappa$ . A DNA fragment (SacII-BglII) containing the C region was replaced with the LoxP-STneo gene having the disrupted KpnI site (Fig. 27). The LoxP-STneo gene was prepared in 1 above. The direction of transcription of STneo was opposite to that of the antibody gene. This plasmid DNA was constructed as follows: the sequence of a multi-cloning site (EcoRI-HindIII) in plasmid pUC18 was changed to the following sequences prepared by DNA chemical synthesis.

5' - AATTCCCGCGGGTCGACGGATCCCTCGAGGGTACCA - 3' (SEQ ID NO: 63)

3' - GGGCGCCCAGCTGCCTAGGGAGCTCCCATGGTTCGA - 5' (S

EQ ID NO: 64)

[0242] The EcoRI-SacII DNA fragment containing J  $\kappa$  (Fig.25) was inserted into the EcoRI and SacII sites of this plasmid. Then, a 3'-end BglII-BglII-BglII-XhoI DNA fragment (Fig.25) was inserted into the BamHI and XhoI sites of the resulting plasmid. A XhoI-Sall DNA fragment containing DT and a XhoI DNA fragment containing LoxP-STneo having the disrupted KpnI fragment were inserted sequentially into the XhoI and Sall sites of this plasmid. The direction of transcription of the DT gene is the same as that of the light-chain  $\kappa$  gene. This plasmid DNA was amplified using E. coli DH5 and purified by cesium chloride equilibrium centrifugation. The purified plasmid DNA was cleaved at one site with restriction enzyme KpnI and used for transfection of TT2F ES cells. As a probe for Southern blot analysis of transformant genomic DNA to detect from transformant TT2F ES cells those clones in which homologous recombination has taken place in the antibody light-chain portion with the targeting vector, a DNA fragment at the 3' end of the light-chain J $\kappa$  - C $\kappa$  genomic DNA fragment (see Fig. 25) (XhoI-EcoRI; about 1.4 kbp) was used. Genomic DNA from those TT2F ES cells transformed with the targeting vector was digested with restriction enzyme EcoRI, and separated by agarose gel electrophoresis. Then, Southern blotting was performed using the above probe.

#### Example 49

##### Production of a mouse ES cell antibody heavy-chain gene-disrupted clone

[0243] In order to obtain a recombinant in which an antibody heavy-chain gene has been disrupted by homologous recombination (hereinafter, referred to as an "antibody heavy-chain homologous recombinant"), the antibody heavy-chain targeting vector prepared in Section 3, Example 48 was linearized with restriction enzyme SacII (Takara Shuzo), and transferred into mouse TT2F ES cells according to the method described by Shinichi Aizawa, "Biomanual Series 8, Gene Targeting", published by Yodosha, 1995. The TT2F cells were treated with trypsin and suspended in HBS at a concentration of  $2.5 \times 10^7$  cells/ml. To the cell suspension, 5  $\mu$ g of DNA was added. Then, electroporation was performed with a gene pulser (Bio-Rad Laboratories, Inc.; resistor unit not connected). A voltage of 250 V was applied at a capacitance of 960  $\mu$ F using an electroporation cell of 4 mm in length at room temperature. The electroporated cells were suspended in 20 ml of an ES medium and inoculated into two tissue culture plastic plates (Corning) of 100 mm into which feeder cells were seeded preliminarily. Similarly, experiments using 10 and 15  $\mu$ g of DNA were also conducted. After one day, the medium was replaced with a medium containing 300  $\mu$ g/ml of G418 (GENETICIN; Sigma). Seven to nine days thereafter, a total of 176 colonies formed were picked up. Each colony was grown up to confluence in a 12-well plate, and then four fifths of the culture was suspended in 0.2 ml of a preservation medium [ES medium + 10% DMSO (Sigma)] and stored frozen at -80°C. The remaining one fifth was inoculated into a 12-well gelatin coated plate and cultured for 2 days. Then, genomic DNA was obtained from the cells ( $10^6$ - $10^7$  cells) with Puregene DNA Isolation Kit (Gentra System Co.). These genomic DNAs from G418 resistant TT2F cells were digested with restriction enzymes EcoRI and XhoI (Takara Shuzo) and separated by agarose gel electrophoresis. Then, Southern blotting was performed to detect homologous recombinants with the probe described in Section 3, Example 48. As a result, 3 clones out of the 176 clones were homologous recombinants. The results of Southern blot analysis of wild-type TT2F cells and homologous recombinants #131 and #141 are shown in the left-side three lanes in Fig. 28. In wild-type TT2F cells, two bands (a and b) are detected which were obtained by the EcoRI and XhoI digestion. In the homologous recombinants, it is expected that one of these bands disappears and that a new band (c) will appear at the lower part of the lane. Actually, band (a) has disappeared in #131 and #141 in Fig. 28 and a new band (c) has appeared. The size of DNA is shown at the left side of the Figure. These results show that one allele of an antibody heavy-chain gene in these recombinant clones has been disrupted by homologous recombination.

#### Example 50

##### Production of chimeric mice from antibody heavy-chain homologous recombinant ES cells

[0244] The cells in a frozen stock of the antibody heavy-chain homologous recombinant TT2F cell clone #131 from Example 49 were thawed, started to culture and injected into 8-cell stage embryos obtained by mating a male and a female mouse of ICR or MCH(ICR) (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. After the embryos were cultured overnight in the medium for ES cells (see Example 9) to develop into blastocysts, about ten of the TT2F cell-injected embryos were transplanted to each side of the uterus of a foster mother ICR mouse (CREA JAPAN, INC.; 2.5 days after pseudopregnant treatment). As a result of transplantation of a total of 94 injected embryos,

22 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2F cell-derived agouti coat color (dark brown) in the host embryo (ICR)-derived albino coat color (white). Out of the 22 offsprings, 18 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells. Out of the 18 mice, 16 mice were female chimeric mice in which more than 80% of their coat color was agouti (i.e. ES cell-derived). From these results, it was confirmed that the antibody heavy-chain homologous recombinant ES cell clone #131 retains the ability to produce chimera. Since a large number of the resultant chimeric mice are female mice exhibiting extremely high contribution, it is very likely that the ES cells have differentiated into functional germ cells (oocytes). Two female chimeric mice exhibiting 100% contribution were mated with MCH(ICR) male mice. As a result, all of the offspring mice exhibited agouti coat color. These offsprings are derived from #131 (see Example 42), and thus it is considered that a disrupted antibody heavy-chain allele was transmitted to them at a rate of 50%.

#### Example 51

Production of a double knockout clone from the antibody heavy-chain homologous recombinant

[0245] It has been reported that a clone in which both alleles are disrupted can be obtained by disrupting one allele by insertion of a G418 resistance gene, culturing an ES cell clone in a medium with an increased G418 concentration and screening the resultant high concentration G418 resistant clones (Shinichi Aizawa, "Biomanual Series 8, Gene Targeting", published by Yodosha, 1995). Based on this technique, the inventors have conducted the following experiments in order to obtain both alleles-disrupted clones from the TT2F antibody heavy-chain homologous recombinants #131 and #141. First, in order to determine the lethal concentration of G418 for both #131 and #141 clones, each clone was inoculated into ten 35 mm plates at a rate of about 100 cells per plate (in this Example, G418 resistant primary culture cells which were not treated with mitomycin were used as feeder cells) (see Example 9). The cells were cultured in an ES medium containing 0, 0.5, 1, 2, 3, 5, 8, 10, 15 and 20 mg/ml of G418 (GENETICIN, Sigma) for 10 days. As a result, definite colonies were observed at a concentration of up to 3 mg/ml, but no colony formation was observed at 5 mg/ml. Based on these results, the minimum lethal concentration was decided to be 5 mg/ml. Then, high concentration G418 resistant clones were selected at concentrations of 4, 5, 6, 7 and 8 mg/ml. For each of #131 and #141, cells were inoculated into ten 100 mm plates at a rate of about  $10^6$  cells per plate and cultured in an ES medium containing G418 at each of the concentrations described above (5 grades; two plates for each concentration). Twelve days after the start of culture, definite colonies (#131: 12 clones; #141: 10 clones) were picked up from plates of 7 mg/ml and 8 mg/ml in G418 concentration. These clones were stored frozen and genomic DNA was prepared by the same procedures as in Example 49. The genomic DNAs from these high concentration G418 resistant clones were digested with restriction enzymes EcoRI and XhoI (Takara Shuzo) and separated by agarose gel electrophoresis. Then, Southern blotting was performed to detect with the probe from Section 3, Example 48 those clones in which both alleles have been disrupted. As a result, one clone derived from #131 (#131-3) was found to be both alleles-disrupted clone. The results of Southern blot analysis of 6 clones derived from #131 are shown in Fig. 28. In wild-type TT2F cells, two wild-type bands (a, b) are detected after the EcoRI and XhoI digestion. In one allele homologous recombinants (#131, #141), the upper band (a) has disappeared and a new band (c) has appeared (Example 49). Furthermore, it is expected that due to the disruption of both alleles, another wild-type band (b) disappears and that the disruption-type band (c) remains alone.

In Fig. 28, this band pattern is observed in clone No. 3 (#131-3). This demonstrates that both alleles of an antibody heavy-chain gene have been disrupted in this clone.

#### Example 52

Removal of a G418 resistance marker gene from the antibody heavy-chain-deficient homozygote TT2F clone

[0246] The G418 resistance marker gene in the antibody heavy-chain both alleles-disrupted clone (high concentration G418 resistant clone #131-3) from Example 51 was removed by the following procedures. An expression vector, pBS185 (BRL), containing Cre recombinase gene which causes a site-directed recombination between the two LoxP sequences inserted at both the ends of the G418 resistance gene was transferred into #131-3 clone according to the methods described in Shinichi Aizawa, "Biomanual Series 8, Gene Targeting", published by Yodosha, 1995 and Seiji Takatsu et al., "Experimental Medicine (extra number): Basic Technologies in Immunological Researches", p. 255-, published by Yodosha, 1995). Briefly, #131-3 cells were treated with trypsin and suspended in HBS to give a concentration of  $2.5 \times 10^7$  cells/ml. To the cell suspension, 30  $\mu$ g of pBS185 DNA was added. Then, electroporation was performed with a gene pulser (Bio-Rad Laboratories, Inc.; resistor unit not connected). A voltage of 250 V was applied at a capacitance of 960  $\mu$ F using an electroporation cell of 4 mm in length (see Example 1). The electroporated cells were suspended in 5 ml of an ES medium and inoculated into a tissue culture plastic plate (Corning) of 60 mm in which

feeder cells were seeded preliminarily. After two days, the cells were treated with trypsin and reinoculated into three 100 mm plates (preliminarily seeded with feeder cells) such that the three plates have 100, 200 and 300 cells, respectively. A similar experiment was also conducted under the same conditions except that the setting of the gene pulser was changed (resistor unit connected; resistance value infinite). After seven days, a total of 96 colonies formed were picked up and treated with trypsin. Then, the colonies were divided into two groups; one was inoculated into a 48-well plate preliminarily seeded with feeder cells and the other was inoculated into a 48-well plate coated with gelatin alone. The latter was cultured in a medium containing 300 µg/ml of G418 (GENETICIN, Sigma) for three days. Then, G418 resistance was judged from the survival ratio. As a result, 6 clones died in the presence of G418. These G418 sensitive clones were grown to confluence in 35 mm plates, and four fifths of the resultant culture was suspended in 0.5 ml of a preservation medium [ES medium + 10% DMSO (Sigma)] and stored frozen at -80°C. The remaining one fifth was inoculated into a 12-well gelatin coated plate and cultured for two days. Thereafter, genomic DNA was prepared by the same procedures as in Example 2. These genomic DNAs from G418 sensitive TT2F clones were digested with restriction enzyme EcoRI (Takara Shuzo) and separated by agarose gel electrophoresis. Then, Southern blotting was performed to confirm the removal of the G418 resistance gene using a 3.2 kb XhoI fragment (Probe A) from G418 resistance gene-containing pSTneoB. As a result, bands observed in #131-3 clone which hybridize with Probe A were not detected at all in the sensitive clones. From these results, it was confirmed that the G418 resistance marker gene had been surely removed in the G418 sensitive clones obtained. Additionally, as a result of Southern blot analysis performed in the same manner using Probe B obtained by digesting pBS185 DNA with EcoRI, no specific band which hybridizes with Probe B was detected in these G418 sensitive clones. Thus, it is believed that Cre recombinase-containing pBS185 is not inserted into the chromosomes of the sensitive clones. In other words, these sensitive clones can be transformed with the vector for knocking out an antibody light-chain (vector having a loxP sequence at both the ends of a G418 resistance gene) described in Section 4, Example 48.

#### Example 53

Transfer of human chromosome #14 (containing antibody heavy-chain gene) into the antibody heavy-chain-deficient ES cell clone

**[0247]** Human chromosome #14 (containing an antibody heavy-chain gene) marked with a G418 resistance gene is transferred by microcell fusion as described in Example 9 into the mouse ES cell clone (from TT2F, G418 sensitive) obtained in Example 52 which is deficient in an endogenous antibody heavy-chain. In the resultant G418 resistant clone, the retention of human chromosome #14 (fragment) containing a human antibody heavy-chain gene is confirmed by PCR analysis or the like (see Example 9).

#### Example 54

Transfer of human chromosome #2 fragment or human chromosome #22 into the antibody heavy-chain-deficient ES cell clone retaining human chromosome #14 (fragment)

**[0248]** A human chromosome #2 fragment (containing the antibody heavy-chain  $\kappa$  gene) or human chromosome #22 (containing the antibody heavy-chain  $\lambda$  gene) marked with a puromycin resistance gene is transferred into the antibody heavy-chain-deficient mouse ES cell clone retaining a human chromosome #14 partial fragment (G418 resistant) from Example 53 by microcell fusion as described in Examples 18 and 35. In the resultant puromycin and G418 double drug-resistant clone, the retention of the human chromosome #14 (fragment) and human chromosome #2 fragment or #22 (fragment) is confirmed by PCR analysis or the like (see Examples 18 and 35).

#### Example 55

Production of chimeric mice from the endogenous antibody heavy-chain-deficient mouse ES cells retaining human chromosome #14 (fragment) containing a human antibody heavy-chain gene

**[0249]** Chimeric mice from the endogenous antibody heavy-chain gene-deficient mouse ES cell clone obtained in Example 53 retaining human chromosome #14 (fragment) containing a human antibody heavy-chain gene are produced by the same procedures as in Example 10. In the resultant chimeric mice, a human antibody heavy-chain produced in the ES cell clone-derived B cells is detected by the method described in Example 14. Since antibody heavy-chain genes functional in the ES cell clone-derived B cells are only the human-derived gene on the transferred chromosome, many of the ES cell clone-derived B cells produce human antibody heavy-chain.

## Example 56

Production of chimeric mice from the endogenous antibody heavy-chain-deficient mouse ES cells retaining human chromosomes #14+#2 (fragments) or #14+#22 (fragments)

[0250] Chimeric mice are produced by the same procedures as in Examples 19, 36, etc from the endogenous antibody heavy-chain gene-deficient mouse ES cell clone retaining human chromosomes #14+#2 (fragments) or #14+#22 (fragments) obtained in Example 54. In the resultant chimeric mice, human antibody heavy-chain and light-chain  $\kappa$  or  $\lambda$  are detected in the ES cell clone-derived B cells according to the method described in Examples 14, 23 and 32. As in Example 55, antibody heavy-chain genes functional in the ES cell clone-derived B cells are only the human-derived gene on the transferred chromosome. Thus, many of the ES cell clone-derived B cells produce human heavy-chains. Furthermore, complete human antibody molecules both heavy and light-chains which are derived from humans are also detected by the method described in Examples 37 and 38.

## Example 57

Production of human antibody-producing hybridomas from the chimeric mice derived from the endogenous antibody heavy-chain-deficient mouse ES cells retaining human chromosomes #14+#2 (fragments) or #14+#22 (fragments)

[0251] The chimeric mice from Example 56 are immunized with an antigen of interest in the same manner as in Examples 15, 25 and 34. The spleen is isolated from each mice and the spleen cells are fused with myeloma cells to produce hybridomas. After cultivation for 1-3 weeks, the culture supernatant is analyzed by ELISA. The ELISA is performed by the method described in Examples 14, 15, 21, 24, 25, 33, 34, 37 and 38. As a result, human antibody positive clones and clones which are human antibody positive and specific to the antigen used in the immunization are obtained.

## Example 58

Production of an antibody light-chain gene-disrupted clone from the antibody heavy-chain-deficient homozygote mouse ES cells

[0252] A homologous recombinant, which has further disruption in an antibody light-chain gene in the antibody heavy-chain-deficient homozygote TT2F cell clone (G418 sensitive) obtained in Example 52 is produced by the following procedures. Briefly, the antibody light-chain targeting vector prepared in Section 4, Example 48 is linearized with restriction enzyme KpnI (Takara Shuzo), and transferred into the above TT2F cell clone (G418 sensitive) according to the method described in Shinichi Aizawa, "Biomannual Series 8: Gene Targeting", published by Yodosha, 1995. After 7-9 days, colonies formed are picked up. They are stored frozen and genomic DNA is prepared in the same manner as in Example 49. Genomic DNAs from G418 resistant clones are digested with restriction enzymes EcoRI and NotI (Takara Shuzo) and separated by agarose gel electrophoresis. Then, Southern blot analysis is performed to detect homologous recombinants with the probe described in Section 4, Example 48.

## Example 59

Production of an double knockout clone from the antibody light-chain homologous recombinant

[0253] A clone in which both alleles of a light-chain gene are disrupted is prepared from the TT2F antibody light-chain homologous recombinant (and antibody heavy-chain-deficient homozygote) clone from Example 58 by the procedures described below. Briefly, a high concentration G418 resistant clone is prepared and stored frozen, and DNA is prepared in the same manner as in Example 51. Genomic DNA from the high concentration G418 resistant clone is digested with restriction enzymes EcoRI and NotI (Takara Shuzo) and separated by agarose gel electrophoresis. Then, Southern blot analysis is performed to detect those clones in which both alleles have been disrupted, with the probe from Section 4, Example 48.

## Example 60

Removal of the G418 resistance gene from the antibody light-chain-deficient homozygote (antibody heavy-chain-deficient homozygote) TT2F cell clone

[0254] The G418 resistance marker gene in the antibody light-chain both alleles-disrupted clone (high concentration G418 resistant clone) obtained in Example 59 is removed by the same procedures as in Example 52. Briefly, an expression vector, pBS185 (BRL), containing Cre recombinase gene which causes a site-directed recombination between the two loxP sequences inserted at both the ends of the G418 resistance gene (Section 1, Example 48) was transferred into the above clone according to the method described in Example 52. The resultant G418 sensitive clones are grown to confluence in 35 mm plates, and 4/5 of the resultant culture was suspended in 0.5 ml of a preservation medium [ES medium + 10% DMSO (Sigma)] and stored frozen at -80°C by the same procedures as in Example 52. The remaining 1/5 was inoculated into a 12-well gelatin coated plate. After cultivation for two days, genomic DNA is prepared by the method described in Example 2. These genomic DNAs from G418 sensitive TT2F clones are digested with restriction enzyme EcoRI (Takara Shuzo) and separated by agarose gel electrophoresis. Then, Southern blotting is performed to confirm the removal of the G418 resistance gene using a 3.2 kb XhoI fragment from G418 resistance gene-containing pSTneoB as a probe.

## Example 61

(1) Transfer of a human chromosome #14 fragment (containing antibody heavy-chain gene) into the endogenous antibody heavy-chain and  $\kappa$  chain-deficient ES cell clone

[0255] A human chromosome #14 fragment SC20 (containing a human antibody heavy-chain gene) was transferred by microcell fusion as described in Section 2 of Example 68 into the mouse ES cell clone HKD31 (from TT2F, G418 sensitive, puromycin sensitive) obtained in Example 78 which is deficient in both endogenous antibody heavy-chain and  $\kappa$  chain. The microcell fusion and the selection of G418 resistant clones were performed in the same manner as in Example 2. Eight of the resultant G418 resistant clones were subjected to PCR analysis using IgM and D14S543 primers (see Example 68). As a result, both markers were detected in 8 out of the 7 clones analyzed. Hence, it was confirmed that the antibody heavy-chain and  $\kappa$  chain-deficient ES cell clone retains the human chromosome #14 fragment SC20.

(2) Production of chimeric mice from the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cells retaining a human chromosome #14 fragment (containing antibody heavy-chain gene)

[0256] Chimeric mice were produced by the same procedures as in Example 10, etc. from the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cell clone HKD31-8 which was obtained in Section 1 of Example 61 and which retains a human chromosome #14 fragment (containing a human antibody heavy-chain gene). As a result of transplantation of a total of 188 injected embryos, 25 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo (ICR) - derived albino coat color (white). Out of the 25 offsprings, 17 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells. Out of the 17 mice, three were chimeric mice in which more than 95% of their coat color was (ES cell-derived) agouti.

[0257] From these results, it was confirmed that the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cell clone retaining the human chromosome #14 fragment (containing a human antibody heavy-chain gene) maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mice.

(3) Detection of human antibody (having human  $\mu$ ,  $\gamma$  or  $\alpha$  chain) in sera of the chimeric mice derived from the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cells retaining a human chromosome #14 fragment (containing antibody heavy-chain gene)

[0258] The chimeric mice produced in Section 2 of Example 61 (derived from HKD31-8) were bled 12 weeks (#1) or 7 weeks (#2-4) after birth. The human antibody concentration in the sera was determined by ELISA in the same manner as in Example 14. Ninety six-well microtiter plates were coated with PBS-diluted anti-human immunoglobulin  $\mu$  chain antibody (Sigma, I6385) or anti-human immunoglobulin  $\gamma$  chain antibody (Sigma, I3382) or anti-human immunoglobulin  $\alpha$  chain antibody (Pharmingen, 08091D) and then a serum sample diluted with mouse serum (Sigma, M5905)-containing PBS was added. Subsequently, peroxidase-labeled anti-human immunoglobulin  $\mu$  chain antibody (The Binding Site Limited, MP008) or peroxidase-labeled anti-human immunoglobulin  $\gamma$  chain antibody (Sigma, A0170) was added to



the plates and incubated. Alternatively, biotin-labeled anti-human immunoglobulin  $\alpha$  chain antibody (Pharmingen, 08092D) was added to the plates and incubated. After the plates were washed, an avidin-peroxidase complex (Vector, ABC Kit PK4000) was added thereto and incubated. TMBZ (Sumitomo Bakelite, ML-1120T) was added as a peroxidase substrate and then enzyme activity was determined by absorbance measurement at 450 nm.

Purified human immunoglobulins IgM (CAPPEL, 6001-1590), IgG (Sigma, I4506) and IgA (Sigma, I2636) of known concentrations having  $\mu$  chain,  $\gamma$  chain and  $\alpha$  chain, respectively, were used as standards for determining human antibody concentrations in the sera. These standards were diluted stepwise with mouse serum-supplemented PBS. The results are shown in Table 20. Chimeric mice having concentrations of human antibody  $\mu$  and  $\lambda$  chains almost as high as in normal mouse sera were confirmed. Also, chimeric mice expressing human  $\alpha$  chain were confirmed. Further, human immunoglobulin  $\gamma$  chain sub-classes were detected in the same manner as in Example 29. As a result, all of the four subclasses ( $\gamma$  1,  $\gamma$  2,  $\gamma$  3 and  $\gamma$  4) were detected.

[0259] These results show that a human antibody heavy-chain gene is expressed efficiently in the chimeric mice derived from the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cells retaining the human chromosome #14 fragment (containing an antibody heavy-chain gene); it was also shown that not only  $\mu$  chain but also all of the  $\gamma$  chain subclasses and  $\alpha$  chain were expressed therein as a result of class switching.

Table 20.

Human Antibody Concentrations in Chimeric Mice (ELISA)				
Chimeric Mouse	Chimerism %	Human Antibody (mg/l)		
		IgM	IgG	IgA
#1	90	270	1250	0.46
#2	99	370	820	0.23
#3	99	550	1460	0.32
#4	95	340	2300	0.06

(4) Acquisition of hybridomas producing anti-HSA antibody comprising human  $\gamma$  chain from the chimeric mice derived from the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cells retaining a human chromosome #14 fragment (containing antibody heavy-chain gene)

[0260] Chimeric mice #3 (derived from HKD31-8; chimerism 99%) and #4 (chimerism 95%) which had exhibited a high human antibody  $\gamma$  chain concentration in the serum in Section 3 of Example 61 were immunized as described below. Human serum albumin (HSA, Sigma, A3782) dissolved in PBS was mixed with an adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc.) to prepare an HSA solution with a concentration of 0.25 mg/ml. When the above-described chimeric mice became 16-week old, 0.2 ml of this HSA solution was administered intraperitoneally twice at an interval of 2 weeks. Two weeks thereafter, the mice were immunized with human serum albumin dissolved in PBS and then bled. The concentration of anti-HSA human antibody in the sera was determined by ELISA in the same manner as in Example 14. Briefly, ELISA plates were coated with HSA and then peroxidase-labeled anti-human Ig $\mu$  antibody (The Binding Site, MP008), anti-human Ig $\gamma$  antibody (Sigma, A1070) and anti-human Ig $\alpha$  antibody (Kirkegaard & Perry Laboratories Inc., 14-10-01) were used for detection. The results are shown Fig. 33. Hybridomas were produced using a myeloma cell SP - 2/0 - Ag14 (Dainippon Pharmaceutical Co., Ltd.) by the method described in Ando, "Monoclonal Antibody Experiment Procedure Manual", published by Kodansha Scientific in 1991. Three days after the final immunization, the spleen was removed from the chimeric mice and then cell fusion was performed using PEG in the same manner as in Example 29 to prepare hybridomas. At the same time, blood samples were collected from the mice to quantitate human Ig $\gamma$  subclasses in the sera. As a result, 920 mg/l of  $\gamma$  1, 520 mg/l of  $\gamma$  2, 11 mg/l of  $\gamma$  3 and 140 mg/l of  $\gamma$  4 were detected in the serum of chimeric mouse #3.

[0261] The fused cells were diluted with a medium (Sanko Pure Chemical, S Cloning Medium CM-B) containing 5% HCF (Air Brown) and HAT (Dainippon Pharmaceutical Co., Ltd., No. 16-808-49) or 1 mg/ml of G418 to give a concentration of  $10^6$  spleen cells/ml and then dispensed into 96-well plates (100  $\mu$ l/well), followed by cultivation. At day 8 of the cultivation, the culture supernatant was collected and screened for human antibody-producing hybridomas by ELISA in the same manner as in Example 14. Briefly, ELISA plates were coated with a HSA solution dissolved in CBB buffer to give a concentration of 5  $\mu$ g/ml. Peroxidase-labeled anti-human immunoglobulin  $\gamma$  chain antibody (Sigma, A0170) and TMBZ (Sumitomo Bakelite, ML-1120T) were used for detection. An absorbance about 3 times higher than the absorbance in the negative control was used as a criterion for judgement. As a result, 74 positive wells were obtained from chimeric mouse #3 and 29 positive wells from chimeric mouse #4. Also, anti-HSA antibody having human  $\mu$  chain was screened in HSA-solution-coated plates using peroxidase-labeled anti-human immunoglobulin  $\mu$  chain antibody (Tago, #2392). Briefly, fused cells from chimeric mouse #3 were inoculated into fifteen 96-well plates, from

which 4 plates were selected by G418 resistance. The culture supernatants of these 4 plates were screened to obtain 5 positive wells. Wells which exhibited colony formation after selection with HAT or 1 mg/ml of G418 were 74 wells/plate for HAT and 29 wells/plate for G418. The cells of those wells which were positive for human  $\gamma$  chain-containing anti-HSA antibody and which had a relatively large number of cells were transferred into 46-well plates and cultured for another 4 days. The isotype of the antibody in the culture supernatant was determined by ELISA. ELISA was performed in HSA-coated plates using alkali phosphatase-labeled anti-human IgG1 antibody (Zymed Laboratories, Inc., 05-3322), anti-human IgG2 antibody (Zymed Laboratories, Inc., 05-3522), anti-human IgG3 antibody (Zymed Laboratories, Inc., 05-3622) and anti-human IgG4 antibody (Zymed Laboratories, Inc., 05-3822) in the same manner as in Example 14. As a result, 27 human IgG1 positive clones, 11 human IgG2 positive clones, 2 human IgG3 positive clones and 13 human IgG4 positive clones were obtained. Fused cells from chimeric mouse #4 were treated in the same manner to obtain 4 positive clones with a large number of cells as human IgG1 producing clones.

[0262] These results show that the immunization by human protein (HSA) of the chimeric mice derived from the endogenous antibody heavy-chain & light-chain-deficient mouse ES cells retaining the human chromosome #14 partial fragment containing a human antibody heavy-chain gene increases the antibody titers of antigen specific human Ig $\mu$ ,  $\gamma$  and  $\alpha$  to thereby enable the acquisition of hybridomas producing anti-HSA antibody containing  $\mu$  chain and all of the human  $\gamma$  chain subclasses.

#### Example 62

Transfer of human chromosome #2 (containing light-chain  $\kappa$  gene) into the endogenous antibody heavy-chain and  $\kappa$  chain-deficient ES cells retaining a human chromosome #14 fragment (containing antibody heavy-chain gene)

[0263] A human chromosome #2 fragment (containing antibody light-chain  $\kappa$  gene) marked with a puromycin resistance gene was transferred into the endogenous antibody heavy-chain and  $\kappa$  chain-deficient mouse ES cell clone HKD31-8 obtained in Section 1 of Example 61 and which retained a human chromosome #14 fragment (containing an antibody heavy-chain gene). The method of transfer was by microcell fusion as described in Example 18. As a result, 13 puromycin and G418 double-resistant clones were obtained. These clones were subjected to PCR analysis (see Example 18) using IgM and D14S543 primers (see Example 68) for the chromosome #14 fragment and V  $\kappa$  1 and FABP1 primers (see Example 12) for the chromosome #2 fragment. As a result, the presence of all the 4 markers was confirmed in 8 clones. Of these clones, KH13 clone was subjected to FISH analysis using human chromosome-specific probes (see Examples 9 and 12). The results are shown in Fig. 34. Two independent, small chromosome fragments hybridizing to the probes were observed in KH13. These results show that KH13 retains both the chromosome #14 fragment and the chromosome #2 fragment.

#### Example 63

Transfer of human chromosome #22 (containing light-chain  $\lambda$  gene) into the endogenous antibody heavy-chain and  $\kappa$  chain-deficient ES cells retaining a human chromosome #14 fragment (containing antibody heavy-chain gene)

[0264] Human chromosome #22 (containing antibody light-chain  $\lambda$  gene) marked with a puromycin resistance gene was transferred into mouse ES cell clone HKD31-8 obtained in Section 1 of Example 61 which was deficient in the endogenous antibody heavy-chain &  $\kappa$  chain and which retained a human chromosome #14 fragment (containing an antibody heavy-chain gene). The method of transfer was by microcell fusion as described in Example 35. As a result, 12 puromycin and G418 double drug-resistant clones were obtained. These clones were subjected to PCR analysis (see Example 35) using IgM and D14S543 primers for the chromosome #14 fragment and Ig  $\lambda$ , D22S315, D22S275, D22S278, D22S272 and D22S274 primers (see Example 2) for the chromosome #22 fragment. As a result, the presence of all of the 8 markers was confirmed in 10 clones. Of the remaining 2 clones, LH13 clone exhibited the presence of 5 markers, IgM, D14S543, IgI, D22S275 and D22S274. Thus, it is believed that this clone contains a fragment of human chromosome #22. LH13 was further subjected to FISH analysis using a human chromosome #22-specific probe and a human chromosome #14-specific probe separately. As a result, independent chromosome fragments hybridizing to the respective probes were observed. This indicates that this clone retains both a chromosome #14 fragment and a chromosome #2 fragment.

## Example 64

Production of endogenous antibody heavy-chain & light-chain-deficient mouse ES cells retaining three human chromosomes, #2 (containing antibody light-chain  $\kappa$  gene), #14 (containing antibody heavy-chain gene) and #22 (containing antibody  $\lambda$  chain gene), or partial fragments thereof

[0265] In order to obtain mouse ES cells retaining three kinds of human chromosomes, human chromosome #2 or #22 is marked by inserting a marker gene such as blasticidin resistance (Izumi et al., Exp. Cell. Res., 197:229, 1991), hygromycin resistance (Wind et al., Cell, 82:321-, 1995), etc. This marking is performed according to the method described in Examples 16 and 26. Human chromosome #22 (containing human antibody light-chain  $\lambda$  gene) marked with blasticidin resistance, hygromycin resistance, etc. is transferred into the mouse ES cell clone (from TT2F, G418 resistant, puromycin resistant) obtained in Example 62 which is deficient in endogenous antibody heavy-chain & light-chain and which retains both human chromosome #14 (fragment) and human chromosome #2 (partial fragment). The method of transfer is by the method described in Example 9. As feeder cells for culturing ES cells, appropriate cells are selected depending on the selection marker used. When a hygromycin resistance marker is used, primary culture fibroblasts obtained from a transgenic mouse strain which retains and expresses the marker (Johnson et al., Nucleic Acids Research, vol. 23, No. 7, 1273-, 1995) are used. It is confirmed by PCR analysis, etc. (see Examples 9, 18 and 35) that the resultant G418, puromycin and hygromycin (or blasticidin) triple drug-resistant clones retain the three kinds of human chromosomes (fragments) described above. In the same manner, a human chromosome #2 fragment marked with a hygromycin or blasticidin resistance gene is transferred into the mouse ES cell clone (from TT2F, G418 resistant, puromycin resistant) obtained in Example 63 which is deficient in endogenous antibody heavy-chain & light-chain and which retains both human chromosome #14 (fragment) and human chromosome #22 (fragment).

## Example 65

Production of chimeric mice from the endogenous antibody heavy-chain & light-chain genes-disrupted mouse ES cells retaining a plurality of human chromosomes (fragments) containing human antibody heavy-chain gene and light-chain gene, respectively

[0266] Chimeric mice are produced by the same procedures as in Example 10, etc. from the endogenous antibody heavy-chain & light-chain genes-disrupted mouse ES cell clones that retain human chromosomes (fragments) containing human antibody genes and which were obtained in Examples 61, 62, 63 and 64. In the resultant chimeric mice, mouse antibodies produced in host embryo-derived B cells and human antibodies produced mainly in ES cell clone-derived B cells are detected by the method described in Examples 14, 23 and 32. Since the antibody heavy-chain gene and the light-chain  $\kappa$  gene which are both functional in the ES cell clone-derived B cells are only human-derived genes on the transferred chromosomes, many of the ES cell clone-derived B cells produce human antibody heavy-chain and light-chain  $\kappa$  (Lonberg et al., Nature, 368:856-, 1994). Furthermore, complete human antibody molecules in which both heavy- and light-chains are derived from human are also detected by the method described in Examples 37 and 38.

(1) Production of chimeric mice from the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cells retaining both a human chromosome #14 fragment (containing antibody heavy-chain gene) and a human chromosome #2 fragment (containing antibody light-chain  $\kappa$  gene)

[0267] Chimeric mice were produced by the same procedures as in Example 10, etc. from mouse ES cell clone KH13 obtained in Example 62 which is deficient in the endogenous antibody heavy-chain and  $\kappa$  chain genes and which retains both a human chromosome #14 fragment (containing an antibody heavy-chain gene) and a human chromosome #2 fragment (containing light-chain  $\kappa$  gene). As a result of transplantation of a total of 176 injected embryos, 20 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo (ICR)-derived albino coat color (white). Out of the 20 offsprings, 7 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells.

[0268] From these results, it was confirmed that the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cell clone retaining both a human chromosome #14 fragment (containing an antibody heavy-chain gene) and a human chromosome #2 fragment (containing light-chain  $\kappa$  gene) maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mice.

(2) Production of chimeric mice from the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cells retaining both a human chromosome #14 fragment (containing antibody heavy-chain gene) and a human chromosome #22 fragment (containing light-chain  $\lambda$  gene)

[0269] Chimeric mice were produced by the same procedures "as in Example 10, etc. from mouse ES cell clone LH13 obtained in Example 63 which is deficient in the endogenous antibody heavy-chain and  $\kappa$  chain genes and which retains both a human chromosome #14 fragment (containing an antibody heavy-chain gene) and a human chromosome #22 fragment (containing light-chain  $\lambda$  gene). As a result of transplantation of a total of 114 injected embryos, 22 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo (ICR)-derived albino coat color (white). Out of the 22 offsprings, 5 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells.

[0270] From these results, it was confirmed that the endogenous antibody heavy-chain and  $\kappa$  chain genes-disrupted mouse ES cell clone retaining both a human chromosome #14 fragment (containing an antibody heavy-chain gene) and a human chromosome #22 fragment (containing light-chain  $\lambda$  gene) maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mice.

(3) Detection and quantitative determination of complete human antibody in sera of the chimeric mice derived from the endogenous antibody heavy-chain and  $\kappa$  chain-deficient mouse ES cells retaining both a human chromosome #2 partial fragment and a human chromosome #14 partial fragment

[0271] The chimeric mice (derived from KH13) produced in Section 1 of Example 65 were bled at day 40 after birth. The concentrations of human antibody in the sera were determined by ELISA in the same manner as in Example 14. Briefly, ELISA plates were coated with PBS-diluted anti-human immunoglobulin  $\kappa$  chain antibody (Kirkegaard & Perry Laboratories Inc., 01-10-10) or anti-human immunoglobulin  $\kappa$  chain antibody (Vector, AI-3060) and then serum samples diluted with mouse serum (Sigma, M5905)-supplemented PBS were added. Subsequently, peroxidase-labeled anti-human immunoglobulin  $\mu$  chain antibody (The Binding Site Limited, MP008) or peroxidase-labeled anti-human immunoglobulin  $\gamma$  chain antibody (Sigma, A0170) was added and incubated. TMBZ (Sumitomo Bakelite, ML-1120T) was added as a peroxidase substrate and then enzyme activity was determined by absorbance measurement at 450 nm. Purified human immunoglobulins IgM (Caltag, 13000) and IgG (Sigma, I4506) of known concentrations having  $\mu$  chain and  $\kappa$  chain were used as standards for determining human antibody concentrations in the sera by comparison. These standards were diluted stepwise with mouse serum-supplemented PBS. The results are shown in Table 21. Chimeric mice were confirmed that had concentrations of complete human antibody more than 10 times higher than in chimeric mice derived from ES cells whose endogenous antibody genes were not knocked out. Also, complete human antibody containing human  $\gamma$  chain was confirmed in the sera of the chimeric mice.

[0272] From these results, it was confirmed that the concentration of complete human antibody in which both heavy- and light-chains were derived from human increased in the chimeric mice derived from the endogenous antibody heavy-chain and  $\kappa$  chain-deficient mouse ES cells retaining both a human chromosome #14 partial fragment and a human chromosome #22 partial fragment.

Table 21.

Concentrations of Human Antibodies in Chimeric Mice (ELISA)				
ES clone	Chimeric mouse	Chimerism (%)	IgM, $\kappa$ (mg/l)	IgG, $\kappa$ (mg/l)
KH13	CKH13-1	95	0.1	0.07
KH13	CKH13-2	85	0.9	0.13

(4) Detection and quantitative determination of complete human antibody in sera of the chimeric mice derived from the endogenous antibody heavy-chain and  $\kappa$  chain-deficient mouse ES cells retaining both a human chromosome #14 partial fragment and a human chromosome #22 partial fragment

[0273] The chimeric mice (derived from KH13) produced in Section 2 of Example 65 were bled at day 49 after birth. The concentrations of human antibody in the sera were determined by ELISA in the same manner as in Example 14. Briefly, ELISA plates were coated with PBS-diluted anti-human immunoglobulin  $\lambda$  chain antibody (Kirkegaard & Perry Laboratories Inc., 01-10-11) or anti-human immunoglobulin  $\lambda$  chain antibody (Vector, AI-3070) and then serum samples diluted with mouse serum (Sigma, M5905)-supplemented PBS were added. Subsequently, peroxidase-labeled anti-human immunoglobulin  $\mu$  chain antibody (The Binding Site Limited, MP008) or peroxidase-labeled anti-human immunoglobulin  $\gamma$  chain antibody (Sigma, A0170) was added and incubated. TMBZ (Sumitomo Bakelite, ML-1120T) was

added as a peroxidase substrate and then enzyme activity was determined by absorbance measurement at 450 nm. Purified human immunoglobulins IgM (Caltag, 13000) and IgG (Sigma, 14506) of known concentrations having  $\mu$  chain and  $\kappa$  chain were used as standards for determining human antibody concentrations in the sera by comparison. These standards were diluted stepwise with mouse serum-supplemented PBS. The results are shown in Table 22. Chimeric mice individuals were confirmed that had concentrations of complete human antibody about 40 times higher than in chimeric mice derived from ES cells whose endogenous antibody genes were not knocked out. Also, complete human antibody containing human  $\gamma$  chain was confirmed in the sera of the chimeric mice.

[0274] From these results, it was confirmed that the concentration of complete human antibody in which both heavy- and light-chains were derived from human increased in the chimeric mice derived from the endogenous antibody heavy-chain and  $\kappa$  chain-deficient mouse ES cells retaining both a human chromosome #14 partial fragment and a human chromosome #22 partial fragment.

Table 22.

Concentrations of Human Antibodies in Chimeric Mice (ELISA)				
ES clone	Chimeric mouse	Chimerism (%)	IgM, $\lambda$ (mg/l)	IgG, $\lambda$ (mg/l)
LH13	CLH13-1	95	13	2.6
LH13	CLH13-2	90	2.8	0.36

#### Example 66

Production of complete human antibody-producing hybridomas from chimeric mice prepared by transferring the endogenous antibody heavy-chain and light-chain-deficient mouse ES cells retaining both a human chromosome #14 partial fragment and a human chromosome #22 partial fragment into immunodeficient mouse host embryos

[0275] A chimeric mouse CLH13-3 (derived from TT2FES clone LH13; chimerism 35%) obtained in Section 3 of Example 67 was immunized with HSA from day 43 after birth. Briefly, human serum albumin (HSA, Sigma, A3782) dissolved in PBS was mixed with an adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc.) to prepare a HSA solution with a concentration of 0.25 mg/ml, 0.2 ml of which was administered intraperitoneally twice at an interval of 1 week. One week thereafter, the mouse was immunized with human serum albumin dissolved in PBS. The mouse was bled every 1 week to determine the concentrations of anti-HSA human antibodies in the serum by ELISA in the same manner as in Example 14. The results are shown in Fig. 35. The spleen was removed from the chimeric mouse at day 3 after the final immunization and then cell fusion was performed using PEG in the same manner as in Example 24 to prepare hybridomas. Briefly, the fused cells were diluted with a medium (Sanko Pure Chemical, S Cloning Medium CM-B) containing HAT (Dainippon Pharmaceutical Co., Ltd., No. 16-808-49) or 1 mg/ml of G418 to give a concentration of  $10^6$  spleen cells/ml and then dispensed into 96-well plates (100  $\mu$ l/well), followed by cultivation. Both of the selection media contained 5% HCF (Air Brown). At day 6 of the cultivation, colonies were formed in almost all wells in both the G418 selection and HAT selection plates. A total of about 770 hybridoma-positive wells were obtained. The culture supernatants were collected and subjected to screening for human antibody-producing hybridomas by ELISA in the same manner as in Example 14. Briefly, ELISA plates were coated with anti-human immunoglobulin  $\lambda$  chain antibody (Vector, AI-3070). Biotin-labeled anti-human immunoglobulin  $\lambda$  chain antibody (Vector, BA-3070) and an avidin-peroxidase complex (Vector ABC Kit PK4000) were used for detection with TMBZ (Sumitomo Bakelite, ML-1120T) used as a substrate. An absorbance about 2 times higher than the absorbance in the negative control was used as a criterion for judgement. As a result, 17 positive wells were obtained. The cells of the positive wells were transferred into 24-well plates and cultured in IMDM medium containing 10% FBS. The culture supernatants were analyzed by ELISA in the same manner as in Section 4 of Example 65. As a result, the presence of 0.09-11 mg/ml of complete human antibody having both human Ig $\mu$  & Ig $\lambda$  was confirmed in 16 wells. The antibody titer of anti-HSA human  $\lambda$  chain was determined in the same manner as in Example 33 to obtain one positive well. The cells of the well which was complete human antibody-positive and anti-HSA human  $\lambda$  chain-positive were cloned by limiting dilution according to the method described in Ando, "Monoclonal Antibody Experiment Procedure Manual", published by Kodansha Scientific in 1991. As a result, 2 clones of anti-HSA human  $\lambda$  chain-positive hybridomas were obtained.

[0276] From these results, it was confirmed that complete human antibody-producing hybridomas could be obtained from chimeric mice prepared by transferring the endogenous antibody heavy-chain and light-chain-deficient mouse ES cells retaining both a human chromosome #14 partial fragment and a human chromosome #22 partial fragment into immunodeficient mouse host embryos. Furthermore, it was confirmed that the antibody titers of antigen-specific human Ig $\mu$  and Ig $\lambda$  increased in response to the stimulation of the HSA antigen. It was further confirmed that hybridomas producing a HSA-specific antibody consisting of human Ig $\mu$  and Ig $\lambda$  could be obtained from this chimeric mouse.

[0277] Since the fused cells had a drug resistance marker on their chromosome, it was possible to select hybridomas using G418 without adding HAT. After G418 selection, only those cells having a human chromosome grow and, thus, hybridomas can be obtained selectively. Also, it is expected that a human chromosome can be prevented from falling off fused cells.

Furthermore, it is expected that even myeloma cells unsuitable for HAT selection such as those having HGPRT (hypoxanthine-guanine-phosphoribosyltransferase) enzyme may become available for cell fusion.

#### Example 67

#### Production of chimeric mice with heavy-chain gene-disrupted host embryos

[0278] From those mice exhibiting agouti coat color among the progeny of the endogenous antibody heavy-chain one allele-disrupted TT2F cell clone - derived chimeric mice produced in Example 49, mice retaining the disrupted allele are selected by Southern blot analysis (Example 49) or the like (the expected possibility is 1/2). Offsprings born by the mating of those antibody heavy-chain-deficient heterozygous male and female mice are subjected to Southern blot analysis (see Example 49), analysis of the production of antibody heavy-chains in sera (Kitamura et al., Nature, 350:423-, 1991), etc. Thus, antibody heavy-chain-deficient homozygotes can be obtained which are deficient in both alleles and which can hardly produce functional antibodies of their own (the expected possibility is 1/4; for the results in membrane-type  $\mu$  chain-deficient mice, see Kitamura et al., Nature, 350:423-, 1991).

#### (1) Establishment of an antibody heavy-chain knockout mouse strain

[0279] Those mice that exhibited agouti coat color among the progeny of the endogenous antibody heavy-chain one allele-disrupted TT2F cell clone-derived chimeric mice produced in Example 49 were subjected to Southern blot analysis (Example 49) to select those mice that retained the disrupted allele. Offsprings born by the mating of these antibody heavy-chain-deficient heterozygous male and female mice were subjected to Southern blot analysis (see Example 49) and analysis of the production of antibody  $\mu$  chain in sera (Kitamura et al., Nature, 350:423-, 1991), etc. As a result, antibody heavy-chain-deficient homozygotes could be obtained which were deficient in both alleles and which could hardly produce functional antibodies of their own (for the results in membrane-type  $\mu$  chain-deficient mice, see Kitamura et al., Nature, 350:423-, 1991).

[0280] Thus, an antibody heavy-chain knockout mouse strain could be established from the antibody heavy-chain one allele-disrupted TT2F cell clone.

[0281] Embryos obtained by mating the homozygous male and female mice bred in a clean environment may be used as hosts for producing chimeric mice. In this case, most of the B cells functional in the resultant chimeric mice are derived from the injected ES cells. Other mouse strains which cannot produce their own functional B cells, such as RAG-2-deficient mouse (Sinkai et al., Cell, 68:855-, 1992), may also be used for this purpose. In this system, chimeric mice are produced by the same procedures as in Example 10, etc. using the mouse ES cell clone from Examples 62, 63 or 64 which is deficient in endogenous antibody heavy-chain & light-chain and which retains human chromosomes #14+#2, #14+#22 or #14+#2+#22 (fragments). The resultant chimeric mice mainly produce human antibodies by the expression of human antibody heavy-chain (on chromosome #14), light-chain  $\kappa$  (on chromosome #2) and light-chain  $\lambda$  (on chromosome #22) genes that are functional in ES cell-derived B cells.

(2) Detection and quantitative determination of complete human antibody in sera of the chimeric mice produced by injecting the endogenous antibody heavy-chain and  $\kappa$  chain-deficient mouse ES cells retaining both a human chromosome #2 partial fragment and a human chromosome #14 partial fragment into immunodeficient mouse host embryos

[0282] Chimeric mice were produced in the same manner as in Section 1 of Example 65 by injecting the ES cell clone KH10 from Example 62 into the embryos obtained by mating male and female mice of the antibody heavy-chain knockout mouse strain established in Section 1 of Example 67. Seven-week old resultant chimeric mice were bled to determine the concentrations of human antibodies in the sera by ELISA in the same manner as in Example 14 and Section 3 of Example 65. The results are shown in Table 23. Complete human antibodies having human  $\mu$  chain +  $\kappa$  chain and human  $\gamma$  chain +  $\kappa$  chain, respectively, were confirmed in the sera of the chimeric mice. It was also confirmed that by transferring ES cells into immunodeficient host embryos, complete antibodies could be obtained even in the resultant chimeric mice of low chimerism since B cells are differentiated only from ES cells.

Table 23.

Concentrations of Human Antibodies in Chimeric Mice (ELISA)				
ES clone	Chimeric mouse	Chimerism (%)	IgM, $\kappa$ (mg/l)	IgG, $\kappa$ (mg/l)
KH13	CKH10-1	6	6.1	0.17
KH13	CKH10-2	3	1.9	0.4

(3) Detection and quantitative determination of complete human antibody in sera of the chimeric mice produced by injecting the endogenous antibody heavy-chain and  $\kappa$  chain-deficient mouse ES cells retaining both a human chromosome #14 partial fragment and a human chromosome #22 partial fragment into immunodeficient mouse host embryos

[0283] Chimeric mice were produced in the same manner as in Section 2 of Example 65 by injecting the ES cell clone LH13 from Example 62 into the embryos obtained by mating male and female mice of the antibody heavy-chain knockout mouse strain established in Section 1 of Example 67. Five-week old resultant chimeric mice were bled to determine the concentrations of human antibodies in the sera by ELISA in the same manner as in Example 14 and Section 4 of Example 65. The results are shown in Table 24. Complete human antibodies having human  $\mu$  chain +  $\lambda$  chain and human  $\gamma$  chain +  $\lambda$  chain, respectively, were confirmed in the sera of the chimeric mice.

Table 24.

Concentrations of Human Antibodies in Chimeric Mice (ELISA)			
ES clone	Chimeric mouse	Chimerism (%)	IgM, $\lambda$ (mg/l)
LH13	CLH13-3	35	51
LH13	CLH13-4	85	32
LH13	CLH13-4	30	27

#### Example 68

Retention of the human chromosome in offsprings of human chromosome #14 fragment (containing antibody heavy-chain gene)-transferred ES cell-derived chimeric mice

(1) Isolation of human-mouse hybrid cells retaining a human chromosome #14 fragment containing an antibody heavy chain gene

[0284] It was observed in Example 42 that a human chromosome #2 fragment transferred into mice was transmitted to their progeny. Thus, it is expected that the possibility of transmission of human chromosome #14 to progeny will be increased if a fragment of this chromosome is used. A9/#14 clone (Example 9; corresponding to A9/14-C11 clone described in Tomizuka et al., Nature Genet. vol 16, 133-143 (1997)) retaining an intact human chromosome #14 marked with a G418 resistance gene was subjected to a more detailed FISH analysis (Example 9). As a result, it was observed that about 10% of the cell population contained only a very small, fragmented human chromosome #14. This chromosome fragment is almost of the same size as the chromosome #2 fragment (Example 12) and believed to contain the G418 resistance marker.

[0285] In order to isolate the cell clones containing the fragmented human chromosome #14, (about 300) A9/#14 cells were seeded on 10 cm plates and cultured. At day 10 of the cultivation, 31 colonies were picked up. Genomic DNAs were prepared from these clones and subjected to PCR analysis in the same manner as in Example 9 using chromosome #14 specific primers (the 18 primers shown in Example 9 were used except PCI and NP). Out of the 16 primers, only IgM, IGG1, IGA2 and IGVH3 were found in one clone (A9/SC20). Since D14S543 (Science, HUMAN GENETIC MAP (1994); the base sequence was obtained from databases of GenBank, etc.) which is a marker located near the human chromosome #14 long arm telomere was also detected in the clone, the fragment of interest (hereinafter referred to as "SC20 fragment") retained in the clone is believed to contain a region adjacent to the chromosome #14 telomere and which contained an antibody heavy-chain gene.

[0286] SC20 fragment was subjected to FISH analysis (Tomizuka et al., Nature Genet. vol 16, 133-143 (1997)) using a human chromosome-specific probe. As a result, it was observed that the size of the chromosome in the clone that hybridized to the probe was smaller than in the control clone (containing an intact chromosome #14). Thus, it was confirmed that A9/SC20 contained a fragment of human chromosome #14.

**[0287]** Further, in order to examine whether SC20 fragment contained a human chromosome #14-derived centromere sequence, chromosome samples from A9/SC20 cells were hybridized to digoxigenin-11-dUTP-labeled human chromosome #14 or #22-specific  $\alpha$  satellite DNA (purchased from COSMOBIO) which was used as a probe, followed by FISH analysis according to the method described in a reference (Tomizuka et al., Nature Genet. vol 16, 133-143 (1997)).  
 5 As a result, a signal hybridizing to the above probe was confirmed. Thus, it has become clear that SC20 fragment contains a human-derived centromere sequence (Fig. 36).

(2) Transfer of human chromosome #14 (fragment) into TT2F cells and stable retention of the chromosome therein

10 **[0288]** A human chromosome #14 fragment was microcell-transferred into TT2F cells using A9/SC20 as a chromosome donor cell in the same manner as in Example 9. As a result of G418 (300  $\mu$ g/ml) selection, 5 resistant clones were obtained. These ES cell clones were subjected to PCR analysis (Example 68, Section 1) and FISH analysis (using human chromosome-specific probes; Tomizuka et al., supra) to confirm the retention of a human chromosome #14 fragment. The results of the FISH analysis are shown in Fig. 37.

15 **[0289]** Stable retention of transferred human chromosomes in mice is important for efficient expression of the transferred genes and efficient transmission of the transferred chromosomes to their progeny. Since selection by addition of drugs is impossible after an ES cell clone has been injected into host embryos, it is desired that transferred human chromosomes be retained stably even under non-selective conditions.

20 **[0290]** TT2F(SC20)-21 clone containing SC20 fragment was cultured in a medium not containing G418 for a long period to examine the retention of SC20 fragment under this condition.

25 **[0291]** Briefly, TT2F(SC20)-21 clone was cultured in a selective medium (G418: 300  $\mu$ g/ml) for 1 week and then subcultured in a non-selective medium for 45 days (subcultured every other day with 8-fold dilution). At day 0, 15, 30 and 45 of the subculture, 300-1000 cells were seeded in six 35 mm plates, three of which contained the selective medium and the other three non-selective medium. The cells were cultured in these plates for about 1 week and then the colonies were counted. Chromosome retention ratio (A/B) was calculated by dividing the total number of colonies in the 3 plates under selective conditions (A) by the total number of colonies in the 3 plates under non-selective conditions (B). For the purpose of comparison, an experiment was conducted using P-21 clone (Example 40) containing W23 fragment derived from human chromosome #2 in the same manner as described above (the selective medium contained 0.75  $\mu$ g/ml of puromycin). The results are shown in Fig. 38. The values shown in Fig. 38 are average values from 3 independent experiments. SC20 fragment exhibited a high retention ratio of 95% or above even after the 45 day cultivation under non-selective conditions. On the other hand, W23 fragment exhibited a retention ratio of 14% under identical conditions.

35 **[0292]** There have been reports of the transfer of a human Y chromosome-derived artificial chromosome (containing human Y-derived centromere) into CHO (hamster fibroblasts), DT40 (chicken B lymphocytes) and mouse ES cell (Shen et al., Hum. Mol. Genet. 6, 1375-1382). Under non-selective cultivation, the artificial chromosome was retained stably in CHO and DT40. In mouse ES cells, however, only the chromosome which accidentally acquired the mouse centromere as a result of rearrangement was retained stably. From these results, an opinion was proposed that a human-derived centromere is unstable in mouse ES cells (Shen et al., supra).

40 **[0293]** The results described above show that SC20, though it contains a human-derived centromere (Example 68, Section 1), is very stable in mouse ES cells. Since the retention of W23 fragment (which is also suggested to contain a human-derived centromere) (Tomizuka et al., supra) in mouse ES cells appeared to be unstable, it is considered that the stability of human-derived chromosomes in mouse ES cells varies depending on the type of the chromosome.

45 **[0294]** From these results, it was demonstrated that SC20 fragment is very useful as a vector for transferring a gene into mice.

(3) Production of chimeric mice from the ES cell clone retaining human chromosome #14 (fragment)

50 **[0295]** Cells in the frozen stock of G418 resistant ES cell clone TT2F(SC20)-21 which was obtained in Example 68, Section 2 and which was confirmed to retain a human chromosome #14 fragment were thawed, started up for culture and injected into 8-cell stage embryos obtained by mating male and female mice of ICR (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. After the embryos were cultured overnight in the medium for ES cells (see Example 9) to develop into blastocysts, about 10 of the injected embryos were transplanted to each side of the uterus of foster mother ICR mice (CREA JAPAN, INC.; 2.5 days after pseudopregnant treatment).

55 **[0296]** As a result of transplantation of a total of 188 injected embryos, 22 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo (ICR)-derived albino coat color (white). Out of the 22 offsprings, 20 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells. Out of the 20 mice, two were chimeric mice in which their coat color was complete agouti (i.e. ES cell-derived).



[0297] From these results, it was confirmed that ES cell clone TT2F(SC20)-21 retaining a human chromosome #2 fragment maintains the ability to produce chimera, that is, the ability to differentiate into normal tissues of mice.

[0298] Two 5-week old chimeric mice [derived from TT2F(SC20) - 21, chimerism 100%, C14m-16 and -17] were bled to determine the concentrations of human antibody IgM and IgG in the sera by ELISA in the same manner as in Example 14. The results are shown in Table 25.

Table 25.

Concentrations of Human Antibody Heavy-Chains in Chimeric Mice (ELISA)			
Chimeric mouse	Chimerism (%)	IgM (mg/l)	IgG (mg/l)
C14m-16	100	7.9	1.0
C14m - 17	100	6.0	1.3

[0299] Human antibodies IgM and IgG were detected in the sera of both chimeric mice. The concentrations of these human antibodies were comparable to the concentrations in the chimeric mice retaining the larger human chromosome #14 fragment (see Example 14). Thus, it was demonstrated that the human antibody gene contained in SC20 fragment is functional.

(4) Confirmation of the retention of human chromosome in the progeny of chimeric mice derived from the mouse ES cells (TT2F, XO) retaining a human chromosome #14 fragment, and detection and quantitative determination of human antibody  $\mu$  chain and  $\gamma$  chain in sera of the progeny

[0300] Examination was made as to whether ES cell-derived offsprings would be produced by mating the female chimeric mice C14m-16 and C14m-17 (both having 100% chimerism in coat color) from Example 68, Section 3 with male ICR mice. By this mating, offsprings with agouti coat color should be produced from TT2F cell (agouti: dominant)-derived oocytes in the chimeric mice fertilized by male ICR mouse (albino: recessive)-derived sperms, and offsprings with albino coat color should be produced from ICR-derived oocytes in the chimeric mice. Actually, all of the viable offspring mice obtained by this mating (30 in total) exhibited ES cell-derived agouti coat color, indicating efficient transmission of ES cells to the germ cell lineage. Genomic DNAs were prepared from the tails of these offspring mice to examine the retention of a human chromosome fragment by PCR. PCR amplification was performed using the three primers (IGVH3, IgM and D14S543) of which the presence in TT2F(SC20)-21 was confirmed. As a result, the presence of the three markers detected in TT2F(SC20)-21 was confirmed in 10 out of the 30 offspring mice (33%). These results show that TT2F cell clone TT2F(SC20)-21 retaining a human chromosome #14 fragment differentiated into functional oocytes in the chimeric mice and that the human chromosome #14 fragment was transmitted to the F1 progeny derived from the oocytes.

[0301] Detection and quantitative determination of human antibodies IgM and IgG in sera were performed on 9 out of the 10 offspring mice which were confirmed to retain a human chromosome #14 fragment, as described below. About 4-8 week-old mice were bled to detect human antibody  $\mu$  chain and  $\gamma$  chain by ELISA in the same manner as in Example 14. As a result, human antibody  $\mu$  chain and  $\gamma$  chain were detected in the sera of all of the mice tested (see Table 26). Thus, it was confirmed that the human antibody heavy chain gene also functions in the F1 progeny born by the chimeric mice.

Table 26.

Concentrations of Human Antibodies IgM and IgG in Chimeric Mice (ELISA)			
Mother Chimeric Mouse	Mouse Individual No.	IgM (mg/l)	IgG (mg/l)
C14m-16	16-5	12.9	2.2
C14m-16	16-14	3.5	2.2
C14m-16	16-16	4.1	2.0
C14m-16	16-17	5.5	3.9
C14m-17	17-7	5.7	1.0
C14m-17	17-8	3.6	1.2
C14m-17	17-19	3.5	0.75
C14m-17	17-22	2.4	1.4
C14m-17	17-23	5.3	1.9

**[0302]** Further, 3 male mice and 4 female mice in the F1 progeny were mated with MCH(ICR) mice (purchased from CREA JAPAN, INC.) to obtain F2 progenies, which were subjected to PCR analysis of tail DNA and analysis for human antibody  $\mu$  chain expression as described above. As a result, it was confirmed that SC20 fragment was transmitted to 30% of the F2 progeny through F1 male mice (43 out of the 142 offsprings were positive) and to 33% of the F2 progeny through F1 female mice (20 out of the 60 offsprings were positive).

**[0303]** These results show that a mouse strain was established which retains the human chromosome #14 fragment (containing a human antibody heavy chain gene), which expresses human antibody heavy-chains and which can transmit the human chromosome to the subsequent generation.

#### (5) Stable retention of a human chromosome #14 fragment in mice

**[0304]** Three F1 mice (16-5, 17-8 and 17-23 shown in Table 26) which were obtained in Example 68, Section 4 and which retained SC20 fragment were used in analysis for the ratio of retention of SC20 fragment in mice. The mice were injected intraperitoneally with 0.3 ml of CORCEMID (100  $\mu$ g/ml) and then killed by dislocation of the cervical vertebrae in an euthanasic manner, followed by removal of the brain, liver, spleen, testis and bone marrow. All of these tissues except the bone marrow were washed with PBS(-), cut into pieces with scissors for anatomy, given hypotonic treatment with KCl (0.075 M) for 15 minutes, and fixed in Carnoy's fixative. Specimens were prepared using the supernatant of the Carnoy fixation by conventional methods. FISH analysis was performed using a human chromosome-specific probe (Human COT-1 DNA) according to the method described in a reference (Tomizuka et al., Nature Genetics, 16, 133-143). As to the brain, spleen, liver and bone marrow, 30 or more nuclei in interphase were selected randomly for each of these tissues. Then, the number of nuclei in which a signal was detected (mark "+" in Fig. 39) and the number of nuclei in which a signal was not detected (mark "-" in Fig. 39) were counted to calculate the retention ratio. The testis were classified into the 1st meiosis phase spreads, the 2nd meiosis phase spreads and sperms. Ten or more spreads or sperms were selected for each group and then counting was performed in the same manner as described above to calculate the retention ratio. As a result, all of the 3 mice exhibited a retention ratio of almost 100% in the brain and liver. A decrease in the retention ratio was observed in the bone marrow and spleen. In the testis, a retention ratio of 80-100% was obtained for the 1st meiosis phase spreads, and a retention ratio of 30-50% for sperms. Assuming that SC20 fragment is retained stably, the theoretical retention ratio should be 100% for the 1st meiosis phase spreads and 50% for the 2nd meiosis phase spreads and sperms. Thus, it is believed that SC20 fragment is retained stably in the testis.

**[0305]** At the same time, fibroblasts were prepared from the tail and then the ratio of retention of SC20 fragment was examined in the same manner as in Example 79. As a result, the retention ratios in mice 16-5, 17-8 and 17-23 were 98%, 96% and 98%, respectively (50 nuclear plate were tested for each mouse).

#### (6) Hereditary relief of antibody production ability-deficient mice by the transfer of a human chromosome #14 fragment (containing antibody heavy-chain gene)

**[0306]** The knockout mouse whose antibody  $\mu$  chain gene essential for the generation of B lymphocytes is disrupted (Section 1, Example 67) cannot produce antibody because the mouse is deficient in mature B lymphocytes responsible for humoral immunity. The following experiment was conducted to examine as to whether this deficiency could be relieved by transferring SC20 fragment (containing a human antibody heavy-chain gene) by mating.

**[0307]** Those mice exhibiting agouti coat color among the progeny obtained by mating the endogenous antibody heavy-chain one allele-disrupted TT2F cell clone-derived chimeric mice from Example 49 with MCH(ICR) mice were subjected to Southern blot analysis to select mice retaining the disrupted allele. A female antibody heavy-chain-deficient heterozygote thus selected was mated with a male F1 offspring (17-7) which was obtained in Example 68, Section 4 and which retains SC20 fragment. The resultant 5 offspring mice were subjected to both PCR analysis for confirming the retention of SC20 fragment and determination of human antibody  $\mu$  chain and  $\gamma$  chain in the sera (see Example 68, Section 4). As a result, it was confirmed that three mice #2, #3 and #5 retained SC20 fragment (Table 27). Furthermore, as a result of the analysis for mouse antibody  $\mu$  chain expression (Example 75), it was demonstrated that mice #2 and #3 are mouse  $\mu$  chain-negative, that is, endogenous antibody heavy-chain-deficient homozygotes (Table 27). These results were consistent with the results of Southern blot analysis (see Example 49) using the DNAs prepared from the tails of the 5 mice. Compared to mouse #1 in which neither mouse nor human antibody heavy chain was detected, very high concentrations of human antibody  $\mu$  chain (310 mg/l) and  $\gamma$  chain (860 mg/l) were detected in mouse #3 which is antibody heavy-chain-deficient homozygote and which retains the human chromosome #14 fragment. Further, quantitative determination of human  $\gamma$  subclasses was performed on mouse #3 in the same manner as in Example 29 to detect all of the 4 subclasses ( $\gamma$  1,  $\gamma$  2,  $\gamma$  3 and  $\gamma$  4). In particular, the concentration of human  $\mu$  chain in this mouse is comparable to the concentration of mouse  $\mu$  chain in wild-type mice (Mendez et al., Nature Genet. 15, 146-156 (1977)). These results show that the symptom of inability for antibody production because of disruption of

endogenous heavy-chain gene (deficiency of B lymphocytes: see Kitamura et al., Nature, 350, 423-, 1991) in this mouse was cured by the transfer of human chromosome #14 fragment SC20 (containing an antibody heavy-chain gene), and that the mouse has recovered the ability to produce antibody and the ability to produce B lymphocytes.

Table 27.

Mouse No.	Retention of SC20 Fragment	Mouse $\mu$ Chain	Human IgM (mg/l)	Human IgG (mg/l)
1	-	-	Below detection limit	0.33
2	+	+	8.4	5.3
3	+	-	310	860
4	-	+	Not measured	Not measured
5	+	+	4.8	0.86

## Example 69

Retention of the human chromosome in offsprings of human chromosome #22 (fragment)-transferred ES cell-derived chimeric mice

## (1) Fragmentation of human chromosome #22 using microcell fusion

[0308] Since it was observed that both a human chromosome #2 fragment (Example 42) and a human chromosome #14 fragment (Example 68, Section 4) once transferred into mice were transmitted to their offsprings, it is expected that fragmentation of human chromosome #22 would increase the possibility of transmission of this chromosome to offspring mice. When a human chromosome is transferred into a recipient cell by microcell fusion, it is observed that 40-80% of the transferred clones retain the human chromosome which has been fragmented at the time of fusion (Oshimura et al., Protein, Nucleic Acid, Enzyme, vol. 35, No. 14, 1990). The present inventors tried to fragment human chromosome #22 utilizing this phenomenon.

[0309] A microcell fusion experiment (see Example 1) was conducted using clone 6-1 from Example 35 as a chromosome donor cell and wild-type mouse A9 cells as a recipient cell, thereby producing seventy-three G418 resistant clones. Genomic DNAs were prepared from the resultant clones and then screened by PCR using Ig  $\lambda$  primers (Example 2). Sixty-seven clones which retained human Ig  $\lambda$  gene were subjected to PCR analysis using primers specific to 8 markers located on human chromosome #22 (D22S315, D22S275, D22S280, D22S278, D22S283, D22S272, D22S282 and D22S274; for the order of location on human chromosome #22, see Nature, vol. 377, 367-379 (1995); base sequences for these primers were obtained from databases of such as GenBank). As a result, it was found that a part of the markers disappeared in 25 clones. Thus, it was suggested that chromosome #22 was fragmented in these clones (Fig. 40). Among them, clone #22 and clone #28 are considered to have a fairly small fragment, because markers other than Ig  $\lambda$  and D22S315 disappeared in the former and markers other than Ig  $\lambda$  disappeared in the latter (Fig. 40). Clone #28 was subjected to FISH analysis (see Example 18) using a human chromosome-specific probe. The results are shown in Fig. 41. It is observed that the size of the chromosome hybridizing to the probe is smaller in this clone than in the control clone (containing an intact chromosome #22). Thus, a human chromosome #22 fragment containing antibody  $\lambda$  gene could be obtained as a result of fragmentation which occurred at the time of microcell fusion.

## (2) Transfer of chromosome #22 (fragment) into TT2F cells

[0310] Chromosome #22 (fragment) was microcell-transferred into TT2F cells by the method described in Example 2, using clones #22, #28 and 6-1 as chromosome donor cells. Clones #22 and A9/#22(6-1) were subjected to puromycin (0.75  $\mu$ g/ml) selection, and clone #28 was subjected to G418 (225  $\mu$ g/ml) selection. As a result, drug resistant clones were obtained as follows: 13 from clone #22, 5 from clone 6-1 and 3 from clone #28. These ES cell clones are subjected to PCR analysis and FISH analysis to confirm the retention of human chromosome #22 (fragment) in the same manner as in Example 69, Section 1.

## (3) Production of chimeric mice from ES cell clones retaining human chromosome #22 (fragment)

[0311] In the same manner as in Example 3, chimeric mice are produced from the drug resistant ES cell clones which were obtained in Example 69, Section 2 and which were confirmed to retain human chromosome #22. Confirmation of the retention of human chromosome #22 (fragment) in the resultant chimeric mice is performed by the method described in Example 4.

## (4) Transmission of human chromosome #22 (fragment) to offsprings

**[0312]** The chimeric mice retaining human chromosome #22 (fragment) are mixed and mated with ICR mice. Retention of a human chromosome #22 fragment in the offsprings is examined by PCR using genomic DNAs prepared from the tails of the offspring mice having agouti coat color (see Examples 30, 42 and 43). As shown in Examples 42 and 43, mouse ES cell clones retaining human chromosome #22 or a fragment thereof can differentiate into oocytes or sperms functional in chimeric mice, thereby allowing to the human chromosome #22 (fragment) to be transmitted their progenies. Thus, it is possible to establish a mouse strain which retains human chromosome #22 (fragment) containing human antibody light-chain  $\lambda$  gene and which can transmit it to the subsequent generation.

## Example 70

Production of mice retaining both human chromosome #2 (fragment) and #14 (fragment) by mating

**[0313]** The human chromosome #2 (fragment)-retaining mouse strain from Example 42 or 43 is mated with the human chromosome #14 (fragment)-retaining mouse strain from Example 68 to produce offsprings. Genomic DNAs are prepared from the tails of the offspring mice. The DNA is analyzed by PCR, etc. (Examples 9, 42 and 43) to produce those mice which retain both human chromosome #2 partial fragment and human chromosome #14 (fragment).

## Example 71

Production of mice retaining both human chromosome #22 (fragment) and #14 (fragment) by mating

**[0314]** The human chromosome #22 (fragment)-retaining mouse strain from Example 69 is mated with the human chromosome #14 (fragment)-retaining mouse strain from Example 68 to produce offsprings. Genomic DNAs are prepared from the tails of the offspring mice. The DNA is analyzed by PCR, etc. (Examples 30, 42 and 43) to produce those mice which retain both human chromosome #22 (fragment) and #14 (fragment).

## Example 72

Production of mice retaining the three human chromosomes #2 (fragment), #14 (fragment) and #22 (fragment) by mating

**[0315]** The mouse strain retaining both human chromosome #2 (fragment) and #14 (fragment) obtained in Example 71 is mated with the mouse strain retaining a human chromosome #22 fragment obtained in Example 42 or 43 to produce offsprings. Genomic DNAs are prepared from the tails of the offspring mice. The DNA is analyzed by PCR, etc. (Examples 9, 30, 42 and 43) to produce those mice which retain all of the three human chromosomes, #22 (fragment), #14 (fragment) and #2 (fragment). Alternatively, mice retaining all of the above three human chromosomes may also be obtained by mating the mouse strain retaining both human chromosome #2 (fragment) and #14 (fragment) from Example 70, with the mouse strain retaining a human chromosome #22 fragment from Example 69.

## Example 73

Production of a complete human antibody-producing mouse strain by mating

**[0316]** The mouse strains retaining human chromosomes #2+#14 (Example 70), #14+#22 (Example 71) and #2+#14+#22 (Example 72), respectively, are repeatedly mated with a mouse strain deficient in endogenous antibody heavy-chain and light-chain  $\kappa$  genes. From the resultant offsprings, those mouse strains which retain human chromosomes #2+#14, #14+#22 or #2+#14+#22 and which are homozygotes in the deficiency of endogenous antibody heavy-chain and light-chain  $\kappa$  genes, are selected by PCR analysis, etc. (Examples 9, 30, 42 and 43). In these strains, complete human antibodies are mainly produced (Green et al., Nature Genetics, 7:13-, 1994; Lonberg et al., Nature, 368:856-, 1994).

**[0317]** Hereinbelow, the establishment of a mouse strain which retains both a human chromosome #2 fragment and a human chromosome #14 fragment and which is homozygote in the deficiency of endogenous antibody heavy-chain and light-chain  $\kappa$  genes will be described. The 4 strains used for the mating and the method for assaying the genotypes of each strain are as follows.

(1) The mouse strain from Example 42 retaining a human chromosome #2 fragment: the retention of the human

chromosome #2 fragment is assayed by PCR analysis of the tail-derived DNA as described in Example 42 and by the expression of human antibody  $\kappa$  chain in the sera.

(2) The mouse strain from Example 68, Section 4 retaining a human chromosome #14 fragment: the retention of the human chromosome #14 fragment is assayed by PCR analysis of the tail-derived DNA as described in Example 68, Section 4 and by the expression of human antibody  $\mu$  chain in the sera.

(3) The antibody heavy-chain knockout mouse strain from Example 67, Section 1: heavy-chain deficiency-homozygotes or heterozygotes were assayed by Southern blot analysis of the tail-derived DNA as described in Example 67, Section 1 and by the presence or absence of the expression of mouse antibody  $\mu$  chain in the sera (see Example 75).

(4) The antibody  $\kappa$  chain knockout mouse strain from Example 80:  $\kappa$  chain deficiency-homozygotes or heterozygotes were assayed by Southern blot analysis of the tail-derived DNA as described in Example 80.

[0318] A mouse strain which retains all of the 4 genotypes (i.e., retaining a human chromosome #2 fragment, retaining a human chromosome #14 fragment, antibody heavy-chain-deficiency homozygote or heterozygote, and antibody  $\kappa$  chain-deficiency homozygote or heterozygote) was established by mating the above 4 strains with each other. Specifically, after the above 4 strains used as starting materials were mated several times, a male mouse having the genotypes of "retaining the human chromosome #14 fragment, antibody heavy-chain-deficiency homozygote and antibody  $\kappa$  chain-deficiency heterozygote" was mated with a female mouse having the genotypes of "retaining the human chromosome #2 fragment, antibody heavy-chain-deficiency homozygote and antibody  $\kappa$  chain-deficiency homozygote" or "retaining the human chromosome #2 fragment, antibody heavy-chain-deficiency homozygote and antibody  $\kappa$  chain-deficiency heterozygote" or "retaining the human chromosome #2 fragment, antibody heavy-chain-deficiency heterozygote and antibody  $\kappa$  chain-deficiency homozygote". As a result, mouse HK23 "retaining the human chromosome #2 fragment, retaining the human chromosome #14 fragment, antibody heavy-chain-deficiency homozygote and antibody  $\kappa$  chain-deficiency heterozygote" and mouse HK29 "retaining the human chromosome #2 fragment, retaining the human chromosome #14 fragment, antibody heavy-chain-deficiency heterozygote or wild-type, and antibody  $\kappa$  chain-deficiency heterozygote" were obtained. Fig. 42 shows the concentration of each antibody in the sera and the genotypes of these mice, together with the data on mouse HK28 which was also produced by the above-described mating and which has the genotypes of "retaining the human chromosome #2 fragment, retaining the human chromosome #14 fragment, antibody heavy-chain-deficiency heterozygote or wild-type, and antibody  $\kappa$  chain-deficiency wild-type". A complete human antibody consisting of human  $\mu$  chain and human  $\kappa$  chain was detected at a concentration of 18 mg/l in the serum of mouse HK23 (Example 38).

[0319] It is possible to produce those mice having the genotypes of "retaining the human chromosome #2 fragment, retaining the human chromosome #14 fragment, antibody heavy-chain-deficiency homozygote and antibody  $\kappa$  chain-deficiency homozygote" by mating the mice obtained by the above mating with each other. In this mouse strain, it is expected that human antibody  $\kappa$  chain will be expressed at a higher concentration than in mouse HK23 because the deficiency of the endogenous  $\kappa$  chain gene is substituted by the human antibody  $\kappa$  chain gene contained in the human chromosome #2 fragment (Lonberg et al., Nature, 368, 856-, 1994). It is also expected that the concentration of a complete human antibody consisting of human heavy-chain and human  $\kappa$  chain will increase further.

#### Example 74

Production of a human antibody-producing hybridoma from a mouse strain which is obtained by mating and which retains a human chromosome(s) containing a human antibody gene(s)

[0320] The mice retaining a human chromosome(s) containing a human antibody gene(s) which were obtained in Example 42, 43, 68, 69, 70, 71, 72 or 73 are immunized with an antigen of interest in the same manner as in Example 25. The spleen is removed from each mice and the spleen cells are fused with myeloma cells to produce hybridomas. After cultivation for 1-3 weeks, the culture supernatant is analyzed by ELISA. The ELISA is performed by the method described in Examples 14, 15, 21, 22, 25, 33, 34, 37 and 38. As a result, human antibody positive clones and clones which are human antibody positive and specific to the antigen used in the immunization are obtained.

#### Example 75

Detection and determination of mouse IgM in sera of chimeric mice derived from the mouse antibody heavy-chain both alleles-disrupted TT2F cell clone

[0321] Offspring mice were born in the same manner as in Example 40 from the mouse antibody heavy-chain both alleles-disrupted TT2F cell clone (#131-3) from Example 51. Three mice having chimerisms of 0%, 50% and 99%,

respectively, were selected from the offspring mice. Mouse IgM in their sera was detected and determined. Briefly, the chimeric mice of about 2 weeks after birth were bled and mouse IgM concentration in the sera was determined by ELISA by the same procedures as in Example 14. A PBS-diluted anti-mouse IgM antibody (Kirkegaard & Perry Laboratories Inc., 01-18-03) was fixed, and then a PBS-diluted serum sample supplemented with 5% FBS was added. Peroxidase-labeled anti-mouse IgM antibody (Kirkegaard & Perry Laboratories Inc., 074-1803) was added and the absorbance at 450 nm was determined using TMBZ as a substrate. Purified mouse IgM (Pharmingen, 03081D) was used as a standard. This standard was diluted stepwise with FBS-supplemented PBS. The results are shown in Table 28. Of the chimeric mice derived from the mouse antibody heavy-chain both alleles-disrupted TT2F cells, the mouse having a chimerism of 99% exhibited a low mouse IgM concentration. Thus, it was confirmed that the mouse heavy-chain gene from the ES cells hardly functions in this mouse.

Table 28.

Concentration of Mouse IgM in Chimeric Mice (ELISA)	
Chimerism %	IgM (mg/l)
0	12
50	11
99	1.5

#### Example 76

Preparation of a targeting vector for knocking out antibody light-chain  $\kappa$  gene in ES cells

**[0322]** Plasmid LoxP-PGKPuro in which LoxP sequence was inserted at both ends of a puromycin resistance gene was prepared in the same manner as in Example 48, Section 1. Briefly, a puromycin resistance cassette PGKPuro was cut out from PGKPuro plasmid DNA (Watanabe et al., Biochem. Biophys. Res. Commun. 213:130-137 (1995); released from Peter W. Laird, Whitehead Institute for Biochemical Research and Massachusetts Institute of Technology, Dept. of Biology, Cambridge, MA) using restriction enzyme Sall and then blunted. PGKPuro was inserted into the SmaI and EcoRV restriction sites of a LoxP-sequence containing plasmid to produce plasmid pLoxP-PGKPuro (Fig. 30). Further, a DNA fragment comprising a genomic DNA constant region containing the mouse antibody light-chain  $\kappa$  J region and constant region was replaced with the LoxP-PGKPuro gene in the same manner as in Example 48 (Fig. 31).

#### Example 77

Production of an antibody light-chain gene both alleles-disrupted strain from antibody light-chain deficient-heterozygote (and antibody heavy-chain-deficient-homozygote) mouse ES cells

(1) A puromycin resistance gene was inserted into the antibody light-chain deficient-heterozygote TT2F clone (HD43) obtained in Example 58 to give a strain in which both alleles of a light-chain gene were disrupted.

**[0323]** The antibody light-chain targeting vector prepared in Example 76 was linearized with restriction enzyme KpnI to transform HD43 clone in the same manner as in Example 58. The resultant transformants were subjected to selective culture at a puromycin concentration of 0.75  $\mu$ g/ml. At day 7-9 of the cultivation, colonies formed were picked up. A part of these colonies was stored frozen, and the remaining part was used to prepare genomic DNA in the same manner as in Example 49. Genomic DNAs from the puromycin resistant strains were digested with restriction enzyme EcoRI (Takara Shuzo), separated by agarose gel electrophoresis and subjected to Southern blot analysis to detect homologous recombinants using the probe described in Example 48, Section 4 (see Examples 58 and 59). As a result, 4 clones in which both alleles of an antibody light-chain were disrupted were obtained from the 74 clones analyzed. Under usual culture conditions, no changes in growth rate and morphology were observed in these clones, as compared to the TT2F clone before gene disruption. This suggests that the clones under consideration retain the ability to produce chimera.

(2) Production of chimeric mice from the antibody heavy-chain-deficient-homozygote and antibody light-chain gene both alleles-disrupted clone

**[0324]** Cells in the frozen stock of antibody light-chain gene both alleles-disrupted TT2F cell clone HD43P-10 from Example 77, Section 1 were thawed, started up for culture and injected into 8-cell stage embryos obtained by mating

male and female mice of ICR (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. After the embryos were cultured overnight in the medium for ES cells (see Example 9) to develop into blastocysts, about 10 of the injected embryos were transplanted to each side of the uterus of foster mother ICR mice (CREA JAPAN, INC.; 2.5 days after pseudopregnant treatment).

**[0325]** As a result of transplantation of a total of 161 injected embryos, 37 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo (ICR)-derived albino coat color (white). Out of the 37 offsprings, 9 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells. Out of the 9 mice, four were chimeric mice in which more than 80% of their coat color was agouti (i.e. ES cell-derived).

**[0326]** From these results, it was confirmed that antibody light-chain both alleles-disrupted ES cell clone HD43P-10 maintains a high ability to produce chimera.

#### Example 78

Removal of the G418 resistance and puromycin resistance marker genes from the antibody light-chain deficient-homozygote (and antibody heavy-chain deficient-homozygote) TT2F cell clone

**[0327]** From the antibody light-chain both alleles-disrupted HD43P-10 clone (puromycin resistant, G418 resistant) which was obtained and confirmed to have a high chimera-forming ability in Example 77, the puromycin resistance and G418 resistance marker genes were removed by the procedures described in Example 52. Briefly, an expression vector pBS185 (BRL) containing a Cre recombinase gene which causes a site-directed recombination between the LoxP sequences inserted at both ends of the G418 resistance marker gene was transferred into the clone described above in the same manner as in Example 52. The resultant puromycin (0.75 µg/ml) sensitive clones (6 clones) were grown to confluence in 35 mm plates in the same manner as in Example 52. Three fifths (3/5) of the resultant culture were suspended in 0.5 ml of a preservation medium [ES medium + 10% DMSO (Sigma)] and stored frozen at -80°C. The remaining two fifth (2/5) were divided into two portions and inoculated into two 12-well gelatin-coated plates. Cells in one plate were cultured in non-selective medium for 2 days. Cells in other plate were cultured in the presence of 300 µg/ml of G418 for 2 days. As a result, 5 puromycin sensitive and G418 sensitive clones which would be killed in the presence of G418 were obtained.

#### Example 79

Increase in the expression of human antibody κ chain in sera as a result of mating a human chromosome #2 fragment-retaining mouse strain with C57BL/6 strain

**[0328]** The hereditary background of the progenies of the chimeric mice [hereinafter referred to as "F1(chimera x MCH)"] which were described in Examples 43 and 44 and which retain a human chromosome #2 fragment (hereinafter referred to as "W23 fragment") is that they are a mixture of TT2F cell (Example 39)-derived CBA mouse strain and C57BL/6 mouse strain, and MCH(ICR) mouse strain mated with the chimeric mice. In order to observe the behavior of W23 fragment under a hereditary background as homogeneous as possible, first, F1 (chimera x MCH) were back-crossed with MCH(ICR). The offspring mice obtained by the mating of F1 (chimera x MCH) (randomly selected 8 male and 6 female mice) X MCH(ICR) were examined as to whether they would retain W23 fragment in the same manner as in Example 43. As a result, it was confirmed that W23 fragment was transmitted through male to 8% of the offsprings (25 out of the 324 offsprings were positive) and through female to 22% of the offsprings (32 out of the 148 offsprings were positive). When the resultant F2(F1 x MCH) (randomly selected 8 male and 8 female mice) were further mated with MCH(ICR), the transmission ratio was 9% through male (30 out of the 346 offsprings were positive) and 24% through female (48 out of the 202 offsprings were positive). Thus, the results was similar to that obtained by the mating of F1(chimera x MCH) x MCH(ICR). F3 (F2 x MCH) were obtained by the latter mating.

**[0329]** The concentrations of human antibody κ chain in the sera of 4-12-week old chimeric mice (Fig. 43, indicated as "Chimera", 4 mice), F1 (chimera x MCH) (19 mice), F2 (F1 x MCH) (39 mice) and F3 (F2 x MCH) (33 mice) were determined in the same manner as in Example 44. The results are shown in Fig. 43. Human antibody κ chain was detected in all of the mice retaining W23 fragment. On the other hand, the κ chain concentrations varied greatly in F2 (F1 x MCH) and F3(F2 x MCH); the averaged values in these groups were lower than those in the chimeric mice and F1(chimera x MCH).

**[0330]** In order to examine the influence which would be caused by the mating with a strain other than MCH(ICR), the same F2(F1 x MCH) mice as used in the experiment of mating with MCH(ICR) were mated with C57BL/6N (purchased from CREA JAPAN, INC.). Concentrations of κ chain were determined in the same manner on the resultant 26 mice retaining W23 fragment [F3(F2 x C57BL/6)]. As a result, these mice exhibited κ chain concentrations as high

as those in the chimeric mice and F1(chimera x MCH) (Fig. 43). As described above, F3(F2 x MCH) and F3(F2 x C57BL/6) are derived from the same F2(F1 x MCH) mice as one of the parents. Therefore, it is believed that the difference between F3(F2 x MCH) and F3(F2 x C57BL/6) is in their hereditary background alone. Thus, it is indicated that difference in hereditary background influences the amount of expression of human antibody  $\kappa$  chain. Further, it has become clear that the hereditary background of C57BL/6 is more desirable than that of MCH(ICR) for efficient expression of human antibody  $\kappa$  chain. From similar experiments, it has been demonstrated that the hereditary background of C3H HeN (purchased from CREA JAPAN, INC.) is comparable to or better than that of C57BL/6 for efficient expression of human antibody  $\kappa$  chain.

**[0331]** The following experiment was conducted to examine as to whether the influence of hereditary background on antibody  $\kappa$  chain concentrations observed above is related to the ratio of chromosome retention (stability) at the level of individual mice. Briefly, metaphase chromosome samples were prepared from tail-derived fibroblasts and bone marrow cells of 2F-1 mouse (serum  $\kappa$  chain concentration: 84 mg/l) and 1F-3 mouse (serum  $\kappa$  chain concentration: 13 mg/l) in F1(chimera x MCH) and subjected to FISH analysis (Tomizuka et al., Nature Genetics, vol 16, 133-143). The ratio of those metaphase spreads containing W23 fragment hybridizing to a human chromosome-specific probe to all of the spreads observed was determined. It is believed that the resultant values represent the W23 fragment retention ratios in fibroblasts and bone marrow cells, respectively. As a result, with respect to 2F-1, the retention ratio was 51% in fibroblasts and 34% in bone marrow cells; and with respect to 1F-3, the retention ratio was 23% in fibroblasts and 18% in bone marrow cells (more than 50 nuclear plate were measured for each case). These results suggest that the  $\kappa$  chain concentrations in sera correlated to the ratios of retentions of W23 fragment in fibroblasts and bone marrow cells. In other words, it is very likely that hereditary background influences the stability of the transferred human chromosome fragment itself. Thus, it is believed that the hereditary background of C57BL/6 or C3H strain is desirable for efficient expression of a gene not only on the chromosome #2 fragment described herein but also on other chromosome fragments (e.g. chromosome #14 fragment).

**[0332]** In order to verify the above conjecture, a male mouse #17-7 in F1(chimera x MCH) which retains the human chromosome #14 fragment obtained in Example 68 (hereinafter referred to as "SC20 fragment") and which expresses a human antibody heavy-chain in the serum was mated with MCH(ICR) and C57BL/6. Of the resultant offsprings, two F2(F1 x MCH) mice and two F2(F1 x C57BL/6) mice, each retaining SC20 fragment, were subjected to determination of human antibody heavy-chain concentrations in the sera (see Example 68). Furthermore, metaphase chromosome samples were prepared from the tails of these mice and then the ratio of SC20 fragment retention was determined in the same manner as for W23 fragment. As a result, the human  $\mu$  chain concentration was 11.0 mg/l and 1.1 mg/l and the chromosome retention ratio 74% and 54% in F2(F1 x MCH), whereas the human  $\mu$  chain concentration was 47 mg/l and 54 mg/l and the chromosome retention ratio 84% and 88% in F2(F1 x C57BL/6). F2(F1 x C57BL/6) mice exhibited higher values in both the human  $\mu$  chain concentration and the chromosome retention ratio. Thus, it has become clear that the hereditary background of C57BL/6 is desirable for stable retention of a transferred human chromosome and for efficient expression of a gene located thereon, as presumed from the results obtained on W23 fragment-retaining mice.

#### Example 80

Production of chimeric mice from an antibody heavy-chain-deficient and antibody  $\kappa$  chain-homologous recombinant ES cell clone

**[0333]** Cells in the frozen stock of antibody heavy-chain-deficient and antibody  $\kappa$  chain-homologous recombinant ES cell clone HD43 from Example 58 were thawed, started up for culture and injected into 8-cell stage embryos obtained by mating male and female mice of ICR (CREA JAPAN, INC.); the injection rate was 10-12 cells per embryo. After the embryos were cultured overnight in the medium for ES cells (see Example 9) to develop into blastocysts, about 10 of the injected embryos were transplanted to each side of the uterus of foster mother ICR mice (CREA JAPAN, INC.; 2.5 days after pseudopregnant treatment).

**[0334]** As a result of transplantation of a total of 314 injected embryos, 51 offspring mice were born. Chimerism in the offsprings can be determined by the extent of TT2 cell-derived agouti coat color (dark brown) in the host embryo (ICR)-derived albino coat color (white). Out of the 51 offsprings, 26 mice were recognized to have partial agouti coat color, indicating the contribution of the ES cells. Out of the 26 mice, two were chimeric female mice in which 100% of their coat color was agouti (i.e. ES cell-derived).

**[0335]** From these results, it was confirmed that antibody heavy-chain-deficient and antibody light-chain-homologous recombinant ES cell clone HD43 maintains the ability to produce chimera. In the female mice exhibiting 100% contribution, it is highly possible that the ES cells have been differentiated into functional germ cells (oocytes).

**[0336]** Examination was made as to whether ES cell-derived offsprings would be produced by mating the above female chimeric mice (both having 100% chimerism in coat color) with male ICR mice. By this mating, offsprings with



agouti coat color should be produced from TT2F cell (agouti: dominant)-derived oocytes in the chimeric mice fertilized by male ICR mouse (albino: recessive)-derived sperms, and offsprings with albino coat color should be produced from ICR-derived oocytes. Actually, all of the viable offspring mice obtained by one mating for each female mouse exhibited ES cell-derived agouti coat color. Genomic DNAs were prepared from the tails of these offspring mice to examine the presence of an antibody  $\kappa$  chain disrupted allele by Southern blot analysis (Example 58). As a result, mice having an antibody  $\kappa$  chain disrupted allele were obtained.

[0337] Twenty-seven offspring mice produced by the mating of the antibody light-chain deficient-heterozygote male and female mice were subjected to Southern blot analysis (Example 58). As a result, antibody light-chain wild-type alleles disappeared and only disrupted alleles were observed in 7 offspring mice. Hence, these mice were believed to be antibody light-chain-deficient homozygotes. Fig. 44 shows the results of detection and quantitative determination of mouse antibody  $\kappa$  chain and  $\lambda$  chain in the sera.

[0338] In those mice which were judged to be antibody light-chain-deficient homozygotes by the Southern blot analysis (Nos. 4, 6, 14, 22, 24, 25 and 26 in Fig. 44), the concentrations of  $\kappa$  chain are greatly reduced (the remaining  $\kappa$  chain appears to be derived from their mother mice). Instead, the concentrations of A chain are greatly increased in these mice. These results are consistent with the reported results of analysis of the antibody  $\kappa$  chain knockout mouse (Yong-Rui Zou et al., EMBO J. 12, 811-820 (1993)).

[0339] Thus, an antibody  $\kappa$  chain knockout mouse strain could be established from antibody  $\kappa$  chain homologous recombinant ES cell clone HD43.

#### Example 81

Preparation of a targeting vector for inserting human telomere sequence into human chromosome #22

[0340] Fragmentation of human chromosome #22 on which human antibody  $\lambda$  chain gene (hereinafter referred to as "Ig $\lambda$  gene") was located was attempted by inserting human telomere sequence by homologous recombination (J. E. Itzhaki et al., Nature Genet., 2, 283-287, 1992). Specifically, a targeting vector for inserting human telomere sequence into the LIF locus located very close to Ig  $\lambda$  gene (on the telomere side) was prepared.

[0341] Human telomere sequence was synthesized by PCR according to the method of J. J. Harrington et al. (Nature Genet., 15, 345-355, 1997). The PCR product was purified by agarose gel electrophoresis and then blunted with DNA Blunting Kit (Takara Shuzo). The blunted PCR product was inserted into the Eco RV site of pBluescript SK II(+) (Toyobo) by ligation using DNA Ligation Kit (Takara Shuzo) (pBS-TEL). This plasmid pBS-TEL was sequenced. As a result, it was found that the telomere sequence had been inserted in the following direction: Hind III - (TTAGGG) $n$  - Eco RI.

[0342] Subsequently, the LIF gene region on human chromosome #22 to be used in the homologous recombination was amplified by PCR as described below, and then cloned into plasmid pBS-TEL. The sequences of the primers used in the PCR were as follows.

**Sense primer: 5' - TCGAACTAGTAGGAGAAGTGAACCTGAGGAGGC - 3'**

**(SEQ ID NO: 65)**

**Antisense primer: 5' - TCGAACTAGTGATTTCAGTGATGCTGTGCAGG - 3'**

**(SEQ ID NO: 66)**

[0343] The PCR reaction mixture was composed of 5  $\mu$ l of 10 x LA PCR buffer II (Mg $^{2+}$  free) (Takara Shuzo); 5  $\mu$ l of 25 mM MgCl $_2$ ; 8  $\mu$ l of dNTP mixture (2.5 mM each) (Takara Shuzo); 10 pmol of sense primer; 10 pmol of antisense primer; 100 ng of template DNA (HFL1, genomic DNA from primary culture human fibroblasts); 0.5  $\mu$ l of LA Taq (5 U/ $\mu$ l) (Takara Shuzo) and sterile distilled water to make a total volume of 50  $\mu$ l. All of the operations for preparing the reaction mixture were carried out on ice. Then, reaction tubes were placed in the well of a thermal cycler (PCR System 9600, Perkin-Elmer) preset at 85°C. After the tubes were heated at 94°C for 1 minute, 35 cycles of reaction were carried out at 98 °C for 10 seconds and at 65 °C for 5 minutes. The PCR product was purified and then digested with Spe I (Spe I site was present in the primers), followed by insertion into the Spe I site in pBS-TEL. The plasmid in which the LIF gene had been inserted in a opposite direction of the human telomere sequence (TTAGGG) $n$  was selected (M. Giovannini et al., Cytogenet Cell Genet 64, 240-244, 1993) (pBS-TEL/LIF).

[0344] Subsequently, plasmid pGKpuro containing a puromycin resistance gene (S. Watanabe et al., Biochem. Bi-

ophys. Res. Comm., 213, 130-137, 1995) was digested with Eco RI and blunted, followed by insertion of Not I linker. The puromycin resistance gene was cut out by digesting the resultant plasmid with Not I and then inserted into the Not I site of pBS-TEL/LIF. The plasmid in which the direction of transcription of the puromycin resistance gene was the same as that of the LIF gene was selected (pBS-TEL/LIFPuro, see Fig. 45). The resultant plasmid was amplified in *E. coli* DH5, purified with QUIAGEN column (Funakoshi) and used for transfection (as described later).

#### Example 82

##### Transfer of human chromosome #22 into chicken DT40 cells

**[0345]** Mouse A9 cells containing human chromosome #22 marked with a G418 resistance gene (Tomizuka et al., Nature Genet., vol 16, 133-143, 1997; hereinafter referred to as "A9/#22neo") were cultured in Dulbecco's modified Eagle's Minimal Essential Medium (hereinafter referred to as "DMEM") supplemented with 10% fetal bovine serum and G418 (800 µg/ml). Chicken DT40 cells were cultured in DMEM supplemented with 10% FBS, 1% chicken serum and  $10^{-4}$  M 2-mercaptoethanol.

**[0346]** Microcells were prepared as described below (for details, see Shimizu et al., "Cell Technology Handbook", published by Yodosha, p. 127-).

A9/#22neo cells were cultured in twelve 25 cm<sup>2</sup> centrifuge flasks (Costar) until the cell concentration reached about 90% saturation. Then, the medium was exchanged with a medium (DMEM + 20% FBS) supplemented with COLCEMID (0.07 µg/ml; demecolcine, Wako Pure Chemical Industries, Inc.).

The cells were cultured for another 2.5-3 days to form microcells. Thereafter, the culture solution was removed from the centrifuge flasks, into which a solution of cytochalasin B (10 µg/ml, Sigma) prewarmed at 37°C was filled and centrifuged at 34 °C at 8000 rpm for 1 hour. The microcells were suspended in DMEM and purified by filtration with filters. After the purification, the microcells were centrifuged at 1500 rpm for 10 minutes and then suspended in 5 ml of DMEM. DT40 cells ( $2 \times 10^7$ ) were centrifuged at 1000 rpm for 5 minutes, washed with DMEM twice and suspended in 5 ml of DMEM. The microcells prepared above were recentrifuged at 1500 rpm for 10 minutes and then, without removal of the supernatant, 5 ml of the previously prepared DT40 suspension was overlaid gently. After centrifugation at 1300 rpm for 5 minutes, the supernatant was removed. The cell pellet was suspended in 2 ml of PHA-P (100 µg/ml, DIFCO) and left to stand in an incubator at 37°C under 5% CO<sub>2</sub> for 15 minutes. Then, the suspension was centrifuged at 1700 rpm for 7 minutes. The supernatant was removed and the cell pellet was loosened by tapping. To the cell pellet, 1 ml of PEG1500 (polyethylene glycol, Boehringer) was added gently and the pellet was treated for 1.5-2 minutes under agitation. After this treatment, 1 ml of DMEM was added over approximately 1 minute. Then, 3 ml of DMEM was added over approximately 2 minutes. Thereafter, DMEM was added to make a total volume of 11 ml and the resultant mixture was mixed gently. The mixture was left to stand for 10 minutes at room temperature and then centrifuged at 1300 rpm for 5 minutes. The supernatant was removed. The cells were suspended in 10 ml of the above-described culture medium and cultured in  $\phi$  100 mm plates for 24 hours. Twenty-four hours later, the medium was exchanged with one supplemented with G418 (1 mg/ml). The resultant culture was dispensed into three 24-well plates (Sumitomo Bakelite), followed by selective culture for about 2 weeks to isolate G418 resistant clones.

##### (1) PCR analysis

**[0347]** As a result of the selective culture, about thirty G418 resistant clones were obtained. Genomic DNAs were extracted from these clones using Puregene DNA Isolation Kit (Gentra System). Using the genomic DNA as a template, PCR was performed with human Ig  $\lambda$  gene-specific primers to identify clones having human chromosome #22 containing Ig $\lambda$  gene. The Ig $\lambda$  gene-specific primers used were as follows.

5' - GAGAGTTGCAGAAGGGGTGACT - 3' (SEQ ID NO: 67)

5' - GGAGACCACCAAACCCTCCAAA - 3' (SEQ ID NO: 68)

**[0348]** The PCR reaction mixture was composed of 5 µl of 10 x Ex Taq buffer (Takara Shuzo); 8 µl of dNTP mixture (2.5 mM each) (Takara Shuzo); 10 pmol of each primer; 100 ng of genomic DNA; 0.5 µl of Ex Taq (5 U/µl) (Takara Shuzo) and sterile distilled water to make a total volume of 50 µl. All of the operations for preparing the reaction mixture were carried out on ice. Then, reaction tubes were placed in the well of a thermal cycler (PCR System 9600, Perkin-

Elmer) preset at 85 °C. After the tubes were heated at 94 °C for 1 minute, 35 cycles of reaction were carried out at 98 °C for 10 seconds, at 56 °C for 30 seconds and at 72 °C for 30 seconds. As a result, 2 clones having human Ig λ gene were identified. The presence of polymorphic markers (D22S315, D22S280, D22S283 and D22S274; Polymorphic STS Primer Pair, BIOS; J.E. Collins et al., Nature 377 suppl.: 367, 1995) located on human chromosome #22 were detected in these clones by PCR (Fig. 46). The PCR conditions were the same as used for the detection of human Ig λ gene. Mark "O" indicates that the marker was detected. Mark "X" indicates that the marker was not detected. The diagram at the left side shows the location of each marker on chromosome #22 based on a physical map. From these results, it was suggested that these 2 clones have a almost intact human chromosome #22. As to the other clones, although human Ig λ gene was not detected, some of the polymorphic markers on chromosome #22 described above were detected.

## (2) FISH analysis.

[0349] One of the above 2 clones (clone No. 52-18) was subjected to FISH analysis to examine how the human chromosome #22 actually existed in cells. Basic operations such as preparation of chromosome samples, hybridization and detection were performed according to Tomizuka et al. (Nature Genet. 16, 133-143, 1997). As a probe, human COT-1 DNA (labeled with Rhodamine) was used. As a result of observation of 20-30 spreads, it was confirmed that an almost intact human chromosome #22 was present independently (Fig. 50). Those stained in red are human chromosome #22.

[0350] From these results of analysis, it was thought that chicken DT40 cell clone 52-18 (hereinafter referred to as "DT40/#22neo") has intact human chromosome #22.

## Example 83

### Targeted truncation of human chromosome #22 in chicken DT40 cells

[0351] DT40/#22neo from Example 82 was transfected with plasmid pBS-TEL/LIFPuro prepared in Example 81 and an attempt was made to perform targeted truncation of the human chromosome #22 on the LIF locus.

[0352] DT40/#22neo cells were cultured under the same conditions as described in Example 82 in the presence of G418 (1 mg/ml).  $10^7$  cells were washed with cold PPS once, suspended in 0.5 ml of PBS and placed on ice. Then, 25-30 μg of pBS-TEL/LIFPuro linearized with Eco RI was added to the cells, mixed with a pipette, transferred into an electroporation cuvette (Bio-Rad) and left to stand in ice for 10 minutes. The cuvette was set in a gene pulser (Bio-Rad) and then a voltage of 550 V was applied at a capacitance of 25 μF. After the cuvette was left to stand on ice for 10 minutes, the cells were transferred into 72 cm<sup>2</sup> culture flasks containing the above-described medium and cultured for 24 hours. Twenty-four hours later, the medium was exchanged with a medium supplemented with G418 (1 mg/ml) and puromycin (0.3 μg/ml, Sigma). The resultant culture was dispensed into five to eight 96-well culture plates, followed by selective culture for about 2 weeks to isolate resistant clones.

## (1) PCR analysis

[0353] As a result of the selective culture, about 80 resistant clones were obtained. Genomic DNAs were extracted from these cells as described above and subjected to PCR to identify homologous recombinants in which a human telomere sequence was integrated into the LIF locus. One of the primers was designed such that its sequence was complementary to a part of the LIF gene region which was not contained in the vector (see Fig. 47). The other primer was designed such that its sequence was complementary to a part of the puromycin resistance gene which was contained in the vector. The sequences of the primers are as follows.

Puro.1: 5' - GAGCTGCAAGAACTCTTCCTCAG - 3' (SEQ ID NO.: 69)

LIF1: 5' - ATGACTCTAAGGCAGGAACATCTGTACC - 3' (SEQ ID NO.: 70)

[0354] The PCR reaction mixture was composed of 5 μl of 10 x LA PCR buffer II (Mg<sup>2+</sup> free) (Takara Shuzo); 5 μl of

25 mM MgCl<sub>2</sub>; 8 µl of dNTP mixture (2.5 mM each) (Takara Shuzo); 10 pmol of each primer; 100 ng of template DNA; 0.5 µl of LA Taq (5 U/µl) (Takara Shuzo) and sterile distilled water to make a total volume of 50 µl. All of the operations for preparing the reaction mixture were carried out on ice. Then, reaction tubes were placed in the well of a thermal cycler (PCR System 9600, Perkin-Elmer) pre-set at 85 °C. After the tubes were heated at 94 °C for 1 minute, 35 cycles of reaction were carried out at 98 °C for 10 seconds and at 65 °C for 10 minutes. A 6.3 kb PCR product as shown in Fig. 47 should be detected only in the homologous recombinants of interest. As a result of the PCR, this 6.3 kb band was detected in 8 clones (homologous recombination ratio: about 10%). When this PCR product was digested with Sal I, a cut pattern was obtained in exactly the same way as expected. Thus, it was confirmed that these 8 clones were homologous recombinants.

[0355] Subsequently, whether or not the truncation occurred as expected in these 8 clones was examined by PCR detection of the presence of genes (Ig λ, LIF, MB, IL2RB, CYP2D6, DIA1, ECGF1 and ARSA; J.E. Collins et al., Nature 377 suppl:367, 1995) and polymorphic markers (D22S315, D22S275, D22S280, D22S281, D22S277, D22S278, D22S283, D22S272, D22S282 and D22S274; J.E. Collins et al., Nature 377 suppl:367, 1995) on chromosome #22.

[0356] A part of the primer sequences used is as described below. The remaining primer sequences were the same as used by Tomizuka et al. (Nature Genet. 16, 133-143, 1997). The presence of LIF is evident from the experiment described above.

**Sense primer: 5'-CTGCGTGTGTAATCGTGTCC-3' (SEQ ID NO:71)**

**Antisense primer: 5'-TCTGCTGTGAGTGAACCTGC-3' (SEQ ID NO:72)**

ECGF1

**Sense primer: 5'-AGGAGGCACCTTGGATAAGC-3' (SEQ ID NO:73)**

**Antisense primer: 5'-TCACTCTGACCCACGATACAGC-3' (SEQ ID NO:74)**

[0357] The PCR reaction mixture was composed of 5 µl of 10 x Ex Taq buffer (Takara Shuzo); 8 µl of dNTP mixture (2.5 mM each) (Takara Shuzo); 10 pmol of each primer; 100 ng of genomic DNA; 0.5 µl of Ex Taq (5 U/µl) (Takara Shuzo) and sterile distilled water to make a total volume of 50 µl. All of the operations for preparing the reaction mixture were carried out on ice. Then, reaction tubes were placed in the well of a thermal cycler (PCR System 9600, Perkin-Elmer) pre-set at 85 °C. After the tubes were heated at 94 °C for 1 minute, 35 cycles of reaction were carried out at 98 °C for 10 seconds, at 56 °C (65 °C for CYP2D6 and ECGF1) for 30 seconds and at 72 °C for 30 seconds. The results are shown in Fig. 48. Marks "O" and "X" have the same meanings as described above. As is clear from this Figure, none of the genes and markers located on the telomere side of the LIF locus into which a human telomere sequence had been integrated were detected in clones 67, 68, 328 and 343. It is suggested that truncation by the integration of a telomere sequence did occur as expected in at least those 4 clones.

## (2) FISH analysis

[0358] Whether the human chromosome #22 had been actually truncated or not was examined by FISH analysis. The experimental method was the same as described above. As probes, human COT1 DNA (labeled with Rhodamine) and plasmid pGKPuro (labeled with FITC) were used. By COT1 staining, the human chromosome #22 can be visually checked for truncation in comparison with DT40/#22neo having intact human chromosome #22. Furthermore, if the chromosome #22 is truncated as expected on the LIF locus into which the vector has been integrated, a signal from the Puro probe should be detected at one end of the telomere of the chromosome #22 fragment. A part of the results is shown in Fig. 49. As a result of observation of 20-30 spreads for each clone, a small fragment of human chromosome #22 (red) having a Puro probe-derived signal (yellow green) at one end of the telomere was surprisingly observed in

all of the 8 homologous recombinant clones. As for clones 64, 212, 222 and 305 which were presumed not to have undergone truncation from the results of the PCR analysis, cells having intact chromosome #22 occupied about 10% of all cells.

[0359] These experimental results show that homologous recombinants in which a human telomere sequence has been integrated into the LIF locus can be obtained at an efficiency of about 10% in chicken DT40/#22neo cells and that truncation of the human chromosome #22 has occurred at the integration site in all of the homologous recombinants (efficiency 100%).

#### Example 84

[0360] Preparation of a complete human antibody-producing hybridoma from a chimeric mouse produced by transferring an endogenous antibody heavy-chain and light-chain - deficient mouse ES cells, which retains partial fragments of human chromosomes 14 and 22, into an immunodeficient mouse host embryo

##### (1) Preparation of anti-TNF- $\alpha$ human IgM antibody

[0361] The chimeric mouse CLH13-7 (derived from TT2FES clone LH13, 50% chimerism) as produced in Example 67-(3) was immunized with human TNF- $\alpha$ , thereby producing hybridomas. Human Tumor Necrosis Factor- $\alpha$  (TNF- $\alpha$ , PEPRO TECH EC LTD., 300-01A) dissolved in PBS was mixed with adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc., R-700) to prepare 0.025 mg/ml of a TNF- $\alpha$  solution. 0.2ml of TNF- $\alpha$  solution was used for immunization by intraperitoneal injection. Immunization has been performed for 70 days at an interval of 1 or 2 weeks. Two weeks after final immunization, the chimeric mouse was immunized with 0.2 ml of 0.025 mg/ml TNF- $\alpha$  dissolved in PBS. Three days after final immunization, the spleen was excised from the chimeric mouse, followed by cell fusion with PEG according to Example 24, thereby producing hybridomas. The fused cells were diluted to achieve 250 thousand spleen cells / ml in a medium (Sanko Junyaku Co., Ltd. S-Clone Cloning Medium CM-B) supplemented with 1 mg/ml G418, 2 % fetal calf serum, and 5% HCF (Air Brown). 100  $\mu$ l of the diluted cells was added to each well of a 96-well plate and cultured. On day 14 after culturing, colonies appeared in about 40% of the wells. The culture supernatant was analyzed by ELISA according to Example 14, followed by screening for a human antibody-producing hybridoma. TNF- $\alpha$  was diluted to 0.5  $\mu$ g/ml with carbonic acid buffer (Sigma, C-3041), and then dispensed, 50  $\mu$ l each well, into a 96-well plate (NUNC, 442404) for coating. Detection was made by TMBZ (DAKO, S1599) using biotin-labeled anti-human immunoglobulin  $\lambda$  chain antibodies (Vector, BA-3070) and an ABC kit (Vector, PK4000). The results were determined using an absorbance as an index, approximately twice or more intense than the negative control. Cells in the positive wells were transferred into a 96-well plate, and then cultured using a medium containing IMDM supplemented with 1 mg/ml G418 and 10% FBS. The culture supernatant was analyzed by ELISA. TNF- $\alpha$  was diluted to 0.5  $\mu$ g/ml with carbonic acid buffer, and then dispensed 50  $\mu$ l per well of a 96-well plate for coating. Detection was made by TMBZ using peroxidase-labeled anti-human immunoglobulin  $\mu$  chain antibody (Biosorce, AH10604). The results were determined using an absorbance as an index, approximately twice or more intense than the negative control. The cells in two positive wells showing strong coloration were cloned by the limiting dilution method in the same manner as in Example 66, thereby obtaining hybridomas producing complete human antibodies that bind to TNF- $\alpha$  and have human Ig $\mu$  and Ig $\lambda$ .

##### (2) Preparation of anti-TNF- $\alpha$ human IgM antibody

[0362] The chimeric mouse CLH13-13 (derived from TT2FES clone LH13, 50% chimerism) as produced in Example 67-(3) was immunized with human TNF- $\alpha$ , thereby producing hybridomas. Human Tumor Necrosis Factor- $\alpha$  (TNF- $\alpha$ , PEPRO TECH EC LTD., 300-01A) dissolved in PBS was mixed with adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc.) to prepare 0.025 mg/ml TNF- $\alpha$  solution. 0.2ml of the TNF- $\alpha$  solution was used for immunization by 5 times intraperitoneal injection at 1 week interval. Two weeks after the fifth immunization, the TNF- $\alpha$  solution was used to boost the immunization. Two weeks after boosting, the chimeric mouse was immunized with 0.025 mg/ml TNF- $\alpha$  solution dissolved in PBS. The blood was collected every week, and the anti-TNF- $\alpha$  human IgY concentration in serum was detected by ELISA according to Example 14. As shown in Fig. 50, an increased human IgY antibody titer against TNF- $\alpha$  was shown. Three days after final immunization, the spleen was excised from the chimeric mouse, followed by cell fusion with PEG according to Example 24, thereby producing hybridomas. The fused cells were diluted to achieve 250 thousand spleen cells / ml in a medium (Sanko Junyaku Co., Ltd. S-Clone Cloning Medium CM-B) supplemented with 1 mg/ml G418, 2 % fetal calf serum, and 5% HCF (Air Brown). 50  $\mu$ l of the diluted cells was added to each well of a 96-well plate and cultured. On day 14 after culturing, colonies appeared in about 60% of wells. The culture supernatant was analyzed by ELISA according to Example 14 followed by screening for a human antibody-producing hybridoma. TNF- $\alpha$  was diluted to 0.5  $\mu$ g/ml with carbonic acid buffer, and then dispensed, 50  $\mu$ l per well, into a 96-well plate

for coating. Detection was performed by TMBZ using peroxidase-labeled anti-human immunoglobulin  $\gamma$  chain antibody (Sigma, A-0170). The results were determined using an absorbance as an index, approximately twice or more intense than the negative control. Cells in the positive wells were transferred into a 96-well plate, and then cultured in a medium containing IMDM supplemented with 1 mg/ml G418 and 10% FBS. The culture supernatant was analyzed by ELISA. TNF- $\alpha$  was diluted to 0.5  $\mu$ g/ml with carbonic acid buffer, and then dispensed, 100  $\mu$ l to each well of a 96-well plate for coating. Detection was made by TMBZ using peroxidase-labeled anti-human immunoglobulin  $\lambda$  chain antibody (Southern Biotechnology Associates Inc., 2070-05). The results were determined using an absorbance as an index, approximately twice or more intense than the negative control. The presence of complete human antibodies containing human Ig $\gamma$  and Ig $\lambda$  was confirmed in two wells.

The result suggests that in a chimeric mouse produced by transferring endogenous antibody heavy chain- and light chain-deficient mouse ES cells retaining partial fragments of human chromosomes 14 and 22 into a host embryo of an immunodeficient mouse, increases the antibody titer of antigen-specific human Ig $\gamma$  against boosting with TNF- $\alpha$  antigen. Furthermore, it was confirmed that the hybridoma producing a complete human antibody IgM or IgG specific to TNF- $\alpha$  can be obtained from these chimeric mice.

### (3) Preparation of human IgM antibody against sugar chain antigen

**[0363]** The chimeric mice CLH13-10 (derived from TT2FES clone LH13, 60% chimerism) and CLH13-18 (derived from TT2FES clone LH13, 30% chimerism) produced in Example 67-(3) were immunized with ganglioside. An adjuvant (MPL + TDM Emulsion, RIBI Immunochem Research Inc., R-700) was dissolved in 2 ml of the mixture of chloroform and methanol (2:1). 1ml of each solution was mixed with 1 mg of GM2 (Sigma, G8397) dissolved in 1 ml of a mixture of chloroform and methanol (2:1) or with 1 mg of ASIALO-GM1 (ISOSEPAB, 65/12). The mixture was transferred into a round-bottom flask, and then dried with a rotary evaporator. 1ml of PBS was added to each product, and then the solution was vigorously stirred using a Vortex mixer, thereby preparing a suspension. The suspension containing GM2 and that containing ASIALO-GM1 were mixed in equal quantities. The chimeric mouse CLH13-18 was immunized intraperitoneally, and the chimeric mouse CLH13-10 was immunized subcutaneously, both with 0.2 ml of the mixture three times a week. The blood was collected every week, ELISA was performed according to Example 14, so that an increase in the antibody titer was confirmed. Ganglioside GM2 or ASIALO-GM1 was dissolved in ethanol to 6  $\mu$ g/ml, and then 50  $\mu$ l of each solution was dispensed into a ELISA plate. The solution was allowed to stand for 2 hours at room temperature, and then dried. A control was prepared by adding only ethanol (without ganglioside) to a plate. PBS supplemented with 2 % bovine serum albumin (BSA, Oriental yeast) was added 200  $\mu$ l/well, thereby blocking for 3 hours at room temperature. The sera of the chimeric mice were diluted 1:100 with PBS supplemented with 4 % fetal bovine serum (FBS) after washing. The diluted sera were added 50  $\mu$ l/well, and then allowed to stand at 4 °C overnight. A peroxidase-labeled anti-human IgM antibody (Biosorce, AH10604) was diluted with PBS supplemented with 0.1 % BSA after washing. Next the diluted chimeric mouse sera were added 50  $\mu$ l/well and then allowed to stand overnight at room temperature. TMB was added to the product and reacted for 15 minutes. Then the reaction was stopped with 1N sulfuric acid. Absorbance at 450 nm was measured. Then an increase in the antibody titer was determined by reducing absorbance of a control well. Fig. 51 shows the results concerning anti-ASIALO GM1 human Ig $\mu$  in sera of the chimeric mice CLH13-18; Figure 52 shows the results concerning anti-GM2 human Ig $\mu$  in sera of the chimeric mice CLH13-10. AS shown in these figures, immunization with ganglioside resulted in the increased human Ig $\mu$  antibody titer specific to the antigen. That is, immune response against the sugar chain antigen was confirmed in the human antibody-producing chimeric mice of the present invention.

**[0364]** Three weeks after the third immunization, re-immunization was performed for boosting. Three days after final immunization, the spleen was excised from the chimeric mouse, followed by cell fusion with PEG according to Example 24, thereby producing hybridomas. The human antibodies against the ganglioside GM2 or ASIALO-GM1 obtained in this manner are useful for treatment of HIV or cancer, such as melanoma.

### Example 85

#### Production of mouse strains mainly producing the complete human antibody by mating (B)

**[0365]** A male or female mouse, which retains SC20 fragment, and is homo- or heterozygous for heavy-chain-deficiency, and homo or heterozygous for  $\kappa$  chain deficiency as produced according to Example 73, was mated with a male or female mouse, which retains W23 fragment, and is homo or heterozygous for heavy-chain deficiency, and homo- or heterozygous for  $\kappa$  chain-deficiency as produced according to Example 73. This mating is expected to yield progeny mice (double Tc/KO), which can meet four conditions of retaining SC20 fragment and W23 fragment, and being homozygous for heavy-chain-deficiency and for  $\kappa$  chain-deficiency.

## (1) Analysis of single Tc/KO mice

[0366] First, the following four types of analysis (A) to (D) were performed for a total of 189 offspring produced by the mating above. A search was performed for a mouse (single Tc/KO), which meets only two of the above conditions (retaining no W23 fragment, and being heterozygous or wild type for  $\kappa$  chain-deficiency). In these individuals, B lymphocytes were developed by function of human heavy-chain introduced instead of mouse heavy-chain that had been deleted. In addition, most of the antibody molecules in sera were thought to contain human heavy-chain.

(A) PCR was performed for DNA prepared from individual mouse to study the retention of SC20 fragment in the same manner as in Example 68-(4). SC20 fragment-specific D14S543 was used as a primer.

(B) Southern blotting was performed to study endogenous heavy-chain deficiency in the same manner as in Example 49.

(C) PCR was performed for DNA prepared from individual mouse to study the retention of W23 fragment in the same manner as in Example 1. W23 fragment-specific D2S1331 (Tomizuka et al., Nature Genetics, 1997, supra) was used as a primer.

(D) Southern blotting was performed to study endogenous  $\kappa$ -chain deficiency in the same manner as in Example 58.

[0367] Twenty out of 189 offspring were determined as single Tc/KO mice by this analysis.

[0368] 50  $\mu$ l of blood was collected from the single Tc/KO mouse HK83 (5 months old) and then 2  $\mu$ l of 0.5 M EDTA (pH 8.0) was added to prevent the blood from clotting. The blood sample was transferred into a centrifugation tube. Then 400  $\mu$ l of sterile water was added to the tube. After 30 seconds, 2 x PBS containing 10 % fetal calf serum (FCS) was added to the solution, followed by centrifugation using a SEROMATIC-2 (KA2200, Kubota). After centrifugation, the supernatant was removed, and then the precipitate was suspended in the remaining small amount of solution. Staining Medium (SM: 1 mM EDTA, 5 % FCS, PBS containing 0.05 % sodium azide) was added to the suspension, followed by recentrifugation. The precipitate was suspended in approximately 10  $\mu$ l of the remaining SM after removal of the supernatant, 1  $\mu$ l of Fc-block (Pharmingen, 01241A, anti-mouse CD32/CD16) was added to the suspension, and then the product was allowed to stand on ice for 1 hour. After 1 hour, 1  $\mu$ l of each PE-labeled anti-mouse CD45R/B220 (Pharmingen, 01125A) and FITC-labeled anti-human IgM (Pharmingen, 08074D) was added, and then the product was allowed to stand for 1 hour. Thus stained cells were washed twice with SM as described above. Finally the cells were suspended in 200  $\mu$ l of SM, thereby preparing stained samples of peripheral blood nuclear cells. Flow cytometry analysis was made using a FACSVantage (Becton Dickinson). Figure 53 shows the results. B220 positive cells, that is, B lymphocytes were absent in mouse antibody heavy-chain-deficient (KO) mice ( $\Delta H/\Delta H$  in the figure); but in single Tc/KO mice ( $\Delta H/\Delta H$ , SC20 in the figure), the presence of a cell population of B lymphocytes which were positive for both B220 and human  $\mu$  chain (h $\mu$  in the figure). That is, it was shown in single Tc/KO mice that introduction of SC20 fragment rescued the deficiency of B lymphocytes.

[0369] Blood was collected from 8 single Tc/KO mouse individuals, HK7 (18 week-old), HK38 (13 week-old), HK45 (13 week-old), HK47 (13 week-old), HK50 (13 week-old), HK61 (13 week-old), HK78 (17 week-old), and HK83 (17 week-old). The concentration of human  $\mu$  chain and of human  $\gamma$  chain in sera were measured by ELISA as described in Example 14. Mouse  $\mu$  chain was detected by the method shown in Example 75 and mouse  $\gamma$  chain by the following method for the sera collected as control from four individuals (litters born from the same mother as that of single Tc/KO mice; 13 to 18 week-old) that were heterozygous for antibody heavy-chain deficiency and heterozygous or wild type for antibody  $\kappa$  chain deficiency (containing no human chromosomal fragment). The concentration of mouse  $\gamma$  chain was determined by ELISA according to Example 14. Anti-mouse immunoglobulin  $\gamma$  chain antibodies (Sigma, I4280) diluted with PBS were coated over a 96-well microtiter plate. Then the serum samples diluted with PBS containing fetal bovine serum (FBS), and peroxidase-labeled anti-mouse immunoglobulin  $\gamma$  chain antibodies (Caltag, M30107) were added in order to incubate. The plate was washed, and then TMBZ (SUMITOMO BAKELITE COMPANY LIMITED, ML-1120T) was added as a peroxidase substrate, so that enzyme activity was evaluated with an absorbance at 450 nm. The results were compared with mouse immunoglobulin IgG (Pharmingen, 03171D), having a  $\gamma$  chain with a known concentration in its purified state and sequentially diluted with PBS containing FBS, so as to determine the concentration of mouse immunoglobulin in sera. Figure 54 shows the results. The mean value of the measurements for each Ig is also shown in this figure. The mean concentration of human  $\mu$  chain in single Tc/KO individuals was larger than that of mouse  $\mu$  chain in antibody heavy-chain-deficient heterozygous individuals. Similarly, the concentration of mouse  $\mu$  chain and of mouse  $\gamma$  chain were measured for single Tc/KO individuals. No mouse  $\mu$  chain was detected (1  $\mu$ g/ml or less) in all the single Tc/KO mice; the mean concentration of mouse  $\gamma$  chain was approximately one tenth of that of human  $\gamma$  chain. These results suggested that in sera of single Tc/KO mice, most of antibody molecules contained human heavy-chain.

[0370] The concentration of human  $\gamma$  chain subclass in sera was measured for the serum samples from four out of 7 said single Tc/KO mice by ELISA as described in Example 29. Figure 55 shows the results. Four types of  $\gamma$  subclass

were detected in all single Tc/KO mice.

[0371] 0.5 ml of adjuvant (TiterMaxGold, Cytex corporation, G-1) was mixed with 0.5 ml of human serum albumin (HSA, Sigma, A3782) which had been dissolved at the concentration of 0.25 mg/ml in PBS. Two single Tc/KO mice, HK45 (5 months old) and HK108 (3.5 months old), were immunized subcutaneously with 0.1 ml of the mixture. Blood was collected from these mice on day 0, 8, and 15 after immunization. The anti-HSA-human  $\gamma$  chain antibody titer in sera was measured by ELISA as described in Example 21. On day 15 after immunization of the two single Tc/KO mice (immunized with HSA), a significant increase in the anti-HSA-human  $\gamma$  chain antibody titer was observed. Thus, it was shown that in the single Tc/KO mouse, antigen specific antibodies (in this case, consisting of human  $\gamma$  chain and mouse  $\kappa$  chain or mouse  $\lambda$  chain) were produced against proteins derived from human for immunization.

### (2) Analysis of double Tc mice

[0372] The following four types (A) to (D) of analysis were made for a total of 189 offspring born. A search was done for mouse mice (double Tc), which meet at least \*\*\* conditions and express human heavy-chain and human  $\kappa$  chain simultaneously.

(A) To study the retention of SC20 fragment, PCR analysis was performed for DNA prepared from the individual mice with the same method as shown in Example 68-(4). The primer used herein was SC20-specific D14S543.

(B) To study the retention of W23 fragment, PCR analysis was performed for DNA prepared from the individual mice with the same method as shown in Example 68-(4). The primer used herein was W23 fragment-specific D2S1331 (Tomizuka et al., Nature Genetics, 1997, supra).

(C) Blood was collected from 3 to 6 week-old mice to confirm the expression of human  $\mu$  chain, and ELISA was performed according to the method shown in Example 68-(4).

(D) Blood was collected from 3 to 6 week-old mice to confirm the expression of human  $\kappa$  chain, and ELISA was performed according to the method shown in Example 20.

[0373] Twenty out of 189 mice were positive in all of the four types of analysis, and determined to be double Tc mice. Hence, it was confirmed that SC20 and W23 fragments were simultaneously retained in an individual mouse and their functions were expressed simultaneously.

### (3) Analysis of double Tc/KO mice

[0374] DNAs of 20 double Tc mice were subjected to Southern blotting as described in Examples 49 and 58 in order to analyze the state of mouse heavy-chain gene and mouse  $\kappa$  chain gene deficiency. The results were 6 mice homozygous for both antibody heavy-chain-deficiency and  $\kappa$  chain deficiencies. Thus 6 mice were homozygous for mouse heavy-chain and  $\kappa$  chain deficiencies and determined to be double Tc/KO mice.

[0375] The peripheral blood nuclear cells prepared as described in Example 85-(1) from two double Tc/KO mice, HKD5 (9 week-old) and HK190 (6 week-old), were subjected to flow cytometry analysis according to Example 85-(1). Thus, in the two mice, the presence of B lymphocytes which were positive for B220 and for H $\mu$  was confirmed.

[0376] Blood was collected from three double Tc/KO mice, HK77 (17 week-old), HKD4 (6 week-old), and HKD5 (6 week-old), and the concentrations of human  $\mu$  chain and of human  $\gamma$  chain in sera were measured by ELISA as described in Example 14. The mean concentration of human  $\mu$  chain in sera and of human  $\gamma$  chain in sera from the 3 mice were 360 mg/l and 201 mg/l, respectively. Both the concentrations of human  $\mu$  chain and of human  $\gamma$  chain were almost equivalent to that of single Tc/Ko mice shown in Fig. 54. Furthermore, for HK190 (4 week-old) and HK192 (4 week-old) in addition to the above 3 mice, the concentration of human  $\kappa$  chain in sera was detected as described in Example 20 and the concentration of mouse  $\lambda$  chain was detected in the method as described below. The concentration of mouse  $\lambda$  chain was detected by ELISA according to Example 14. Anti-mouse immunoglobulin  $\lambda$  chain antibodies (Caltag, M33600) diluted with PBS were coated over a 96-well microtiter plate, serum samples diluted with PBS containing fetal bovine serum (FBS), and then peroxidase-labeled anti-mouse immunoglobulin  $\lambda$  chain antibodies (Caltag, M33607), were added to the plate for incubation. The plate was washed, and then TMBZ (SUMITOMO BAKELITE COMPANY LIMITED, ML-1120T) was added as a peroxidase substrate to evaluate the enzyme activity with absorbance at 450 nm. The results were compared with that of IgG (Sigma, M6034), having a  $\lambda$  chain with a known concentration in its purified state and sequentially diluted with PBS containing FBS, so as to determine the concentration of mouse immunoglobulin  $\lambda$  chain in sera. Based on the results, first a proportion of the concentration of human  $\kappa$  chain to the concentration of all light-chain (Human  $\kappa$  chain concentration + mouse  $\lambda$  chain concentration) was obtained for individual mice. The mean value of 5 individuals was 60%. Therefore, it was shown that the complete human antibody molecule (human  $\mu$  chain /  $\kappa$  chain or human  $\gamma$  chain /  $\kappa$  chain) expressed dominantly over the hybrid antibody molecule (human  $\mu$  chain / mouse  $\lambda$  chain or human  $\gamma$  chain / mouse  $\lambda$  chain) in sera of these double Tc/KO mice.



[0377] Blood was collected from double Tc/KO mice HKD5 (6 week-old) and the concentration of  $\gamma$  chain subclass in sera was measured by ELISA, similar to Example 85-(1). The results were human  $\gamma$  1 chain: 141 mg/l, human  $\gamma$  2 chain: 61 mg/l, human  $\gamma$  3 chain: 119 mg/l, and human  $\gamma$  4 chain: 8.3 mg/l. All four types of human  $\gamma$  chain subclass were detected.

[0378] 0.5 ml of adjuvant (TiterMaxGold, Cytex corporation, G-1) was mixed with 0.5 ml of human serum albumin (HSA, Sigma, A3782) dissolved at a concentration of 0.25 mg/ml in PBS. Two double Tc/KO mice, HK77 (20 week-old) and HKD5 (10 week-old), were immunized subcutaneously with 0.1 ml of the mixture. Blood was collected from these mice on day 0, 8, and 15 after immunization. The anti-HSA-human  $\gamma$  chain antibody titer in sera was measured by a method similar to that of Example 21, and the anti-HSA-human  $\kappa$  chain antibody titer was measured by ELISA as described in Example 23. The results are shown in figures 56 and 57. On day 15 after immunization of the two double Tc/KO mice (immunized with HSA), a significant increase was observed in both the anti-HSA-human  $\gamma$  chain antibody titer and the anti-HSA-human  $\kappa$  chain antibody titer. Thus, it was shown that in a double Tc/KO mouse, the antigen specific complete human antibody (in this case, containing human  $\gamma$  chain and  $\kappa$  chain) was produced against the proteins derived from human for immunization.

#### (4) Analysis of double Tc/KO mice ( $\lambda$ 1 mutant)

[0379] The mouse  $\lambda$  chain gene was not disrupted in the double Tc/KO mouse (Example 85-1(3)). Thus mouse  $\lambda$  chain protein was still expressed in sera and the concentration thereof was approximately 40% of the whole Ig light-chain. To express human  $\kappa$  chain more efficiently, it is required to inactivate the competitive mouse  $\lambda$  chain gene.

[0380] The presence of mutant mice wherein expression of mouse  $\lambda$  chain is approximately 1/50<sup>th</sup> of that of normal mice is known (Ju et al., J. Immunol., 136: 2684, 1986). Gene analysis of this mouse strain revealed that this was caused by the presence of base substitution in an Ig $\lambda$ 1 chain gene constant region (Ju et al., supra). In fact, almost no  $\lambda$ 1 chain accounting for 80 % or more of the concentration of the whole  $\lambda$  chain (remaining A chains are  $\lambda$  2 chain and  $\lambda$  3 chain) was detected in sera of homozygous mutant mice (herein after referred to as  $\lambda$ 1 mutant) (Ju et al., supra). Hence, it is concluded that  $\lambda$  1 chain, the main constituent of mouse Ig $\lambda$  chain, is inactivated in the  $\lambda$ 1 mutant.

[0381] Double Tc/KO mice ( $\lambda$ 1 mutant) can be obtained by mating Al mutant with double Tc/KO mice described in Example 85-(3). This mouse will show decreased expression of  $\lambda$  chain in serum and increased expression of human  $\kappa$  chain instead.  $\lambda$ 1 mutants were isolated from ICR mice (purchased from CHARLES RIVER JAPAN, INC.) population by the above-mentioned method (Ju et al., supra). The isolated  $\lambda$ 1 mutants were mated with double Tc/KO mice, thereby obtaining double Tc/KO ( $\lambda$ 1 mutant) mice. Concentration of human  $\kappa$  chain and of mouse  $\lambda$  chain in sera of the resultant double Tc/KO ( $\lambda$ 1 mutant) and of double Tc/KO ( $\lambda$ 1 mutant hetero or wild type) as a control were measured (Example 85-(3)). Based on these measurements, first, the percentage by human  $\kappa$  chain concentration of whole light-chain concentration was determined for each mouse. Next, the mean percentage of 5 mice was calculated for the two types of the mouse strains above. The results were 93% for double Tc/KO ( $\lambda$ 1 mutant) and 63% for double Tc/KO ( $\lambda$ 1 mutant hetero or wild type) mice. Furthermore, the mean value of the whole light-chain concentration of 5 mice of each strain was 558 mg/l for double Tc/KO ( $\lambda$ 1 mutant) and 525 mg/l for double Tc/KO ( $\lambda$ 1 mutant hetero or wild type). That is, no significant difference was found between the two.

[0382] These results suggested that the expression level of mouse  $\lambda$  chain decreased in double Tc/KO ( $\lambda$ 1 mutant), but that of human  $\kappa$  chain increased as if it compensated for the decreased level. Therefore, the use of the  $\lambda$ 1 mutant was shown to enable efficient expression of complete human antibody molecules consisting of human  $\kappa$  chain and human heavy-chain.

#### Example 86 Preparation of a HSA-specific human antibody-producing hybridoma from the double Tc/KO mice

[0383] An anti-HSA human antibody-producing hybridoma was obtained from double Tc/KO mice HKD5, for which the antibody titer had increased against HSA immunization in Example 85. On day 30 after the initial immunization, final immunization was performed. 3 days later, hybridomas were produced by the method described in Example 24. The supernatants of approximately 3300 wells, in which G418-resistant colonies had appeared, were analyzed by ELISA (Examples 14 and 61). 11 wells were positive for HSA-specific human  $\mu$  chain; 39 wells were positive for HSA-specific human  $\gamma$  chain. 14 wells out of 39 wells positive for anti-HSA human  $\gamma$  chain were positive for human  $\kappa$  chain, and the remainder was positive for mouse  $\lambda$  chain. In addition, no well was positive for both types of Ig light-chain at the same time. These results suggested that the hybridoma producing HSA-specific complete human antibodies (consisting of  $\gamma$  heavy-chain and  $\kappa$  light-chain) was obtained from the double Tc/KO mice. Moreover, ELISA analysis was performed to detect four types of human  $\gamma$  chain subclass according to Example 61-(4). 7 wells were positive for  $\gamma$ 1, 2 wells positive for  $\gamma$ 2, and 5 wells positive for  $\gamma$ 4. Three typical clones of IgG/ $\kappa$  hybridoma were subjected to limited dilution for subcloning, and then the supernatant was used for measuring affinity. Measurement of the affinity constant using surface plasmon resonance in BIAcore followed by calculation using an attached calculation software resulted in  $1.1 \times 10^{10}$  to  $6.6 \times 10^{10} \text{ M}^{-1}$ . Thus, the anti-HSA human IgG/ $\kappa$  antibody obtained from HKD5 was shown to have

high affinity to HSA.

#### Example 87 Construction of cassette vectors ploxPHyg and ploxPbsr

**[0384]** In the following Examples 87 to 103, production of human artificial chromosomes  $\lambda$ -HAC and  $\kappa$ -HAC, having human antibody  $\lambda$  light-chain and human antibody  $\kappa$  light-chain gene clusters cloned thereto, will be described. In addition, introduction of each of the produced HAC into a mouse, expression of a human antibody gene contained in HAC in a mouse, and transfer of HAC to chimera mouse offspring. The outline of the system for producing HAC-introduced mouse disclosed herein is shown in Figs. 58 and 59.

**[0385]** Cassette vectors ploxPHyg and ploxPbsr were constructed as follows for insertion of a loxP sequence, a recognition sequence of Cre recombinant enzyme, onto a human chromosome. For positive selection of cells carrying translocation as expected, the two cassette vectors were constructed such that the former contained PGK promoter and the latter contained GFP gene to be transcribed by this PGK promoter.

First, the construction of ploxPHyg will be described. A plasmid pBluescript II SK(-) (TOYOBO) was cleaved with a restriction enzyme EcoRV (Boehringer). The product was allowed to react for 30 minutes at 50 °C using dephosphorylase CIAP (alkaline dephosphorylase derived from the small intestine of a calf, TAKARA SHUZO CO., LTD.), thereby dephosphorylating the cleaved ends. DNA fragments were excised from a plasmid pBS302 (GIBCO) using restriction enzymes SpeI and HindIII (Boehringer) and then blunt-ended with a DNA blunting kit (TAKARA SHUZO CO., LTD.). Then the DNA fragment was ligated using a DNA Ligation kit (TAKARA SHUZO CO., LTD.), so that a competent cell DH5 (TOYOBO) was transformed, thereby obtaining a plasmid pBS302HS. Blunt-ending, ligation and transformation were conducted following protocols attached to each kit.

Next, the plasmid was cleaved with a restriction enzyme Sall (Boehringer), and then dephosphorylated. To this plasmid, PGK promoter fragments excised from a plasmid pGKPuro (WHITEHEAD INSTITUTE, distributed by Dr. Peter W. Laird) with restriction enzymes Sall and XhoI (Boehringer) were cloned, thereby obtaining a plasmid pBSPGK302HS. The plasmid was cleaved with restriction enzymes EcoRI and NotI (Boehringer), blunt-ended, and dephosphorylated. Then a NotI linker (TAKARA SHUZO CO., LTD.) was ligated to the plasmid, thereby obtaining a plasmid pBSPGK302HSN.

On the other hand, a hygromycin B-resistant gene cassette was excised from a plasmid #1-133 (distributed by Shunichi Takeda, Professor, Medical School, Kyoto University) with a restriction enzyme BamHI (Boehringer), and then the ends were blunted. The plasmid pBSPGK302HSN was cleaved with a restriction enzyme Sall (Boehringer) and blunt-ended. Next the hygromycin B-resistant gene cassette was cloned, thereby obtaining a plasmid ploxPHyg.

Next, construction of ploxPbsr will be described. First, SfiI linker was inserted into the SacI site of a plasmid pBluescript II SK(-) (TOYOBO) as described above. The SfiI linker had been prepared by synthesizing oligo DNA with the following sequence and phosphorylating the 5' end (synthesized by GREINER JAPAN).

(SfiI linker)

5' - GGCCGC [A/T] GCGGCC - 3' (SEQ ID NO: 75)

The plasmid was cleaved with a restriction enzyme BamHI (Boehringer), and dephosphorylated. Then a DNA fragment excised from a plasmid pBS302 (GIBCO) with a restriction enzyme BamHI (Boehringer) was cloned, thereby obtaining a plasmid pBSSfK302B. A SpeI linker (TAKARA SHUZO CO., LTD.) was inserted into the ClaI site of a plasmid pGREEN LANTERN-1 (GIBCO) as described above. A GFP gene cassette fragment excised from a restriction enzyme SpeI (Boehringer) was cloned into the plasmid pBSSfK302B that had been cleaved with SpeI followed by dephosphorylation, thereby obtaining a plasmid pBSSfK302BGFP. This plasmid was cleaved with a restriction enzyme XbaI (Boehringer) and blunt-ended. Then a DNA fragment (blastocidin S-resistant gene cassette) that had been cleaved with a restriction enzyme BamHI (Boehringer) from a plasmid #1-134 (distributed by Shunichi Takeda, Professor, Medical School, Kyoto University) and blunt-ended was cloned, thereby obtaining ploxPbsr. Fig. 60 shows the structure of the cassette vectors ploxPHyg and of ploxPbsr.

#### Example 88 Construction of targeting vectors pHCF2loxPHyg(F) and (R)

**[0386]** Targeting vectors pHCF2loxPHyg(F) and (R) were constructed as follows for insertion of a loxP sequence into a HCF2 locus (closely linked to Ig $\lambda$  region in the centromere side) on human chromosome #22. Forward (F) and reverse (R) targeting vectors were constructed because the transcriptional direction of HCF2 gene was unknown (whether it is from centromere to telomere or from telomere to centromere). First, the genomic region of the human HCF2 locus was amplified by PCR using the following primers:

HCF2 - F2K; 5' - TCGAGGTACCGTGAGAACAAAGACAGAGAATGAGGGAGG - 3'

(SEQ ID NO: 76)

HCF2 - R2K; 5' - TCGAGGTACCTAATGCAGAGGCTCTTTGGTGTACTTGG - 3'

(SEQ ID NO: 77)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, LA Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94 °C for 1 min. Each cycle consisted of 98°C for 10 sec and 68°C for 15 minutes. PCR products were treated with proteinase K (GIBCO) and then subjected to gel filtration with a CHROMA SPIN-TE400 (CLONTECH). Then the product was cleaved with a restriction enzyme KpnI (Boehringer) and subjected to gel filtration with a CHROMA SPIN-TE1000 (CLONTECH). The resulting PCR fragment was cloned into the KpnI site of a plasmid pBluescriptII, thereby obtaining pHCF2. Next the DNA fragment containing a loxP was cleaved with restriction enzymes KpnI and NotI (Boehringer) from a cassette vector ploxPHyg, blunt-ended, and then cloned into the SnaBI site of the plasmid pHCF2. The product wherein the direction of the loxP sequence was same as that of the cloned HCF2 gene fragment was designated as (F), and the product with that in the opposite direction was designated as (R) [pHCF2loxPHyg(F) and (R), Figs. 61 and 62].

Example 89 Construction of a targeting vector pRNR2loxPbsr

[0387] A targeting vector pRNR2loxPbsr was constructed as follows for insertion of a loxP sequence into a RNR2 locus on a human chromosome #14. First, the genomic region of the human RNR2 locus was amplified by PCR using the following primers:

RNR2 - F10E; 5' - TCGAGAATTTCAGTAGCTGGCACTATCTTTTGGCCATC -

3' (SEQ ID NO: 78)

RNR2 - R10E; 5' - TCGAGAATTCGGAGAAAGAACACACAAGGACTCGGTC - 3'

(SEQ ID NO: 79)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, LA Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94°C for 1 min. Each cycle consisted of 98 °C for 10 sec, and 68 °C for 15 min. PCR products were treated with proteinase K (GIBCO) and then subjected to gel filtration with a CHROMA SPIN-TE400 (CLONTECH). Then the product was cleaved with a restriction enzyme EcoRI (Boehringer) and then subjected to a CHROMA SPIN-TE1000 (CLONTEC). On the other hand, the KpnI site of a plasmid pBluescriptII was deleted by cleaving the site with a restriction enzyme KpnI (Boehringer), blunting and self-ligation. Then a SrfI linker was inserted into the NotI site to construct a plasmid vector [pBS (K) Sr]. In addition, oligo DNA with the following sequence was used as the SrfI linker.

SrfI linker; 5' - GCCCGGGC - 3' (SEQ ID NO: 80)

Next a plasmid pRNR2 was constructed by cloning the PCR fragment of the above RNR2 gene into the EcoRI site of the plasmid [pBS(K)Sr].

Moreover, a KpnI linker (TAKARA SHUZO CO., LTD.) was inserted into the SfiI site of the cassette vector ploxPbsr, a DNA fragment containing a loxP sequence was cleaved with a restriction enzyme KpnI (Boehringer), then the fragment was cloned into the KpnI site of the plasmid pRNR2. The RNR2 gene was reported to be transcribed in the direction from telomere to centromere (R.G. Worton et al., SCIENCE, 239:64-68, 1988). Thus, the product wherein the direction

of the cloned PCR fragment of the RNR2 gene was opposite to that of the loxP sequence was used as a targeting vector (pRNR2loxPbsr, Fig. 63).

#### Example 90 Construction of a targeting vector pYHZloxPHyg

**[0388]** A targeting vector pYHZloxPHyg was constructed as follows for insertion of a loxP sequence into the genomic region (located approximately 30 kb away from the Igx region in the direction of the centromere side) cosYHZ304 (obtained from Shimizu, Professor, School of Medicine, Keio University) on human chromosome #2. First, the human cosYHZ304 genomic region was amplified by PCR using the following primers:

YHZ - F2B; 5' - TCGAGGATCCGATAGAGAGATTGTCTTAAATGGGTGGG - 3'  
(SEQ ID NO: 81)

YHZ - R2B; 5' - TCGAGGATCCAACAGCTGGAAGTCATAAAAGCATAGC - 3'  
(SEQ ID NO: 82)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, LA Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94 °C for 1 min. Each cycle consisted of 98°C for 10 sec, and 68°C for 15 min. PCR products were treated with proteinase K (GIBCO) and then subjected to gel filtration with a CHROMA SPIN-TE400 (CLONTECH). Then the product was cleaved with a restriction enzyme BamHI (Boehringer) and subjected to gel filtration using a CHROMA SPIN-TE1000 (CLONTECH). Moreover, the NotI site of the plasmid pBluescriptII was deleted by cleaving the site with a restriction enzyme NotI (Boehringer), blunting and self-ligation. Then a SrfI linker was inserted into the SacII site, so as to construct a plasmid vector [pBS(N)Sr]. A plasmid pYHZ was constructed by cloning the PCR fragment of the above cosYHZ304 genomic region into the BamHI site of the plasmid [pBS(N)Sr]. The plasmid pYHZ was cleaved with a restriction enzyme Tth111I (TAKARA SHUZO CO., LTD.) and blunted. Then a NotI linker (TAKARA SHUZO CO., LTD.) was inserted into the product, thereby obtaining pYHZN. In addition, Furthermore, a NotI linker (TAKARA SHUZO CO., LTD.) was inserted into the KpnI site of the cassette vector ploxPHyg, a DNA fragment containing a loxP sequence was cleaved with a restriction enzyme NotI (Boehringer), and then the resultant fragment was cloned into the NotI site of the plasmid pYHZN. The direction of the cosYHZ304 sequence is known to be from telomere to centromere according to Shimizu, Professor, School of Medicine, Keio University. Thus, the product wherein the direction of the cloned PCR fragment of the cosYHZ304 genome was opposite to that of the loxP sequence was used as a targeting vector (pYHZloxPHyg, Fig. 64).

#### Example 91 Construction of a cassette vector pTELPuro

**[0389]** A cassette vector pTELPuro was constructed as follows for insertion of a human telomere sequence onto a human chromosome. The human telomere sequence was synthesized by PCR according to J.J.Harrington et al. (Nature Genet., 15, 345-355, 1997) and then cloned into the EcoRV site of a plasmid pBluescript II SK(-) (TOYOBO), thereby obtaining a plasmid pTEL. After the EcoRI site of a plasmid pGKPuro (distributed by WHITEHEAD INSTITUTE, Dr. Peter W. Laird) was changed to the NotI site, the DNA fragment (puromycin-resistant gene cassette) cleaved with a restriction enzyme NotI (Boehringer) was cloned into the NotI site of the plasmid pTEL, thereby obtaining a cassette vector pTELPuro (Fig. 65).

#### Example 92 Construction of targeting vectors pTELPuroCD8A(F) and (R)

**[0390]** Targeting vectors pTELPuroCD8A (F) and (R) were constructed as follows for insertion of a human telomere sequence into a CD8A locus (closely linked to the Igx region in the telomere side) on human chromosome #2. Forward (F) and reverse (R) targeting vectors were constructed because the transcriptional direction of the CD8A gene had been unknown (whether it is from centromere to telomere or from telomere to centromere). First, the genomic region of the human CD8A locus was amplified by PCR using the following primers:

CD8A-F; 5' - TCGAGGATCCCTTTAGTGAAGGCAAAGGAAGGGACACTC - 3'

(SEQ ID NO: 83)

CD8A-R; 5' - TCGAGGATCCTGTAAAGGGGTAGCCTGTCCTCTTTCATG - 3'

(SEQ ID NO: 84)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, LA Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94°C for 1 min. Each cycle consisted of 98 °C for 10 sec, and 65 °C for 10 min. PCR products were treated with proteinase K (GIBCO) and then subjected to gel filtration with a CHROMA SPIN-TE400 (CLONTECH). Then the product was cleaved with a restriction enzyme BamHI (Boehringer) and subjected to gel filtration with a CHROMA SPIN-TE1000 (CLONTECH). The resulting PCR fragment was cloned into the BamHI site of the plasmid pTELPuro. Next the cloned CD8A genomic fragment, whose direction was same as that of the human telomere sequence, was determined as (F), the fragment whose direction was opposite to the cloned CD8A genomic fragment as (R) [pTELPuroCD8A(F) and (R), Figs. 66 and 67].

Example 93 Site-directed insertion of a loxPHyg cassette onto human chromosome #22 in a chicken DT-40 cell

[0391] Insertion of the loxPHyg cassette to the HCF2 locus was attempted by transfecting the targeting vectors pHCF2loxPHyg(F) and (R) constructed as in Example 88 into a chicken DT-40 cell retaining human chromosome #22 fragment in which telomere-directed truncation occurred at the LIF locus (Kuroiwa et al., Nucleic Acid Research, 26: 3447-3448, 1998).

[0392] Chicken DT-40 cells were cultured in a RPMI1640 medium (GIBCO) supplemented with 10% fetal bovine serum (GIBCO, herein after referred to as FBS), 1% chicken serum (GIBCO), and 10<sup>-4</sup>M2-mercaptoethanol (Sigma). Approximately 10<sup>7</sup> cells were washed once with an additive-free RPMI1640 medium, and suspended in 0.5 ml of an additive-free RPMI1640 medium. Twenty five to 30 µg of the targeting vectors pHCF2loxPHyg (F) and (R) linearized with a restriction enzyme NotI (Boehringer) were added to the suspension, and the mixture was transferred to a cuvette for electroporation (Bio-Rad) and allowed to stand for 10 minutes at room temperature. The cuvette was set in a gene pulsar (Bio-Rad), and was impressed with voltage (550 V and 25 µF). The product was allowed to stand for 10 minutes at room temperature, and cultured for 24 hours. After 24 hours, the medium was exchanged with that containing hygromycin B (1 mg/ml). The product was dispensed into five 96 well culture plates, followed by selection culture for approximately 2 weeks. Genome DNAs were extracted from the hygromycin B-resistant clones using a Puregene DNA Isolation Kit (Gentra System). Genome DNAs were cleaved with restriction enzymes HpaI and XhoI (Boehringer), subjected to electrophoresis in 0.8% agarose gel, followed by alkaline blotting onto a cellulose nitrate filter (DuPont). Southern hybridization was performed on this filter using a HCF2 probe to identify the homologous recombinant. The HCF2 probe was constructed by PCR using the following primers and a human genome as a template. A <sup>32</sup>P - labeled DNA probe was constructed by random priming using a PCR product as a template (Amersham, following the attached protocol).

HCF2-F4; 5' - CACATGACAAGAGCTCAGCG - 3' (SEQ ID NO: 85)

HCF2-R4; 5' - TCTGACTTCCTCATGAGAGCC - 3' (SEQ ID NO: 86)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, Ex Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94°C. Each cycle consisted of 1 min, 98°C for 10 sec, 65°C for 30 sec, and 72 °C for 1 min. For the targeting vector pHCF2loxPHyg(F), a band not less than 20 kb band was detected for the non homologous recombinant, and a 8.7 kb band for the homologous recombinant. For the targeting vector pHCF2loxPHyg(R), a band not less than 20 kb band was detected for the non homologous recombinant, and a 9.5 kb band for the homologous recombinant (Figs. 61 and 62). Southern hybridization results showed that 41 out of 52 clones were the target homologous recombinants in the targeting vector pHCF2loxPHyg (F),

and 28 out of 60 clones were the target homologous recombinants in (R)(called HF and HR clone, respectively).

Example 94 Site-directed insertion of a loxPbsr cassette onto human chromosome #14 in a chicken DT-40 cell

[0393] Insertion of the loxPbsr cassette to the RNR2 locus was attempted by transfecting the targeting vector pRNR2loxPbsr constructed as in Example 89 into a chicken DT-40 cell retaining fragment SC20 of a human chromosome #14 fragment SC20(Kuroiwa et al., Nucleic Acid Research, 26: 3447, 1998; distributed by Shunichi Takeda, Professor, Medical School, Kyoto University).

[0394] As described above, the targeting vector pRNR2loxPbsr linearized with a restriction enzyme SrfI (TOYOBO) was transfected, followed by selection culture for about 2 weeks under the presence of blastcidin S (10 µg/ml). Genome DNAs were extracted from the resistant clones, and then the homologous recombinants were identified by PCR using the following primers (Fig. 63).

RNR2 - 1; 5' - TGGATGTATCCTGTCAAGAGACC - 3' (SEQ ID NO:87)

STOP - 3; 5' - CAGACACTCTATGCCTGTGTGG - 3' (SEQ ID NO:88)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, LA Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94 °C for 1 min. Each cycle consisted of 98 °C for 10 sec, and 65 °C for 5 min. Hence, approximately 2.5 kb of a PCR product was amplified as expected in 8 out of 60 clones. That is, the homologous recombinant was obtained (called R clone).

Example 95 Site-directed cleavage of human chromosome #2 in a chicken DT-40 cell

[0395] Cleavage of chromosome #2 at an insertion site was attempted by transfecting the targeting vectors pTELPuroCD8A(F) and (R) constructed as in Example 92 into a chicken DT-40 cell retaining the full length human chromosome #2 (Kuroiwa et al., Nucleic Acid Research, 26: 3447, 1998; distributed by Shunichi Takeda, Professor, Medical School, Kyoto University) so as to insert a human telomere sequence to the CD8A locus.

[0396] As described above, the targeting vectors pTELPuOCDBA(F) and (R) linearized with a restriction enzyme SrfI (TOYOBO) were transfected, followed by selection culture for about 2 weeks in the presence of puromycin (0.3 µg/ml). Genome DNAs were extracted from the resistant clones, and then the homologous recombinants were identified by PCR using the following primers (Figs. 66 and 67).

For the targeting vector pTELPuroCD8A(F)

CD8A - 3; 5' - GCCCTCATGGAAATCTCCTGGG - 3' (SEQ ID NO:89)

CDPuro - 1; 5' - GCAGCAACAGATGGAAGGCCTC - 3' (SEQ ID NO:90)

For the targeting vector pTELPuroCD8A(R)

CD8A - 2; 5' - GAACAGAAAGCCACTCTTGCTTTCCAT - 3' (SEQ ID NO:91)

CDPuro - 2; 5' - ACCGAGCTGCAAGAACTCTTCCTCAC - 3' (SEQ ID NO:92)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, LA Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94°C for 1 min. Each cycle consisted

of 98°C for 10 sec, and 68°C for 10 min. Hence, PCR products were amplified as expected in 2 out of 95 clones for (F); in 2 out of 160 clones for (R). That is, the homologous recombinants were obtained (called CD clone and CR clone, respectively).

[0397] Next, PCR and FISH analysis were performed to confirm cleavage of chromosome #2 occurred at the CD8A locus in the homologous recombinants, CD and CR clones.

#### (1) PCR analysis

[0398] The presence of the gene and polymorphic markers on chromosome #2 (Fig. 68) was detected by PCR using the following primers. Primers used for detection of D2S373, D2S113, D2S388, D2S1331, D2S134, D2S171, and TPO were manufactured by BIOS.

Vk3 detection primers

Vk3 - F; 5' - CTCTCCTGCAGGGCCAGTCA - 3' (SEQ ID NO:93)

Vk3 - R; 5' - TGCTGATGGTGAGAGTGAAGTC - 3' (SEQ ID NO:94)

Ck detection primers

Ck - F; 5' - TGGAAGGTGGATAACGCCCT - 3' (SEQ ID NO:95)

Ck - R; 5' - TCATTCTCCTCCAACATTAGCA - 3' (SEQ ID NO:96)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, Ex Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94 °C for 1 min. Each cycle consisted of 98 °C for 10 sec, and 56-60 °C for 30 sec, and 72 °C for 30 sec. Fig. 68 shows the result. All markers were detected in the chicken DT-40 cell clones 521D4 retaining the full length human chromosome #2, whereas all the markers located on the telomere side beyond CD8A disappeared in the homologous recombinant CD clone. Furthermore in the homologous recombinant CR clone, all markers were detected.

#### (2) FISH analysis

[0399] To visually determine that human chromosome #2 was cleaved at the CD8A locus, FISH was performed using a pGKPuro probe capable of detecting a puromycin-resistant gene in a targeting vector (Fig. 69), according to Kuroiwa et al's method (Nucleic Acid Research, 26: 3447-3448, 1998). COT1 staining (labeled with rhodamin, red) revealed the chromosome #2 was fragmented in the CD clone (CD10) compared to 521D4. Furthermore, the signal (labeled with FITC, yellow) derived from the pGKPuro probe was detected on the one end of the telomere, suggesting that the CD8A locus, at which the targeting vector had been inserted, is the one end of the telomere of the chromosome #2 fragment. In the CR clone, the signal derived from the pGKPuro probe was detected at 2p12, suggesting that no fragmentation occurred.

[0400] In conclusion, the human chromosome #2 was cleaved at the CD8A locus in the homologous recombinant CD clone. In addition, no fragmentation in the CR clone suggests that the transcriptional direction of the CD8A gene is from centromere to telomere. As described above, telomere-directed truncation in either direction can be applied to determine an unknown transcriptional direction for a locus.

Example 96 Site-directed insertion of a loxPHyg cassette onto human chromosome #2 in a chicken DT-40 cell

[0401] Insertion of the loxPHyg cassette at the cosYHZ304 genomic region was attempted by transfecting the targeting vector pYHZloxPHyg constructed as in Example 90 into the chicken DT-40 cell CD clone retaining the human chromosome #2 fragment in which telomere-directed truncation occurred at the CD8A locus obtained in Example 95 above.

[0402] As described above, the targeting vector pYHZloxPHYg linearized with a restriction enzyme SrfI (TOYOBO) was transfected, followed by selection culture for about 2 weeks in the presence of hygromycin B (1 mg/ml). Genome DNAs were extracted from the resistant clones, and then the homologous recombinants were identified by PCR using the following primers (Fig. 64).

YHZ-2; 5' - TCCTCTTTTTCCTTCCTTGCCTC - 3' (SEQ ID NO:97)

Yhyg-2; 5' - ATTATTTTGGGCGTTGCGTGG - 3' (SEQ ID NO:98)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, LA Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94 °C for 1 min. Each cycle consisted of 98 °C for 10 sec, and 65 °C for 5 min. The homologous recombinant (called Y clone) can be identified by this analysis.

Example 97 Construction of a human artificial  $\lambda$ -HAC having both a human antibody heavy-chain gene cluster and a  $\lambda$  light-chain gene cluster

[0403] To express a complete human antibody stably and efficiently in a mouse as described in the general description, construction of a human artificial chromosome  $\lambda$ -HAC having both a human antibody heavy-chain gene cluster and a  $\lambda$  light-chain gene cluster was attempted by translocating of a human chromosome #22 fragment consisting of HCF2-Ig $\lambda$ -LIF, to the RNR2 locus on a human chromosome #14 fragment SC20 containing human IgH.

[0404] First, a DT40 hybrid cell retaining both the human chromosome #22 fragment and the chromosome #14 fragment SC20 was constructed by cell fusion of the homologous recombinants HF and HR clones obtained in Examples 93 and 94 above with the homologous recombinant R clone.

(1) Construction of a DT40 hybrid cell retaining both the human chromosome #22 fragment and the chromosome #14 fragment SC20.

[0405] The R clone was cultured in a RPMI1640 medium containing blastcidin S (10  $\mu$ g/ml) and HF clone in the same medium containing hygromycin B (1 mg/ml). The two types of clones ( $1-2 \times 10^7$  clones) were mixed, subjected to centrifugation, and washed twice with a serum-free RPMI1640 medium. After complete removal of the remaining medium, 0.5 ml of 50% PEG 1500 (Boehringer) kept at 37 °C was added gently to the product, followed by vigorous mixing for approximately 2 minutes with a pipette. To the mixture, 1 ml of a serum-free RPMI1640 medium was added slowly for 1 minute, and then 9 ml of a serum-free RPMI1640 medium was added slowly for 3 minutes. Then the mixture was allowed to stand for 10 minutes at 37 °C. Subsequently, the mixture was centrifuged at 1200 rpm for 5 minutes, and cultured in an RPMI1640 medium containing serum for 24 to 48 hours. Then the medium was replaced by an RPMI1640 medium containing blastcidin S (10  $\mu$ g/ml) and hygromycin B (1 mg/ml), dispensed into five 24-well culture plates, and then cultured for 3 to 4 weeks. Similarly, the HR and R clones were fused (cell fusion). Genome DNAs were extracted from a hybrid cell (called RHF clone) obtained from cell fusion of the HF and R clones and a hybrid cell (called RHR clone) obtained from the HR and R clones. PCR was performed using the following primers, confirming retention of two chromosomes, human chromosomes 14 and 22.

Human chromosome #14 detection primers

VH3-F; 5' - AGTGAGATAAGCAGTGGATG - 3' (SEQ ID NO:99)

VH3-R; 5' - GTTGTGCTACTCCCATCACT - 3' (SEQ ID NO:100)

Human chromosome #22 detection primers



Igλ-F; 5' - GAGAGTTGCAGAAGGGGTGACT - 3' (SEQ ID NO:101)

Igλ-R; 5' - GGAGACCACCAAACCTCCAAA - 3' (SEQ ID NO:102)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, Ex Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94°C for 1 min. Each cycle consisted of 98°C for 10 sec, 56°C for 30 sec, and 72°C for 30 sec. PCR showed that 6 clones of the RHF clones and 2 clones of the RHR clones were positive for both VH3 and Igλ. In addition, FISH using a human COT1 DNA as a probe revealed that all of these clones retained two individual human chromosomes. The above results suggest that the RHF and RHR hybrid clones retain two chromosomes, human chromosomes 14 and 22.

(2) Site-directed translocation of the human chromosome #22 fragment to the chromosome #14 fragment SC20 in the RHF and RHR hybrid clones

(2)-1 Construction of a vector pBS185hisD stably expressing Cre recombinant enzyme

[0406] As described in the general description, site-directed translocation of a human chromosome is performed using the Cre-loxP system. With this system, the recombination efficiency between non homologous chromosomes is expected to be very low. Thus the following expression vector was constructed to express Cre enzyme stably, not transiently.

[0407] The Cre recombinant enzyme expression vector pBS185 (GIBCO) was cleaved with a restriction enzyme EcoRI (Boehringer), followed by insertion of a BglII linker, thereby obtaining pBS185Bg. A histidinol-resistant gene cassette was excised from a plasmid #1-132 (distributed by Shunichi Takeda, Professor, School of Medicine, Kyoto University) with a restriction enzyme BamHI (Boehringer), and then cloned into the BglII site of the pBS185Bg, thereby obtaining a vector pBS185hisD (Fig. 70).

(2)-2 Site-directed translocation of the human chromosome #22 fragment to the chromosome #14 fragment SC20 in the RHF and RHR hybrid clones using the Cre-loxP system.

[0408] As described above, the Cre recombinant enzyme stable expression vector pBS185hisD linearized with a restriction enzyme KpnI (Boehringer) was separately transfected to the RHF and RHR hybrid clones, and subjected to selection culture for approximately 2 weeks in the presence of histidinol (0.5 mg/ml). Subsequently, the resistant cell population was dispensed into four 6-well culture plates (24 pools). Two pools were randomly selected and cultured to approximately 10<sup>7</sup> cells. The cell pool was suspended in 4 ml of PBS (phosphoric acid buffer) supplemented with 5% FBS and 1 μg/ml propidium iodide (PI), and subjected to analysis with FACS Vantage (Becton Dickinson). As described above, since recombination (translocation) between loxPs results in reconstruction and expression of GFP gene, cells carrying translocation can be detected by FACS. Sorting was repeated 4 times for the cell fraction suspected to be positive for GFP. Culture was conducted after every sorting in a RPMI1640 medium containing hygromycin B (1 mg/ml) in order to remove the cells retaining acentric chromosomes as described below. As a result, GFP-positive cells could be concentrated with a purity of 98 - 99% from the RHF clone, however, none could be concentrated from the RHR clone. It was suggested that the directions of two loxPs were the same as each other in the RHF clone (from centromere to telomere) as shown in Fig. 71 and that translocation resulted in a normal chromosomal structure. However, in the RHR clone, the directions of two loxPs differed from each other so that translocation resulted in a dicentric chromosome having two centromeres and an acentric chromosome lacking a centromere. For example, this can cause no growth in the presence of hygromycin B. In other words, no cells can be concentrated. The results show the specificity of the experiment for translocation with this system. In addition, HCF2 gene is expected to be transcribed in the direction from centromere to telomere.

[0409] Next, whether recombination occurred between loxPs as expected was confirmed by PCR for the two clones (called SF2-21 and SF2-23) cloned by FACS from RHF. Genome DNAs were extracted from the SF2-21 and SF2-23 and PCR was performed using the following primers.

PGK-1; 5' - ATAGCAGCTTTGCTCCTTCG - 3' (SEQ ID NO: 103)

GFP-1; 5' - TTCTCTCCTGCACATAGCCC - 3' (SEQ ID NO: 104)

5 PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, EX Taq (TAXARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94 °C for 1 min. Each cycle consisted of 98 °C for 10 sec, 61 °C for 30 sec and 72 °C for 1 min. As shown in Fig. 72, when the PCR with these primers yields a product of about 600 bp, this suggests that recombination occurred between loxPs. Fig. 73 shows the result. In the RHF clone to which no Cre recombinant enzyme stable expression vector was transfected, no PCR product was obtained, whereas in SF2-21 and SF2-23, a PCR product of approximately 600 bp was obtained.

10 In addition, FISH was performed using a human 14q ter-specific probe (the long arm telomere region of human chromosome #14 was detected, FITC-labeled) and a pGKPUro probe (the long arm telomere region of human chromosome #22 fragment was detected, Rhodamin-labeled). Thus, FITC and rhodamin signals were detected in the telomere regions on both ends of the same chromosome (Fig. 74). Another FISH using a human chromosome #14-specific probe (labeled with rhodamin) and human chromosome #22-specific probe (labeled with FITC) also resulted in detection of signals derived from the both probes on the same chromosome (Fig. 74).

15 [0410] In conclusion, translocation occurred in SF2-21 and SF2-23 as expected and a human artificial chromosome λ-HAC containing both the human heavy-chain gene cluster and the λ light-chain gene cluster on the same chromosome was constructed.

20

Example 98 Construction of a human artificial chromosome κ-HAC containing both human antibody heavy-chain and κ light-chain gene clusters

25 [0411] A human artificial chromosome κ-HAC containing both human antibody heavy-chain and κ light-chain gene clusters can be constructed by translocation of the human chromosome #2 fragment consisting of cosYHZ304-Igk-CD8A to the RNR2 locus on the chromosome #14 fragment SC20 containing human IgH.

[0412] First, a DT40 hybrid cell retaining both the human chromosome #2 fragment and the chromosome #14 fragment SC20 was constructed by cell fusion of the homologous recombinant Y clone obtained in Example 96 above with the homologous recombinant R clone.

30

(1) Construction of a DT40 hybrid cell retaining both the human chromosome #2 fragment and the chromosome #14 fragment SC20.

35 [0413] The R clone was cultured in a RPMI1640 medium containing blastcidin S (10 µg/ml) and the Y clone in the same medium containing hygromycin B (1 mg/ml). The two types of clones (1-2 x 10<sup>7</sup> clones) were mixed, subjected to centrifugation, and washed twice with a serum-free RPMI1640 medium. After complete removal of the remaining medium, 0.5 ml of 50% PEG 1500 (Boehringer) kept at 37 °C was added gently to the product, followed by vigorous mixing for approximately 2 minutes with a pipette. To the mixture, 1 ml of a serum-free RPMI1640 medium was added slowly for 1 minute, and then 9 ml of a serum-free RPMI1640 medium was added slowly for 3 minutes. Then the mixture was allowed to stand for 10 minutes at 37 °C. Subsequently, the mixture was centrifuged at 1200 rpm for 5 minutes, and cultured in a RPMI1640 medium containing serum for 24 to 48 hours. Then the medium was replaced by a RPMI1640 medium containing blastcidin S (10 µg/ml) and hygromycin B (1 mg/ml), dispensed into five 24-well culture plates, and then cultured for 3 to 4 weeks. Genome DNAs were extracted from the hybrid clones (called RY clone), and subjected to PCR using the following primers, thereby confirming the retention of two chromosomes, human chromosomes 14 and 2.

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Human chromosome #14 detection primers

50 VH3 - F; 5' - AGTGAGATAAGCAGTGGATG - 3' (SEQ ID NO:105)

VH3 - R; 5' - GTTGTGCTACTCCCATCACT - 3' (SEQ ID NO:106)

55

Human chromosome #2 detection primers

CK-F; 5' - TGGAAGGTGGATAACGCCCT - 3' (SEQ ID NO:107)

CK-R; 5' - TCATTCTCCTCCAACATTAGCA - 3' (SEQ ID NO:108)

PCR was performed using GeneAmp9600 (Perkin-Elmer) as a thermal cycler, Ex Taq (TAKARA SHUZO CO., LTD.) as a Taq polymerase, and the attached buffer and dNTP (dATP, dCTP, dGTP and dTTP) following the recommended conditions. PCR was performed for 35 cycles following thermal denaturation at 94 °C for 1 min. Each cycle consisted of 98 °C for 10 sec, 56-60 °C for 30 sec, and 72 °C for 30 sec. Furthermore, FISH was performed using human COT1 DNA as a probe to confirm that the two human chromosomes exist independently. The above results can confirm that the RY hybrid clone retains two chromosomes, human chromosomes 14 and 2.

(2) Site-directed translocation of the human chromosome #2 fragment to the chromosome #14 fragment SC20 in the RY hybrid clone

[0414] As described above, the Cre recombinant enzyme stable expression vector pBS185hisD linearized with a restriction enzyme KpnI (Boehringer) was transfected to the RY hybrid clone, and subjected to selection culture for approximately 2 weeks in the presence of histidinol (0.5 mg/ml). Subsequently, the resistant cell population was dispensed into four 6-well culture plates (24 pools). Two pools were randomly selected and cultured to approximately 10<sup>7</sup> cells. The cell pool was suspended in 4 ml of PBS (phosphoric acid buffer) supplemented with 5% FBS and 1 µg/ml propidium iodide (PI), and subjected to analysis with FACS Vantage (Becton Dickinson). Sorting was repeated several times for the cell fraction suspected to be positive for GFP.

Similarly, whether recombination occurs as expected between loxPs in RY-derived clones to be cloned by FACS can be confirmed by PCR using PGK-1 and GFP-1 primers. This can also be confirmed by FISH using a human 14q ter-specific probe (long arm telomere region of the human chromosome #14 is detected, FITC-labeled) and a pGK Puro probe (short arm telomere region of the human chromosome #2 fragment is detected, Rhodamin-labeled). Furthermore, FISH using a human chromosome #14-specific probe and a human chromosome #2-specific probe can also reach a conclusion that a human artificial chromosome κ-HAC containing both the human heavy-chain gene cluster and the κ light-chain gene cluster on the same chromosome is constructed.

Example 99 Introduction of human artificial chromosomes A-HAC and κ-HAC from a DT40 hybrid cell into a Chinese hamster CHO cell

[0415] λ-HAC and κ-HAC were first introduced into a CHO cell by MMCT for the purpose of introducing λ-HAC and κ-HAC into a mouse ES cell.

[0416] SF2-21 and SF2-23 DT40 hybrid cells were separately cultured on 8 Petri dishes with a diameter of 150 mm. When the cells reached a confluent state, the medium was exchanged with a RPMI1640 medium supplemented with 20% FBS, 1% chicken serum, 10<sup>-4</sup>M2-mercaptoethanol, 0.05 µg/ml colcemid, and cultured for another 36 hours to form a microcell. The cells were suspended in 24 ml of a serum-containing RPMI1640 medium, and dispensed 2 ml into each of twelve 25 cm<sup>2</sup> centrifugation flasks precoated with 100 µg/ml poly L-lysine. Then the cells were cultured for 1 hour at 37°C allowing the cells to adhere to the bottom of the flasks. Following removal of the culture solution, cytochalasin B (10 µg/ml, Sigma) solution kept at 37°C was added to a centrifugation flask, then subjected to centrifugation at 8000 rpm for 1 hour at 34 °C. The microcell was suspended in a serum-free DMEM medium, and purified with a 8 µm and 5 µm filters. Following purification, the product was centrifuged at 1700 rpm, for 10 minutes, and then suspended in 5 ml of a serum-free DMEM medium. On the other hand, approximately 10<sup>7</sup> CHO cells were removed by trypsinization. The cells were washed twice in a serum-free DMEM medium, and then suspended in 5 ml of a serum-free DMEM medium. Again, the microcell was centrifuged at 1700 rpm for 10 minutes. Without removing the supernatant, 5 ml of the CHO suspension was layered gently over the supernatant. Following centrifugation, the culture solution was removed, 0.5 ml of 1:1.4 PEG solution [5 g of PEG1000 (Wako Pure Chemicals Co., Ltd) and 1 ml of DMSO (Sigma) were dissolved in 6 ml of DMEM] was added, followed by vigorous stirring with a pipette for approximately 2 minutes. Subsequently, 10 ml of a serum-free DMEM medium was added gently to the mixture for about 3 minutes, and allowed to stand for 10 minutes at 37°C. After centrifugation, the cells were suspended in a F12 medium (GIBCO) supplemented with 10% FBS, dispensed into ten 24-well culture plates, and cultured for 24 hours at 37 °C. Then the medium was exchanged with a F12 medium containing 800 µg/ml G418, followed by selection culture for 3 to 4 weeks.

[0417] Genome DNAs were extracted from the G418-resistant clones. Then the CHO clone retaining λ-HAC was

identified by PCR in the conditions as described above using Ig $\lambda$  and VH3 detection primers, PGK-1, and GFP-1 primers (data not shown). Moreover, FISH was performed using human chromosome #14-specific and 22-specific probes for the clone positive in the PCR above, thereby visually confirming the presence of  $\lambda$ -HAC. These results suggested that a CHO clone retaining  $\lambda$ -HAC was obtained.

[0418] In exactly the same manner, CHO cells retaining  $\kappa$ -HAC can be cloned.

Example 100 Introduction of  $\lambda$  and  $\kappa$ -HAC from a CHO cell into a mouse ES cell

[0419] To produce a chimeric mouse retaining  $\lambda$  and  $\kappa$ -HAC,  $\lambda$  and  $\kappa$ -HAC retained in a CHO cell were introduced to a mouse ES cell by MMCT.

[0420] According to Tomizuka et al.'s method (Nature Genet. 16: 133, 1997), a microcell was purified from the CHO cell retaining  $\lambda$  and  $\kappa$ -HAC of approximately  $10^8$ , then suspended in 5 ml of DMEM. Approximately  $10^7$  mouse ES cells, TT2F, were removed by trypsin treatment, washed three times with DMEM, and suspended in 5 ml of DMEM. Next the suspension was added to the centrifuged microcell, and subjected to centrifugation at 1250 rpm for 10 minutes so as to completely remove the supernatant. The precipitate was mixed well by tapping. Then 0.5 ml of 1:1.4 PEG solution [5g of PEG 1000 (Wako Pure Chemicals Co., Ltd) and 1 ml of DMSO (Sigma) were dissolved in 6 ml of DMEM], followed by stirring well for approximately 1.5 minutes. Subsequently 10 ml of DMEM was slowly added to the product. Then the product was centrifuged for 10 minutes at 1250 rpm, and suspended in 30 ml of an ES medium. The suspension was dispensed into 3 Petri dishes (CORNING) with a diameter of 100 mm containing feeder cells pre-inoculated therein for culturing. 24 hours later, the medium was exchanged with a medium containing 300  $\mu$ g/ml G418, followed by selection culture for approximately 1 week. Genome DNAs were extracted from the drug-resistant colonies, and then PCR was performed in conditions as described above using Ig $\lambda$  or C $\kappa$  and VH3 detection primers. Furthermore, FISH was performed using probes specific to human chromosomes 14 and 22, or 2. In conclusion, the ES cell clone retaining the target human artificial chromosomes  $\lambda$  and  $\kappa$ -HAC was obtained.

Example 101 Production of a chimeric mouse retaining human artificial chromosomes  $\lambda$  and  $\kappa$ -HAC

[0421] A chimeric mouse was produced in the manner as described in Example 10 using the ES cell clone as obtained in Example 100 above. Possible hosts include ICR and MCH (CLEA JAPAN, INC.) or an embryo at the 8-cell cell stage obtained by mating antibody heavy-chain knockout female and male mice as established in Example 67-(1). The coat color of the progeny mice born as a result of transplantation of the injected embryo into surrogate mice determines whether they are chimeric or not (Example 68-(3)). Chimeric mice can be obtained from the ES cell line retaining human artificial chromosome  $\lambda$ -HAC or that retaining  $\kappa$ -HAC. It is suggested that the ES cell line retaining human artificial chromosome  $\lambda$ -HAC or that retaining  $\kappa$ -HAC possesses chimera-forming ability, that is, retaining ability to differentiate into the normal tissue of a mouse.

Example 102 Expression of a complete human antibody in a chimeric mouse retaining human artificial chromosomes  $\lambda$ -HAC and  $\kappa$ -HAC

[0422] Retention of HAC by chimeric mice is as shown by PCR and FISH analysis (Examples 97 and 98) and the method (Tomizuka et al., Nature Genetics, 16, 133-143). The expression of complete human antibody molecules consisting of human Ig $\lambda$  chain, human Ig heavy-chain and human Ig $\lambda$  chain/heavy-chain in a chimeric mouse retaining  $\lambda$ -HAC is confirmed by the method described in Example 65. Moreover, the expression of complete human antibody molecules consisting of human Ig $\kappa$  chain, human Ig heavy-chain and human Ig $\kappa$  chain/heavy chain in a chimeric mouse retaining  $\kappa$ -HAC is also confirmed by the method described in Example 65.

Example 103 Transfer of human artificial chromosomes  $\lambda$ -HAC and  $\kappa$ -HAC from the chimeric mice to its progeny

[0423] Transfer of  $\lambda$ -HAC and  $\kappa$ -HAC from the chimeric mice to its progeny was examined by the method described in Example 68-(4). As a result, the two mouse strains, retaining and transferring  $\lambda$ -HAC and  $\kappa$ -HAC, respectively to its progeny are established. The retention of HAC in these mice strains is shown by PCR and FISH analysis (Examples 97 and 98) and the method (Tomizuka et al., Nature Genetics, 16, 133-143). In addition, the expression of complete human antibody molecules consisting of human Ig $\lambda$  chain, human Ig heavy-chain and human Ig $\lambda$  chain/heavy-chain in the mouse strain retaining and transferring  $\lambda$ -HAC to its progeny is confirmed by the method as described in Example 65. The expression of complete human antibody molecules consisting of human Ig $\kappa$  chain, human Ig heavy-chain and human Ig $\kappa$  chain/heavy-chain in the mouse strain retaining and transferring  $\kappa$ -HAC to its progeny is confirmed by the method as described in Example 65. As described in Example 73, the mouse strain retaining and transferring  $\lambda$ -HAC or  $\kappa$ -HAC to its progeny is repeatedly mated with a mouse strain deficient in endogenous antibody heavy-chain and

light-chain  $\kappa$  genes, whereby the mouse strains retaining  $\lambda$ -HAC or  $\kappa$ -HAC and being homozygous for endogenous antibody heavy-chain gene and  $\kappa$  chain gene deficiency can be obtained. Complete human antibodies are mainly produced in these mouse strains.

[0424] Stable retention of each HAC is examined for the mouse strains retaining and transferring  $\lambda$ -HAC or  $\kappa$ -HAC to its progeny by the method described in Example 68-(5). The results show the stable retention of each HAC in the somatic cells of the mouse strains.

Example 104 Introduction of the human chromosome #22 fragment in which telomere-directed truncation occurred at LIF locus into a Chinese hamster CHO cell

[0425]

[0426] The human chromosome #22 fragment in which telomere-directed truncation occurred at LIF locus was introduced into CHO cells by MMCT for introduction of the fragment into mouse ES cells.

[0427] The chicken DT-40 cell clones retaining the human chromosome #22 fragment in which telomere-directed truncation had occurred at LIF locus were separately cultured on 8 Petri dishes with a diameter of 150 mm. When the cells reached a confluent state, the medium was exchanged by RPMI1640 media supplemented with 20% FBS, 1% chicken serum,  $10^{-4}$ M 2-mercaptoethanol, and 0.05  $\mu$ g/ml colcemid, and cultured for another 36 hours to form the microcell. The cells were suspended in 24 ml of a serum-containing RPMI1640 medium, and dispensed 2 ml into each of twelve 25 cm<sup>2</sup> centrifugation flasks (CORNING) precoated with 100  $\mu$ g/ml poly L-lysine. Then the cells were cultured for 1 hour at 37°C for the cells to adhere to the bottom of the flask. Following removal of the culture solution, a centrifugation flask was filled with cytochalasin B (10  $\mu$ g/ml, Sigma) solution kept at 37°C, and then subjected to centrifugation at 8000 rpm for 1 hour at 34°C. The microcells were suspended in serum-free DMEM media, and purified with a 8  $\mu$ m and 5  $\mu$ m filters. Following purification, the product was centrifuged at 1700 rpm, for 10 minutes, and then suspended in 5 ml of a serum-free DMEM medium. On the other hand, approximately  $10^7$  CHO cells were removed by trypsinization. The cells were washed twice in a serum-free DMEM medium, and then suspended in 5 ml of a serum-free DMEM medium. Again, the microcell was centrifuged at 1700 rpm for 10 minutes. Without removing the supernatant, 5 ml of the previously prepared CHO suspension was layered gently. Following centrifugation, the culture solution was removed, 0.5 ml of 1:1.4 PEG solution [5 g of PEG1000 (Wako Pure Chemicals Co., Ltd) and 1 ml of DMSO (Sigma) were dissolved in 6 ml of DMEM], followed by vigorous stirring with a pipette for approximately 2 minutes. Subsequently, 10 ml of a serum-free DMEM medium was added gently to the mixture for about 3 minutes, and allowed to stand for 10 minutes at 37 °C. After centrifugation, the cells were suspended in a F12 medium (GIBCO) supplemented with 10% FBS, dispensed into ten 24-well culture plates, and cultured for 24 hours at 37 °C. Then the medium was exchanged with a F12 medium containing 800  $\mu$ g/ml G418, followed by selection culture for 3 to 4 weeks.

[0428] Genome DNAs were extracted from the G418-resistant clones and subjected to PCR using Ig $\lambda$ , D22S315, D22S272, D22S278 detection primers and Puro-1 and LIF-1 primers (Kuroiwa et al., Nucleic Acid Research, 26: 3447-3448, 1998). Thus, two markers, D22S272 and D22S278 were not detected as expected, however, the other markers were all detected. In addition, FISH using human COT1 DNA and pGKPuro probes revealed that two human chromosome #22 fragments were retained and a signal derived from pGKPuro probe was observed on each of the one end of the telomere. Accordingly, it was concluded that the CHO cell clone retaining two individual human chromosome #22 fragments in which telomere-directed truncation had occurred at LIF locus was obtained.

Example 105

Introduction of human chromosome #22 fragment C68 into an endogenous antibody heavy-chain and  $\kappa$  chain deficient mouse ES cell retaining human chromosome #14 fragment SC20

[0429] The human chromosome #22 fragment C68 obtained in Example 83 was introduced by the microcell method into the mouse ES cell line HKD2-1 as produced by the method in Example 61-(1) to be deficient in endogenous antibody heavy-chain and  $\kappa$  chain and to retain the human chromosome #14 fragment SC20. Except the use of the CHO cell clone C68-6 retaining two human chromosome #22 fragments obtained in Example 104 as chromosome donor cells, the method described in Example 35 was employed. The resultant puromycin- and G418-resistant strain (double resistant strain) NLH (CHOI) was subjected to PCR analysis, confirming the retention of the introduced chromosomal fragments. Three types of primers, D14S543 (Example 68) for the chromosome #14 fragment, and Ig $\lambda$  and D22S315 for chromosome #22 (Example 2) were used in total. Thus, all three types of the markers were confirmed to be present in the NLH(CHO1) strain. In addition, FISH analysis was performed using a human chromosome-specific probe (Example 68). Microscopic study of 50 nuclear plates showed that two individual chromosomal fragments hybridizing to the probes were observed for 45 (90%) nuclear plates. These two chromosomal fragments were thought to be human chromosome #14 fragment SC20 and human chromosome #22 fragment C68, respectively. Therefore,

NLH(CH01) strain was confirmed to retain the chromosome #14 fragment SC20 and the chromosome #22 fragment C68 simultaneously.

Example 106 Detection and quantification of human antibodies in sera of the chimeric mice produced by injecting endogenous antibody heavy-chain and  $\kappa$  chain gene deficient mouse ES cells retaining the human chromosome #14 fragment SC20 and the human chromosome #22 fragment C68 into immunodeficient mouse host embryos

[0430] A chimeric mouse was produced by the method shown in Example 10 and the like from the mouse ES cell line NLH(CH01) obtained in Example 105. The host embryo used herein was an embryo at the 8-cell stage obtained by mating male and female antibody heavy-chain knockout mice established in Example 67-(1). A total of 307 injected embryos were transplanted so that 32 progeny mice were born. Whether progeny are chimeric or not was determined by their coat color. That is, if progeny exhibits wild type color (dark brown) among white coat color derived from the host embryo (ICR), it is determined as a chimeric mouse. Fourteen out of 32 progeny mice born clearly exhibited wild type coat color, for which the contribution of the ES cells was confirmed. These results confirms that the endogenous antibody heavy-chain gene- and  $\kappa$  chain gene-deficient mouse ES cell line NLH(CH01), retaining human chromosome #14 fragment SC20 and human chromosome #22 fragment C68 simultaneously, retains chimera-forming ability, that is, retains the ability to differentiate into normal mouse tissue.

[0431] Blood was collected from 6 to 10 week-old chimeric mice derived from NLH (CH01), and subjected to ELISA according to Examples 14, 32 and 85, for quantification of various human immunoglobulin concentrations and mouse immunoglobulin  $\lambda$  chain concentration in sera. Table 29 shows the result. High concentrations of human  $\mu$  chain,  $\gamma$  chain, and  $\lambda$  chain were detected in sera of the chimeric mice. The mouse  $\lambda$  chain carrying undisrupted genes was also detected, however, its concentration was lower than that of human  $\lambda$  chain.

[0432] These results suggested that the expression of complete human antibody molecules consisting of human heavy-chain and human  $\lambda$  chain in the chimeric mouse was dominant over that of hybrid antibody molecules consisting of human heavy-chain and mouse  $\lambda$  chain. That is, it was confirmed that the human chromosome introduced via the chicken DT-40 cell functions in the mouse and protein encoded by the human gene on the human chromosome is expressed.

[0433] Transfer of fragment C68 from the chimeric mice to its progeny mice was examined by the method described in Example 68-(4). Thus a mouse strain retaining fragment C68 and transferring it to progeny was established. The retention of fragment C68 in the mouse strain is shown by PCR and FISH analysis (Example 105). Furthermore, a mouse strain (C68+SC20) retaining both fragments SC20 and C68 can be obtained by mating this mouse with the mouse strain retaining and transferring chromosome #14 fragment SC20 to its progeny. The expression of complete human antibody molecules consisting of human Ig $\lambda$  chain, human Ig heavy-chain and human Ig $\lambda$  chain/heavy-chain in the mouse strain (C68+SC20) is confirmed by the method described in Example 65.

Table 29

Mouse name	Human $\mu$ chain (mg/l)	Human $\gamma$ chain (mg/l)	Human $\lambda$ chain (mg/l)	Mouse $\lambda$ chain (mg/l)
C-1	840	90	1300	150
C-3	560	170	470	170
C-12	980	650	1700	440

Example 107 Preparation of a HSA-specific human antibody-producing hybridoma from the chimeric mice produced by injecting endogenous antibody heavy-chain and  $\kappa$  chain gene- deficient mouse ES cells, which retain human chromosome #14 fragment SC20 and human chromosome #22 fragment C68, into immunodeficient mouse host embryos.

[0434] The chimeric mice C-10 (derived from NLH(CH01); 30% chimerism) as produced in Example 106 were immunized with HSA. A HSA solution 0.25mg/ml was prepared by mixing human serum albumin (HSA, Sigma, A3782) dissolved in PBS and an adjuvant (TiterMaxGold, Cytrx). The mouse was immunized twice with 0.15ml of the HSA solution (0.05 ml each was injected subcutaneously at three positions in total). Initial immunization was performed on the 7 week-old mice, and 21 days later, a second immunization was performed. According to Example 61 or 66, anti-HSA antibody concentration in sera diluted 1:100 was measured. Figure 75 shows the results. Since an increase in anti-HSA human antibody concentration was confirmed, final immunization was performed on day 30 with HSA dissolved in PBS. One clone producing HSA-specific human  $\mu$  chain and human  $\lambda$  chain was obtained by the production and cloning of hybridomas according to Example 66.

[0435] These results show that human Ig $\lambda$  chain expressed by the human chromosome introduced via the chicken

DT-40 cell binds specifically to HSA and is functional.

Example 108 Retention of the introduced human chromosomal fragments by the somatic cells of the chimeric mice produced by injecting endogenous antibody heavy-chain and  $\kappa$  chain gene-deficient mouse ES cells, which retain human chromosome #14 fragment and human chromosome #22 fragment, into immunodeficient mouse host embryos.

[0436] Fibroblasts derived from the tails of the chimeric mice C-12 (derived from NLH(CH01); chimerism 99%) produced in Example 106 were cultured following the method (Tomizuka et al., Nature Genetics, 16, 133-143). Preparation of chromosome samples from the fibroblasts and FISH analysis were performed according to the method (Tomizuka, et al., supra). Microscopic examination for 50 nuclear plates revealed that 21 (42%) nuclear plates contained two independent chromosomes hybridizing to human COT1 probes. These two chromosomes are thought to be chromosome #14 fragment SC20 and chromosome #22 fragment C68, respectively. Hence, it was shown that the somatic cell of the chimeric mouse retains two types of chromosomal fragments.

Example 109 Preparation of a complete human antibody-producing hybridoma against sugar chain antigen (GM2) from the chimeric mice as produced by injecting endogenous antibody heavy-chain and light-chain-deficient mouse ES cells, which retains human chromosome #14 partial fragment and human chromosome #22 partial fragment, into immunodeficient mouse host embryos.

[0437] The hybridomas as obtained in Example 84-(3) were subjected to selection culture in the presence of G418 using 96 well plates. Then the antibodies in the culture solution were subjected to screening by ELISA according to Example 84-(3). Cloning was performed by the limited dilution method, thereby obtaining 11 clones of the antibody-producing hybridoma. The antibodies in the culture supernatant were analyzed by ELISA so that binding of the obtained monoclonal antibodies to GM2 was confirmed.

[0438] These results suggest that the complete human antibody-producing hybridoma against sugar chain antigen ganglioside GM2 can be obtained from the chimeric mouse. The obtained human antibody is expected to apply for treating diseases such as HIV and melanoma.

Example 110 Preparation of a complete human antibody-producing hybridoma against sugar chain antigen (asialo-GM1) from the endogenous antibody heavy-chain and light-chain-deficient mice retaining human chromosome #14 partial fragment and human chromosome #2 partial fragment.

[0439] The Tc mice HK232 as produced in Example 85 were immunized with asialo-GM1. An adjuvant (MPL+TDM Emulsion, RIBI Immunochem Research Inc., R-700) was dissolved in 2 ml of a mixture of chloroform and methanol (2:1). This solution was mixed with 1 mg of asialo-GM1 (ISOSEP AB, 65/12) dissolved in 2 ml of a mixture of chloroform and methanol (2:1). The mixture was transferred into a round-bottom flask, and dried with a rotary evaporator. Two ml of PBS was added to the dried product and mixed vigorously using a Vortex mixer, thereby preparing suspension Gg4-RIBI. The mouse was immunized intraperitoneally with 0.2 ml of Gg4-RIBI and 500  $\mu$ g of anti-mouse CD40 antibodies. One week after the immunization, blood was collected, and an increase in the antibody titer was confirmed by ELISA according to Example 84. Two weeks after the immunization, final immunization was performed intraperitoneally with the suspension Gg4-RIBI while 5  $\mu$ g of recombinant human IL-6 was subcutaneously injected. Three days after final immunization, the mouse spleen was excised, followed by cell fusion according to Example 24. Thus the hybridomas were produced. Selection culture was performed in the presence of G418, or G418 and puromycin using 96-well plates. Then the antibodies in the culture solution were subjected to screening by ELISA according to Example 84. Moreover, 0.15 mg of asialo-GM1, 1.5 mg of Galactocerebroside, and 1.5 mg of cholesterol were dissolved in 2 ml of a mixture of chloroform and methanol (2:1) and dried. Next the dried product was resuspended in 100 ml of PBS, dispensed 100  $\mu$ l per well into ELISA plates for solid phase formation. Screening was performed by ELISA using the prepared plates. The cells in the wells shown as positive by the two types of screening above were cloned. The culture supernatant of the anti-asialo-GM1 human IgM antibody-producing hybridomas were allowed to react with HIV-infected cells and analyzed by a flowcytometry. Therefore, one clone of the hybridoma producing monoclonal antibodies that bind to HIV-infected cells rather than HIV-uninfected cells was obtained.

[0440] These results suggest that the complete human antibody-producing hybridoma against sugar chain antigen Asialo-GM1 is obtained from the double Tc mouse. Thus obtained human antibody is expected to apply for treatment for diseases, such as HIV.

Example 111 Preparation of a complete human antibody-producing hybridoma against human TNF- $\alpha$  from the endogenous antibody heavy-chain and light-chain-deficient mouse retaining human chromosome #14 partial fragment and human chromosome #2 partial fragment.

- 5 [0441] Hybridomas were produced by immunizing the 8 week-old Tc mice HK492 as produced in Example 85 with human TNF- $\alpha$  according to the methods of Examples 84 and 85. TNF- $\alpha$  solution dissolved in PBS was mixed with the same amount of adjuvant (Titer Max Gold, CytRx). The mice were immunized with this mixture 25  $\mu$ g/mouse. Immunization was performed over approximately 80 days at an interval of 2 to 3 weeks. After an increase in the antibody concentration was confirmed, final immunization was performed intraperitoneally with only TNF- $\alpha$  solution. Then hybridomas were produced according to Example 86. The cells in the wells positive for the antibody were cloned, so that one clone of the anti-TNF- $\alpha$  human IgG/k antibody-producing hybridoma was obtained.

[0442] All of the publications, patents, and patent applications cited in this specification are all incorporated as reference into this specification.

#### 15 Industrial Applicability

[0443] The present invention provides a chimeric non-human animal retaining single or multiple foreign chromosome(s) or fragment(s) thereof, and expressing the gene on the chromosome(s) or on the fragment(s) thereof. Biologically active substances can be produced using the chimeric non-human animal of this invention.

- 20 [0444] The present invention provides a pluripotent cell retaining single or multiple foreign chromosome(s) or fragment(s) thereof, and expressing the gene on the chromosome(s) or on the fragments thereof. Using this cell, hereditary diseases can be treated by bone marrow transplantation and the like.

[0445] The present invention provides a pluripotent cell carrying at least two types of disrupted endogenous genes. By using the cell of this invention as a recipient cell, into which single or multiple foreign chromosomes or fragments thereof containing the same gene as the disrupted endogenous gene or a gene homologous to the same are introduced, a functional cell or a chimeric non-human animal, which retains single or multiple foreign chromosome(s) or fragment(s) thereof, and expresses the gene on the foreign chromosome(s) or on fragment(s) thereof, can be produced. Expression of the gene on single or multiple foreign chromosome(s) or on fragment(s) thereof in such a chimeric non-human animal or its progeny, its tissues or its cells thereof allows production of a biologically active substance.

- 30 [0446] Furthermore, the present invention provides a method for artificially modifying a chromosome(s) or fragment(s) thereof.

#### Free Text of Sequence List

- 35 [0447] SEQ ID NOS:1-58 and 61-74 show nucleotide sequences of primers.  
 [0448] SEQ ID NOS:59 and 60 show nucleotide sequences of probes.  
 [0449] SEQ ID NO:75 shows a nucleotide sequence of Sfi I linker.  
 [0450] SEQ ID NO:76-79 show nucleotide sequences of primers.  
 [0451] SEQ ID NO:80 shows a nucleotide sequence of Srf I linker.  
 40 [0452] SEQ ID NOS:81-108 show nucleotide sequences of primers.

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55



SEQUENCE LISTING

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<120> Method for Modifying Chromosomes

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&lt;223&gt; Description of Artificial Sequence: Primer

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22

#### Claims

1. A method for producing a cell comprising a modified foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

(a) preparing a microcell comprising a foreign chromosome(s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell with high homologous recombination efficiency through its fusion with said microcell;

(b) in said cell with high homologous recombination efficiency, inserting a targeting vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking the desired site; and



(c) in said cell with high homologous recombination efficiency, causing deletion and/or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof.

2. The method of claim 1, wherein a plurality of said cells with high homologous recombination efficiency are subjected to whole cell fusion after steps (a) and (b) and are subjected to step (c).
3. The method of claim 2, wherein a plurality of said cells with high homologous recombination efficiency each comprises a distinct foreign chromosome(s) or a fragment(s) thereof.
4. The method of claim 1, wherein said cell comprising a modified foreign chromosome(s) or a fragment(s) thereof is an animal cell.
5. The method of claim 4, wherein said animal cell is a mammalian cell.
6. The method of claim 4, wherein said animal cell is a non-human animal cell.
7. The method of claim 1, wherein said targeting vector contains a telomere sequence which is introduced into a desired site by insertion of the targeting vector.
8. The method of claim 2, wherein said deletion occurs at a site into which said telomere sequence has been introduced.
9. The method of claim 1, wherein said targeting vector comprises a recognition sequence for a site-directed recombination enzyme, and said recognition sequence is introduced into a desired site by insertion of the targeting vector.
10. The method of claim 9, wherein a vector, which is capable of expressing a site-directed recombination enzyme, is introduced into said cell with high homologous recombination efficiency simultaneously with or after insertion of said targeting vector comprising the recognition sequence for a site-directed recombination enzyme, so that an activity of said site-directed recombination enzyme is expressed, resulting in deletion and/or translocation of said foreign chromosome(s) or a fragment(s) thereof at a site in which said recognition sequence for a site-directed recombination enzyme is introduced.
11. The method of claim 10, wherein said translocation occurs between a plurality of foreign chromosomes or fragments thereof.
12. The method of claim 11, wherein a plurality of said foreign chromosomes are derived from the same species.
13. The method of claim 12, wherein said same species is a human.
14. The method of claim 11, wherein a plurality of said foreign chromosomes are derived from different species.
15. The method of claim 14, wherein said species are a human and a mouse.
16. The method of claim 10, wherein said translocation occurs between a foreign chromosome(s) or a fragment(s) thereof and a chromosome(s) derived from said cell with high homologous recombination efficiency.
17. The method of claim 9, wherein said site-directed recombination enzyme is a Cre enzyme.
18. The method of claim 9, wherein said recognition sequence for a site-directed recombination enzyme is a LoxP sequence.
19. The method of claim 1, wherein said cell with high homologous recombination efficiency is an embryonic stem cell (or ES cell).
20. The method of claim 1, wherein said cell with high homologous recombination efficiency is a chicken DT-40 cell.
21. The method of claim 1, which further comprises a step of screening a cell comprising said foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred.

22. The method of claim 21, wherein said screening is based on expression of a marker gene.

23. The method of claim 22, wherein said marker gene is a drug-resistance gene.

5 24. The method of claim 22, said marker gene is a green fluorescent protein-encoding gene derived from the jellyfish *Aequorea victoria* or a modified gene thereof.

25. The method of claim 1, wherein said foreign chromosome(s) or a fragment(s) thereof is derived from a human.

10 26. A method for producing a chimeric non-human animal comprising a modified foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

(a) preparing a microcell comprising a foreign chromosome(s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell with high homologous recombination efficiency through its fusion with said microcell;

15 (b) in said cell with high homologous recombination efficiency, inserting a vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof, and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking said desired site;

20 (c) in said cell with high homologous recombination efficiency, causing deletion and/or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof; and

(d) preparing a microcell comprising said foreign chromosome(s) or a fragment(s) thereof in which deletion or translocation has occurred, and transferring said foreign chromosome(s) or a fragment(s) thereof into a pluripotent non-human animal cell through its fusion with said microcell.

25 27. The method of claim 26, wherein a plurality of said cells with high homologous recombination efficiency are subjected to whole cell fusion after steps (a) and (b) and are subjected to step (c).

30 28. The method of claim 27, wherein a plurality of said cells with high homologous recombination efficiency each comprise a distinct foreign chromosome(s) or a fragment(s) thereof.

29. The method of claim 26, wherein said targeting vector comprises a telomere sequence which is introduced into a desired site by insertion of the targeting vector.

35 30. The method of claim 29, wherein said deletion occurs at a site where said telomere sequence has been introduced.

31. The method of claim 26, wherein said targeting vector comprises a recognition sequence for a site-directed recombination enzyme, and said recognition sequence is introduced into a desired site by insertion of the targeting vector.

40 32. The method of claim 31, wherein a vector, which is capable of expressing a site-directed recombination enzyme, is introduced into said cell with high homologous recombination efficiency simultaneously with or after insertion of said targeting vector comprising said recognition sequence for a site-directed recombination enzyme, so that an activity of said site-directed recombination enzyme is expressed, resulting in deletion and/or translocation of said foreign chromosome(s) or a fragment(s) thereof at a site into which said recognition sequence is introduced.

33. The method of claim 32, wherein said translocation occurs between a plurality of foreign chromosomes or fragments thereof.

50 34. The method of claim 32, wherein said translocation occurs between said foreign chromosome(s) or a fragment(s) thereof and said chromosome(s) derived from said cell with high homologous recombination efficiency.

35. The method of claim 31, wherein said site-directed recombination enzyme is a Cre enzyme.

55 36. The method of claim 31, wherein said recognition sequence for site-directed recombination enzyme is a LoxP sequence.

37. The method of claim 26, wherein said cell with high homologous recombination efficiency is an embryonic stem

cell (or ES cell).

38. The method of claim 26, wherein said cell with high homologous recombination efficiency is a chicken DT-40 cell.

39. The method of claim 26, which further comprises a step of screening cells comprising a foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred.

40. The method of claim 39, wherein said screening is based on expression of a marker gene.

41. The method of claim 40, wherein said marker gene is a drug-resistance gene.

42. The method of claim 40, the marker gene is a green fluorescent protein-encoding gene derived from the jellyfish *Aequorea victoria* or a modified gene thereof.

43. The method of claim 26, wherein in the step (d), a microcell is produced from said cell with high homologous recombination efficiency; said foreign chromosome(s) or a fragment(s) thereof, in which deletion and/or translocation has occurred is transferred into a CHO cell through its fusion with said microcell; a microcell is produced from the CHO cell; and then said foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred is transferred into a pluripotent cell through its fusion with said microcell.

44. The method of claim 26, said pluripotent cell is an embryonic stem cell (or ES cell).

45. The method of claim 26, said foreign chromosome(s) or a fragment(s) thereof is derived from a human.

46. A method for producing a non-human animal comprising a modified foreign chromosome(s) or a fragment(s) thereof, which comprises the steps of:

(a) preparing a microcell comprising a foreign chromosome(s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment (s) thereof into a cell with high homologous recombination efficiency through its fusion with said microcell;

(b) in said cell with high homologous recombination efficiency, inserting a vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof, and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking said desired site;

(c) in said cell with high homologous recombination efficiency, causing deletion and/or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof;

(d) preparing a microcell comprising said foreign chromosome(s) or a fragment(s) thereof, in which deletion and/or translocation has occurred, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell derived from a non-human animal through its fusion with said microcell; and

(e) transplanting the nucleus of said cell derived from the non-human animal into an enucleated unfertilized egg derived from a homologous non-human animal of the same species.

47. The method of claim 46, wherein a plurality of said cells with high homologous recombination efficiency are subjected to whole cell fusion after steps (a) and (b) and are subjected to the step (c).

48. The method of claim 47, wherein a plurality of said cells with high homologous recombination efficiency comprise a distinct foreign chromosome(s) or a fragment (s) thereof.

49. The method of claim 46, wherein said targeting vector comprises a telomere sequence, which is introduced into a desired site by insertion of the targeting vector.

50. The method of claim 49, wherein said deletion occurs at a site into which a telomere sequence has been introduced.

51. The method of claim 46, wherein said targeting vector comprises a recognition sequence for site-directed recombination enzyme, and said recognition sequence is introduced into a desired site by insertion of the targeting vector.

52. The method of claim 51, wherein a vector, which is capable of expressing a site-directed recombination enzyme, is introduced into said cell with high homologous recombination efficiency simultaneously with or after insertion of

said targeting vector comprising said recognition sequence for a site-directed recombination enzyme, so that an activity of said site-directed recombination enzyme is expressed, resulting in deletion and/or a translocation of said foreign chromosome(s) or fragment(s) thereof at a site into which said recognition sequence is introduced.

- 5 53. The method of claim 52, wherein said translocation occurs between a plurality of foreign chromosomes or fragments thereof.
54. The method of claim 52, wherein said translocation occurs between said foreign chromosome(s) or a fragment(s) thereof and said chromosome derived from a cell with high homologous recombination efficiency.
- 10 55. The method of claim 51, wherein said site-directed recombination enzyme is a Cre enzyme.
56. The method of claim 51, wherein said recognition sequence for a site-directed recombination enzyme is a LoxP sequence.
- 15 57. The method of claim 46, wherein said cell with high homologous recombination efficiency is an embryonic stem cell (or ES cell).
58. The method of claim 46, wherein said cell with high homologous recombination efficiency is a chicken DT-40 cell.
- 20 59. The method of claim 46, which further comprises a step of screening cells containing a foreign chromosome(s) or a fragment(s) thereof in which deletion and/or translocation has occurred.
60. The method of claim 59, wherein said screening is based on expression of a marker gene.
- 25 61. The method of claim 60, wherein said marker gene is a drug-resistant gene.
62. The method of claim 60, wherein said marker gene is a green fluorescent protein-encoding gene derived from the jellyfish *Aequorea victoria* or a modified gene thereof.
- 30 63. The method of claim 46, wherein, in the step (d), a microcell is produced from said cell with high homologous recombination efficiency; said foreign chromosome(s) or fragment(s) thereof, in which deletion and/or translocation have/has occurred, is/are transferred into a CHO cell through its fusion with the microcell; a microcell is produced from the CHO cell; and then said foreign chromosome(s) or a fragment(s) thereof, in which deletion and/or translocation has occurred, is transferred into a cell derived from a non-human animal through its fusion with the microcell.
- 35 64. The method of claim 46, said cell derived from a non-human animal is a culture cell derived from an embryo or a blastocyst.
- 40 65. The method of claim 46, said cell derived from a non-human animal is a culture cell derived from a fetus or an adult.
66. The method of claim 46, said cell derived from a non-human animal is a fibroblast cell derived from fetus.
- 45 67. The method of claim 46, said foreign chromosome(s) or a fragment(s) thereof is derived from a human.
68. A non-human animal, which retains a chromosomal fragment(s) obtained by deletion of a foreign chromosome(s) or a fragment(s) thereof.
- 50 69. The non-human animal of claim 68, wherein said chromosomal fragment(s) comprises:
  - (i) a marker gene and a telomere sequence, and/or
  - (ii) a recognition sequence for a site-directed recombination enzyme.
- 55 70. A non-human animal, comprising a recombinant foreign chromosome(s) obtained by translocation between a plurality of foreign chromosomes or fragments thereof.
71. The non-human animal of claim 70, wherein said recombinant chromosomal fragment(s) comprises:

- (i) a marker gene and a telomere sequence; and/or
- (ii) a recognition sequence for a site-directed recombination enzyme.

72. The non-human animal of claim 70, wherein said recombinant foreign chromosome(s) is independently maintained in the nucleus of the non-human animal cell.

73. The non-human animal of claim 70, wherein said recombinant foreign chromosome(s) is derived from a human.

74. The non-human animal of claim 70, wherein the recombinant foreign chromosome(s) is derived from human chromosomes #14 and #2.

75. The non-human animal of claim 70, wherein said recombinant foreign chromosome(s) is derived from human chromosomes #14 and #22

76. The non-human animal of claim 70, wherein said recombinant foreign chromosome(s) comprises genes for a heavy-chain and a light-chain  $\lambda$  of a human antibody.

77. The non-human animal of claim 70, wherein said recombinant foreign chromosome(s) comprises genes for a heavy-chain and a light-chain  $\kappa$  gene of a human antibody.

78. The non-human animal of claim 70, which is a mouse.

79. The non-human animal of claim 70, which is an ungulata.

80. The non-human animal of claim 70, which is a bovine.

81. The non-human animal of claim 70, which is an ovine.

82. The non-human animal of claim 70, which is an avian.

83. The non-human animal of claim 70, which is a chicken.

84. A cell, comprising a recombinant chromosome(s) or a fragment(s) thereof obtained by deletion and/or translocation of a chromosome(s) or a fragment(s) thereof, which comprises at least a part of a human chromosome and into which

- (i) a marker gene and telomere sequence, and/or
- (ii) a recognition sequence for site-directed recombination enzyme is/are introduced.

85. A method for modifying a foreign chromosome(s) or a fragment(s) thereof in a cell, which comprises the steps of:

- (a) preparing a microcell containing a foreign chromosome (s) or a fragment(s) thereof, and transferring said foreign chromosome(s) or a fragment(s) thereof into a cell with high homologous recombination efficiency through its fusion with the microcell;
- (b) in said cell with high homologous recombination efficiency, inserting a targeting vector by homologous recombination into a desired site of said foreign chromosome(s) or a fragment(s) thereof and/or a desired site of a chromosome(s) derived from said cell with high homologous recombination efficiency, thereby marking said desired site; and
- (c) in said cell with high homologous recombination efficiency, causing deletion or translocation to occur at the marked site of said foreign chromosome(s) or a fragment(s) thereof.

86. An artificial chromosome vector, which comprises a centromere sequence derived from a human chromosome #14 or #21, and a recognition sequence for a site-directed recombination enzyme.

87. The artificial chromosome vector of claim 86, wherein said recognition sequence for a site-directed recombination enzyme is a LoxP sequence.

88. A recombinant chromosome or a fragment thereof obtained by deletion and/or translocation of a chromosome(s)

or a fragment(s) thereof, into which

(i) a marker gene and a telomere sequence, and/or

(ii) a recognition sequence for a site-directed recombination enzyme is/are introduced, and which comprises  
at least a part of a human chromosome.

89. The recombinant chromosome or a fragment thereof of claim 88, which comprises fragments of human chromosomes #14 and #22.

90. The recombinant chromosome or a fragment thereof of claim 88, which comprises fragments of human chromosomes #14 and #2.

91. The recombinant chromosome or a fragment thereof of claim 88, which comprises genes for a heavy-chain and a light-chain  $\lambda$  of a human antibody.

92. The recombinant chromosome or a fragment thereof of claim 88, which comprises genes for a heavy-chain and a light-chain  $\kappa$  of a human antibody.

## MICROCELL HYBRID CLONES

	C8 and others from group c				N2 and others from group N									Z-15
	B-5	B-24	D-1	D-2	D-8	D-14	E-21	L-34	M-16	T-9	W-23	Y-68		
S207	X	X	X	X	●	●	●	●	X	X	X	X	●	
S177	X	X	X	X	●	●	●	●	X	●	X	X	●	
POLYMORPHIC	●	●	X	X	●	●	●	●	X	X	●	X	●	
AND GENETIC	●	●	●	●	●	●	●	●	●	●	●	●	●	
MARKERS	●	●	X	X	●	●	●	●	X	X	●	X	●	
S156	X	X	X	X	●	●	●	●	X	X	X	X	●	
S159	X	X	X	X	●	●	●	●	X	X	X	X	●	
CHROMOSOME	I	I	T	I	I	I	I	I	T	I	I	I	I	

I: INDEPENDENT      T: TRANSLOCATED

FIG. 1

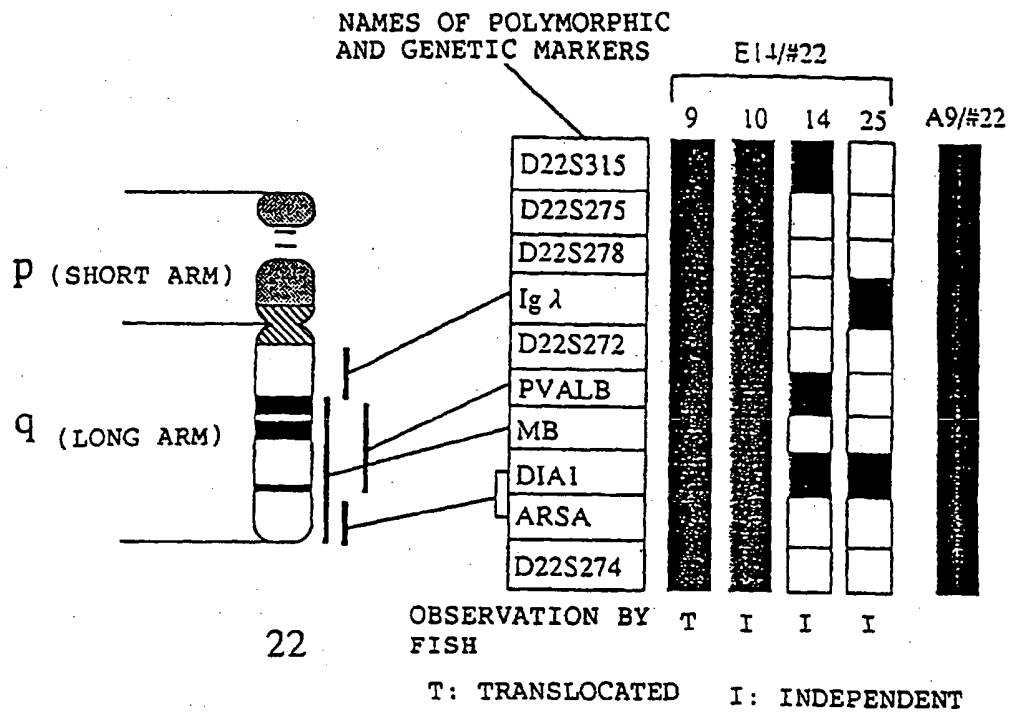


FIG. 2



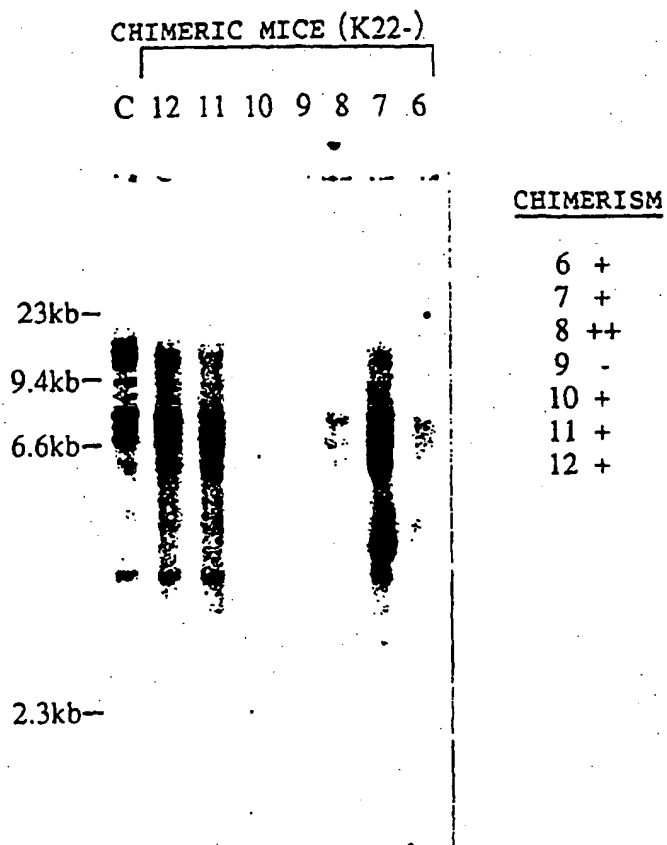


FIG. 3

B : BRAIN  
 L : LIVER  
 SM : SKELETAL MUSCLE  
 H : HEART  
 Sp : SPLEEN  
 Th : THYMUS  
 Ov : OVARY  
 K : KIDNEY  
 nc : NEGATIVE CONTROL  
 pc : POSITIVE CONTROL



DIA1 : HUMAN CYTOCHROME b5 REDUCTASE

FIG. 4

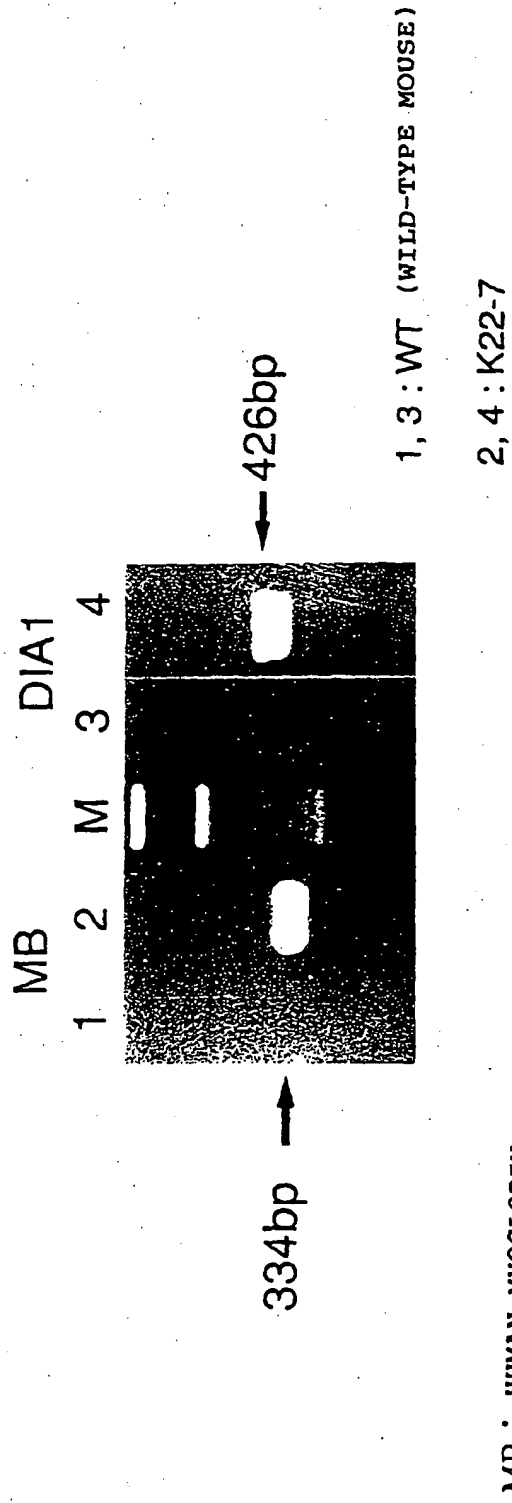


FIG. 5

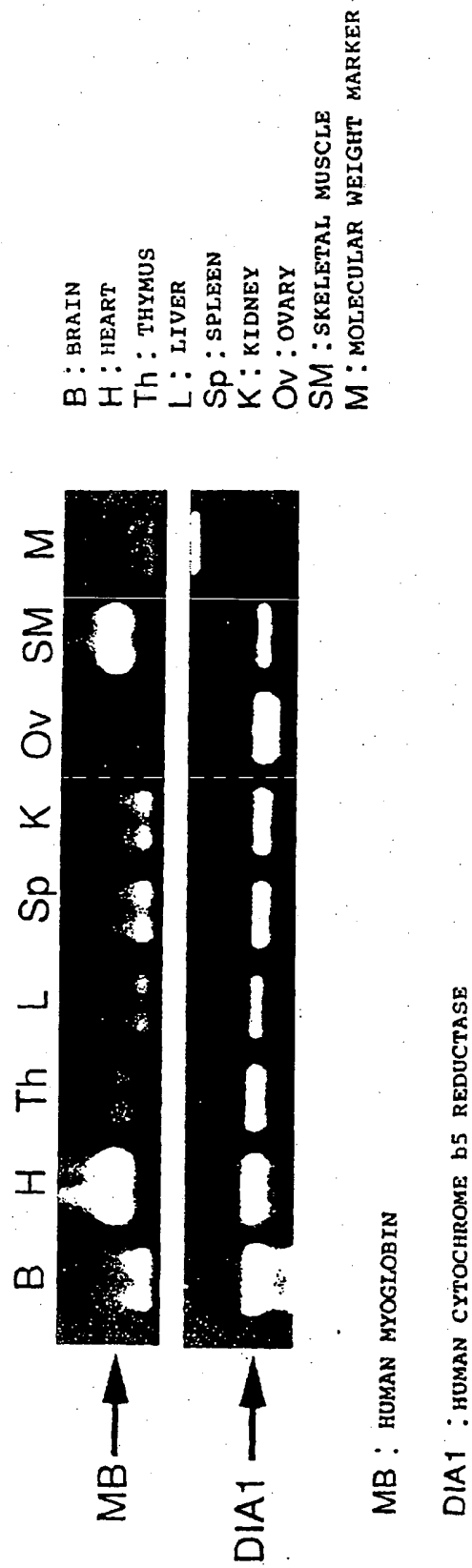
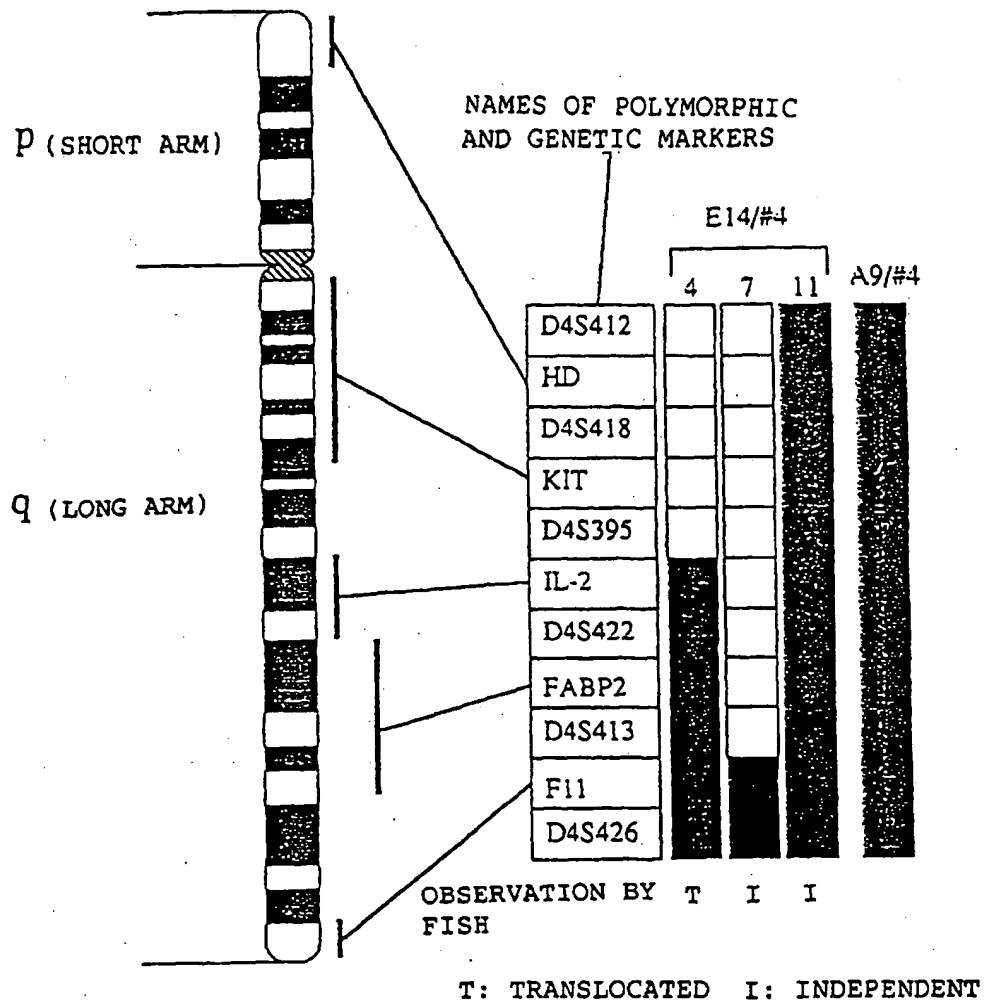


FIG. 6



4

FIG. 7

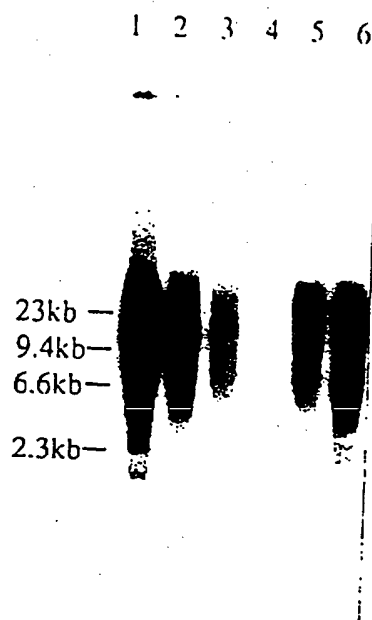


FIG. 8

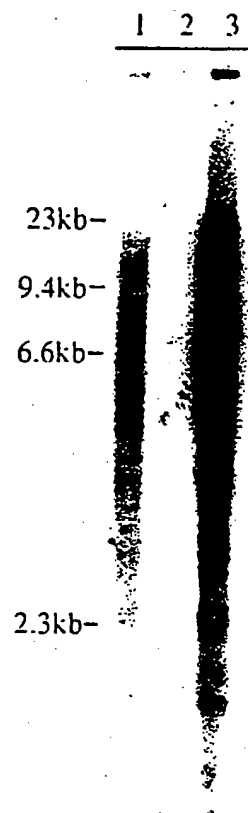


FIG. 9

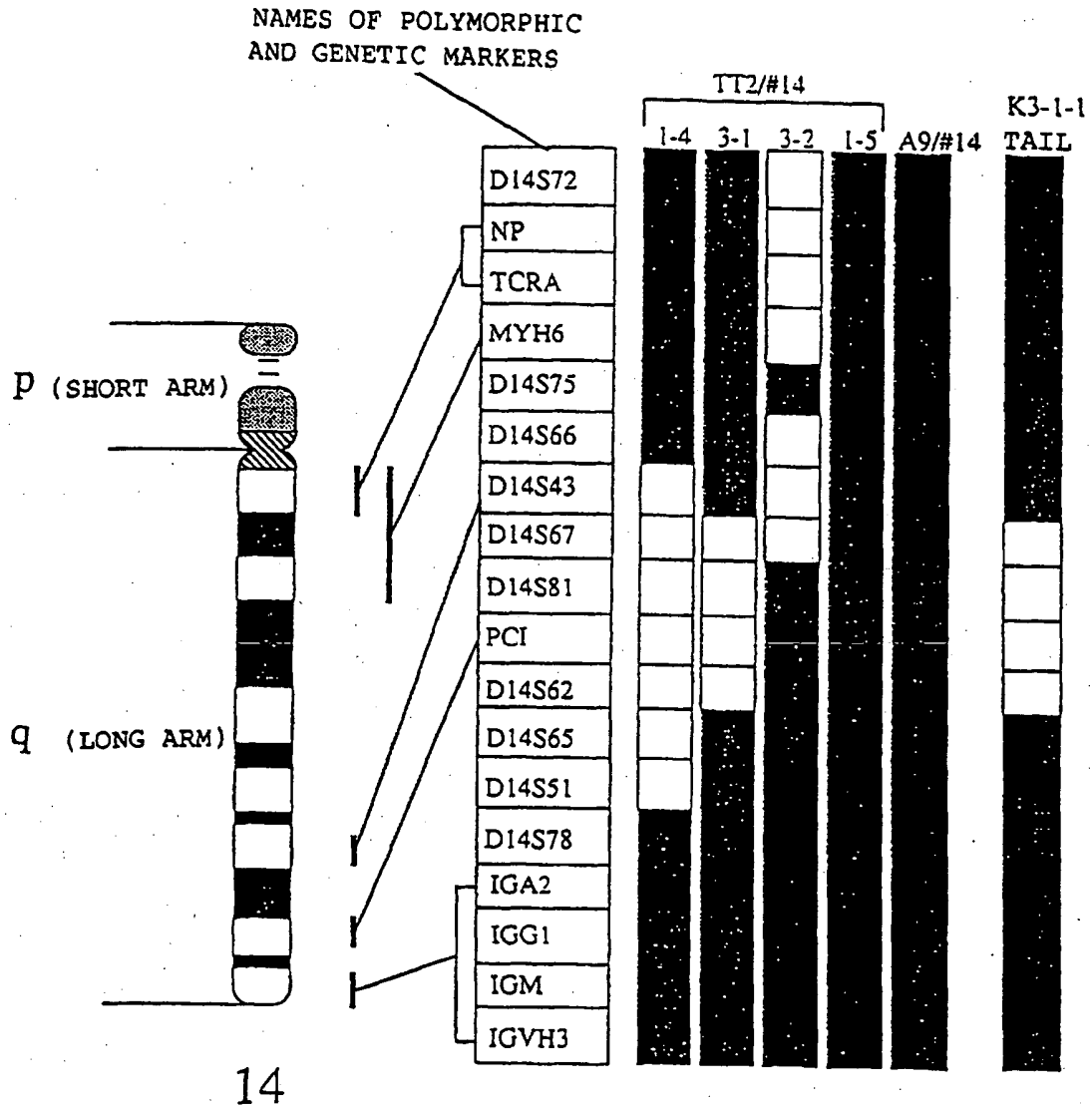


FIG. 10



B : BRAIN  
 K : KIDNEY  
 Sp : SPLEEN  
 H : HEART  
 L : LIVER  
 Th : THYMUS  
 pc : POSITIVE CONTROL  
 nc : NEGATIVE CONTROL  
 M : MOLECULAR WEIGHT MARKER



IgM : HUMAN IMMUNOGLOBULIN  $\mu$  CHAIN

FIG. 11

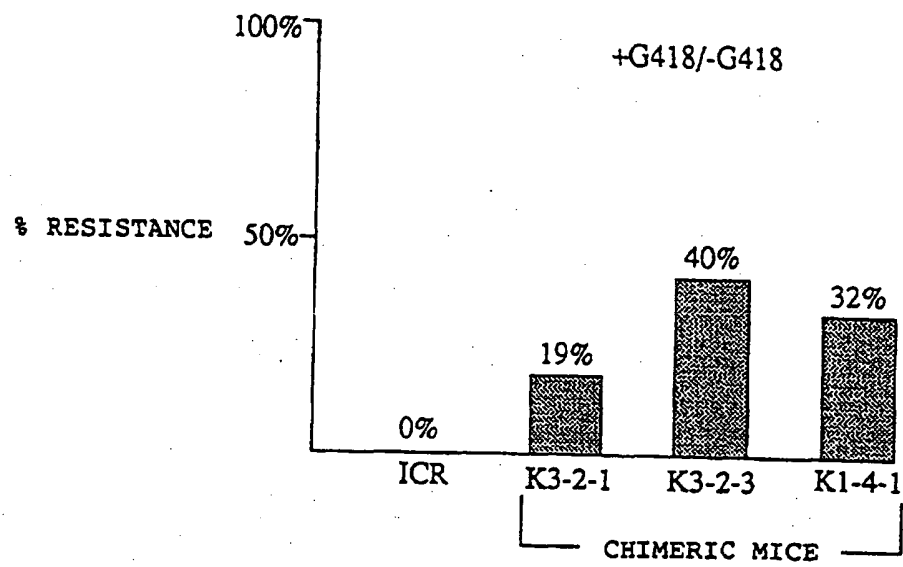


FIG. 12

FIG. 13

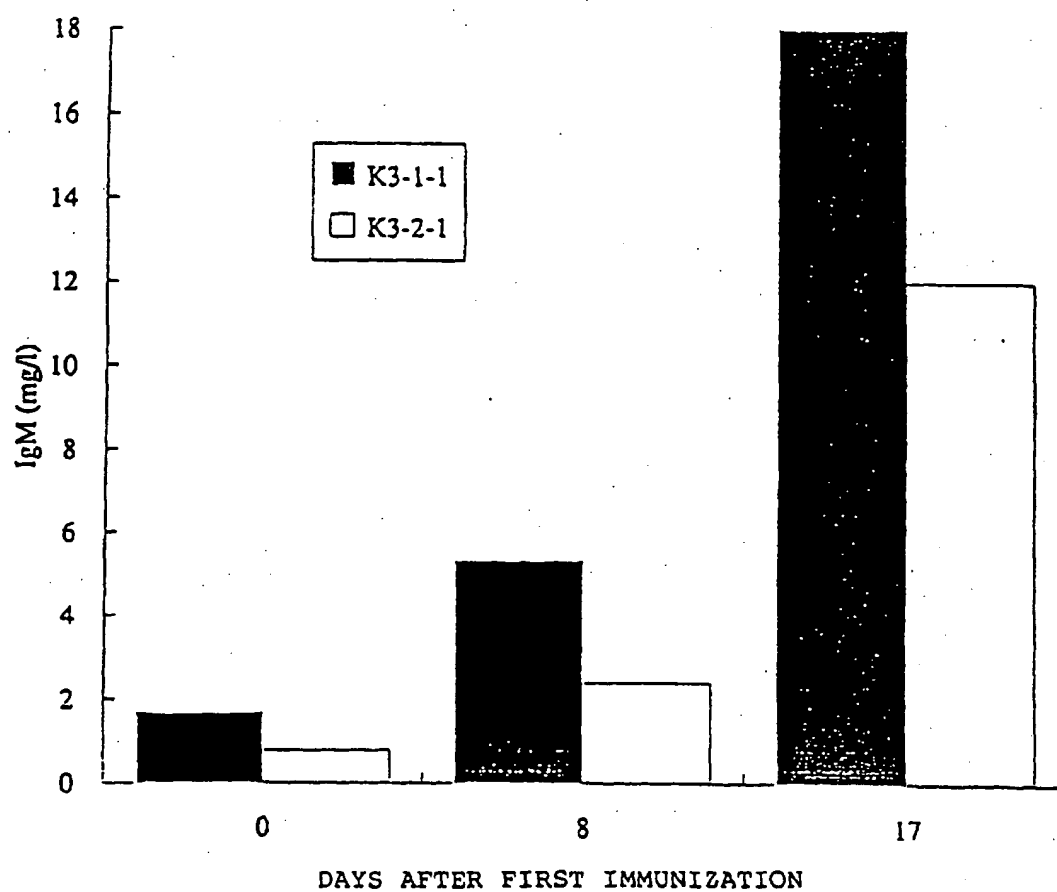


FIG. 14

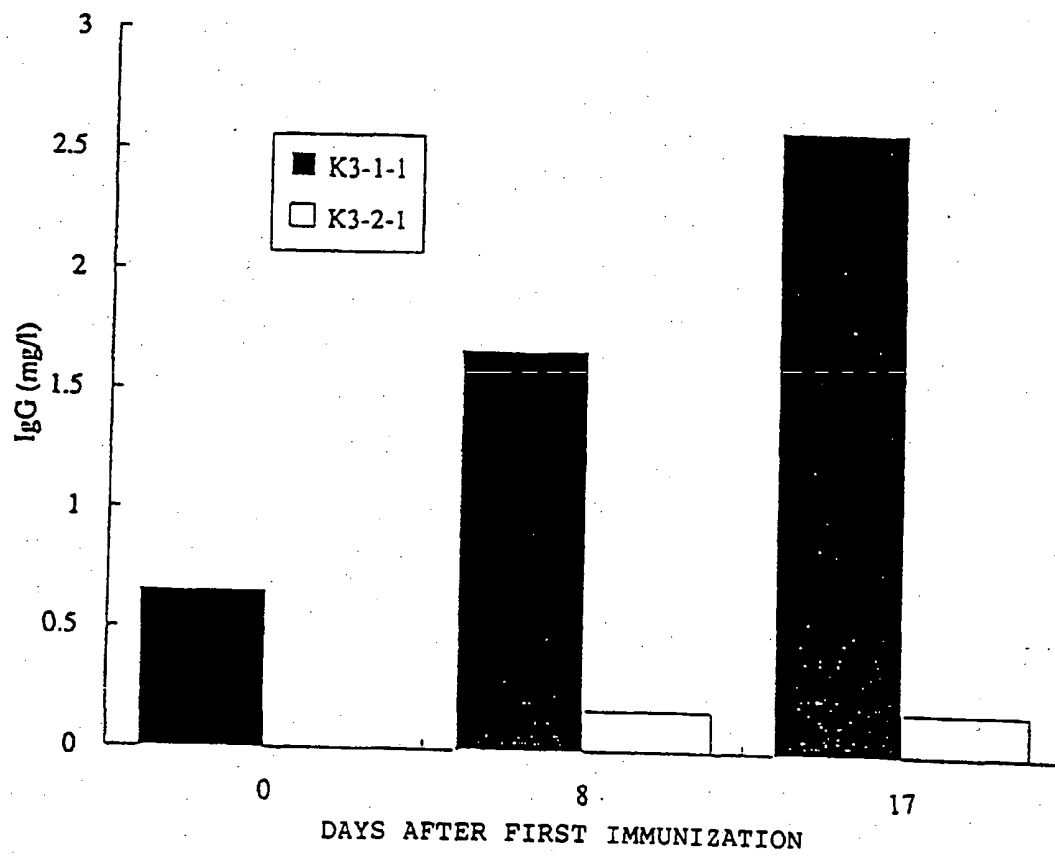
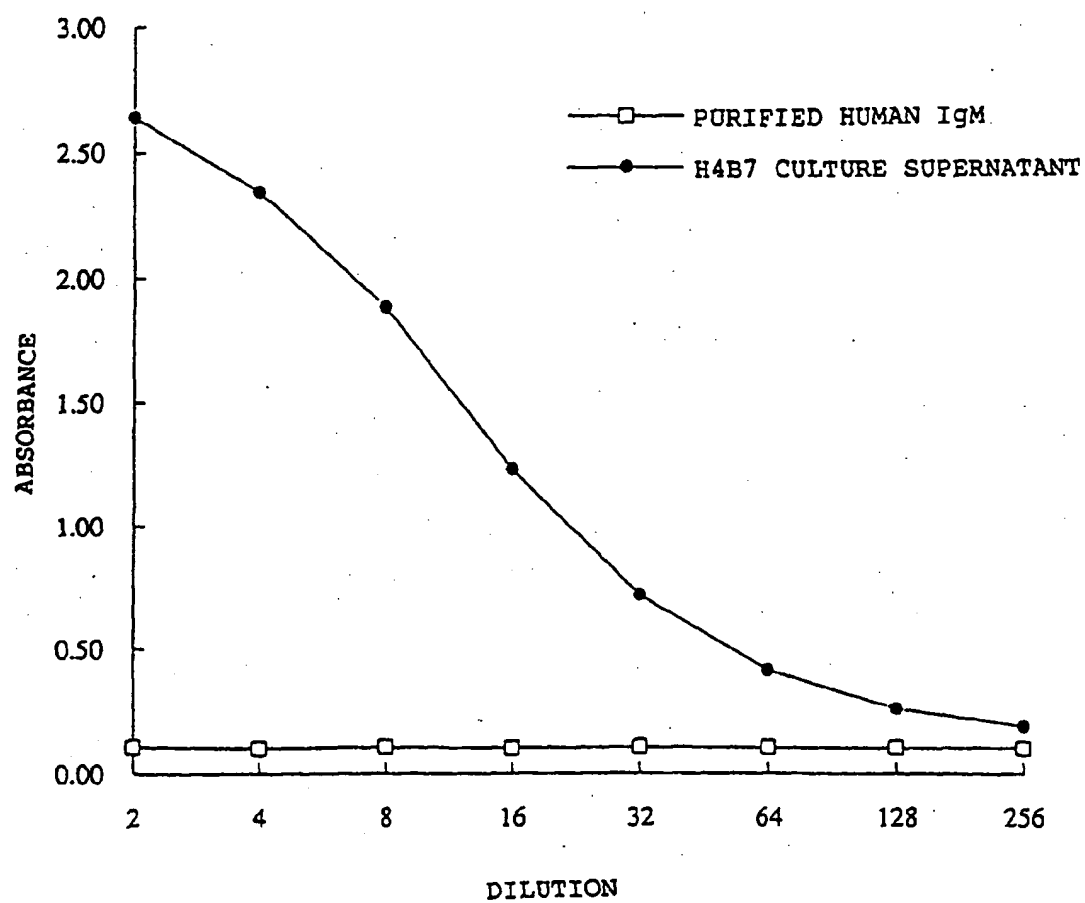


FIG. 15



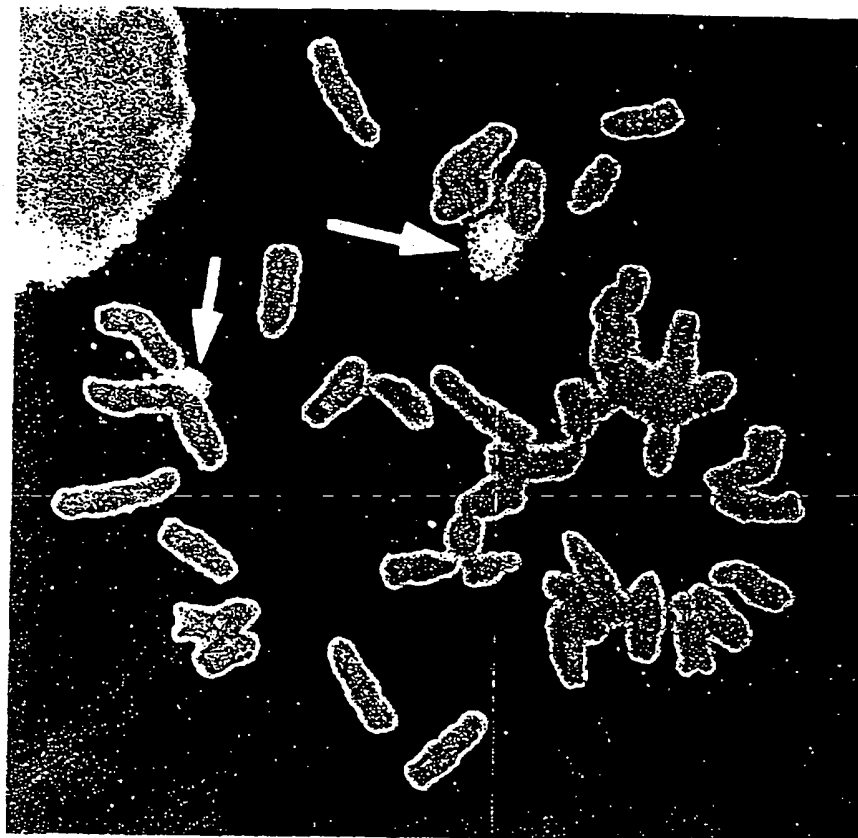


FIG. 16

FIG. 17

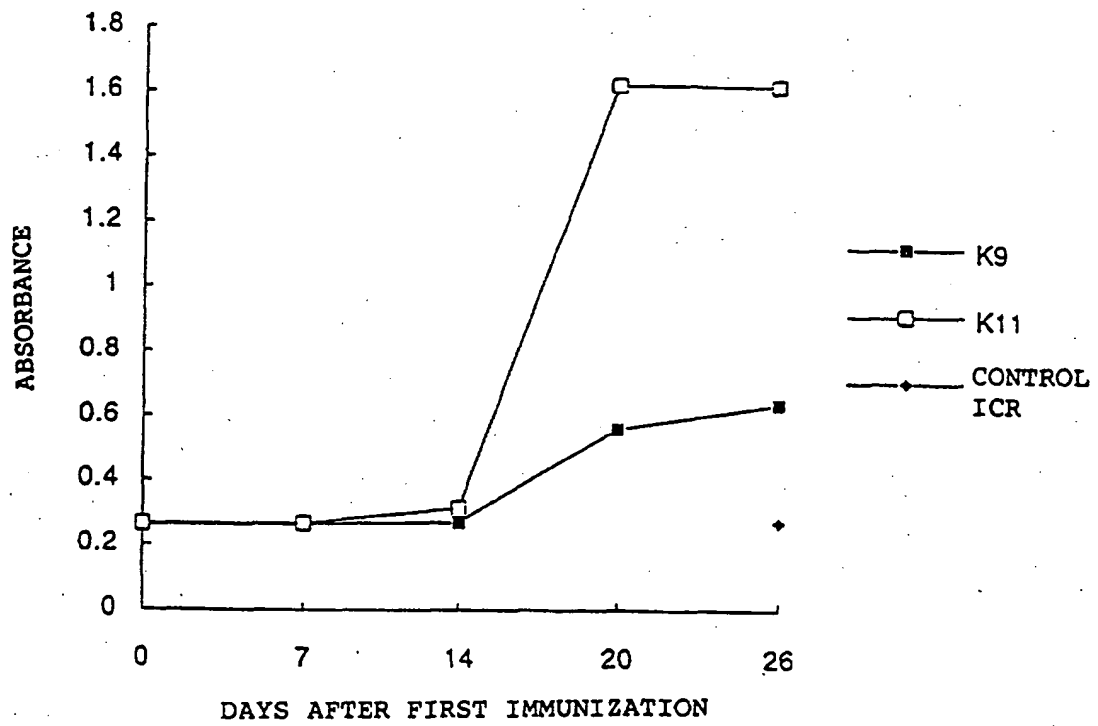


FIG. 18

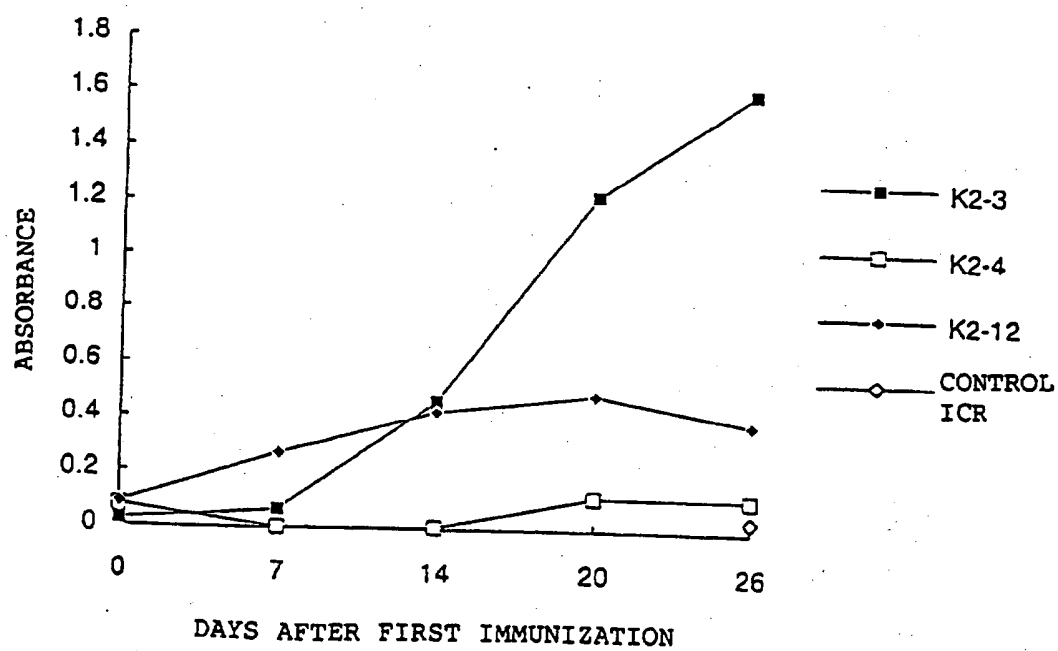




FIG. 19

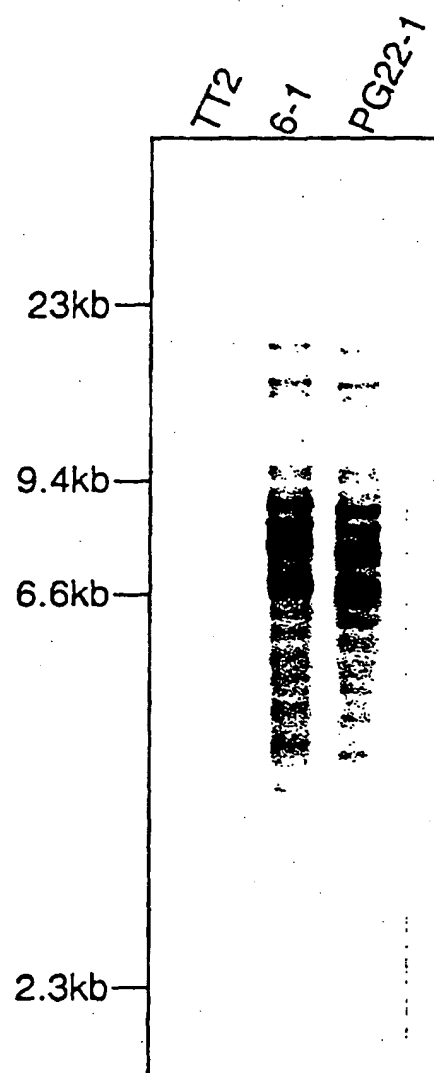


FIG. 20

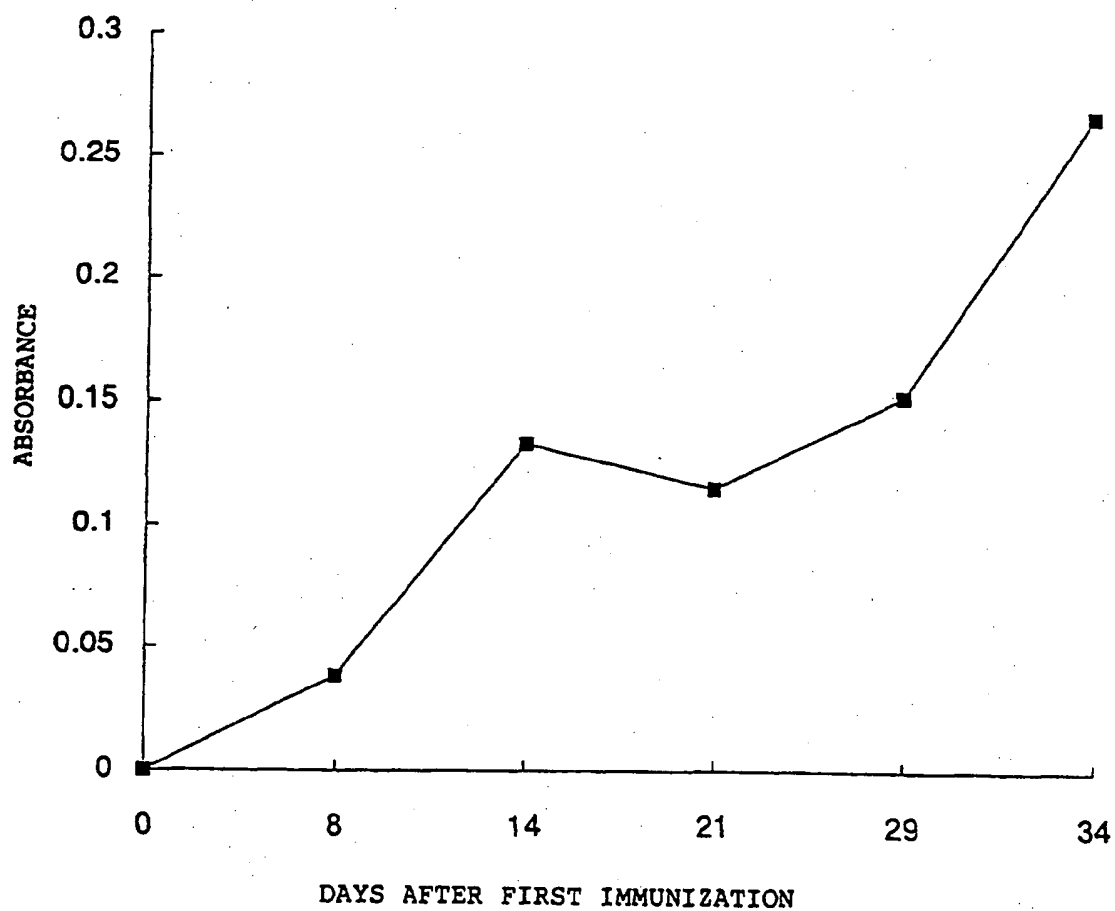
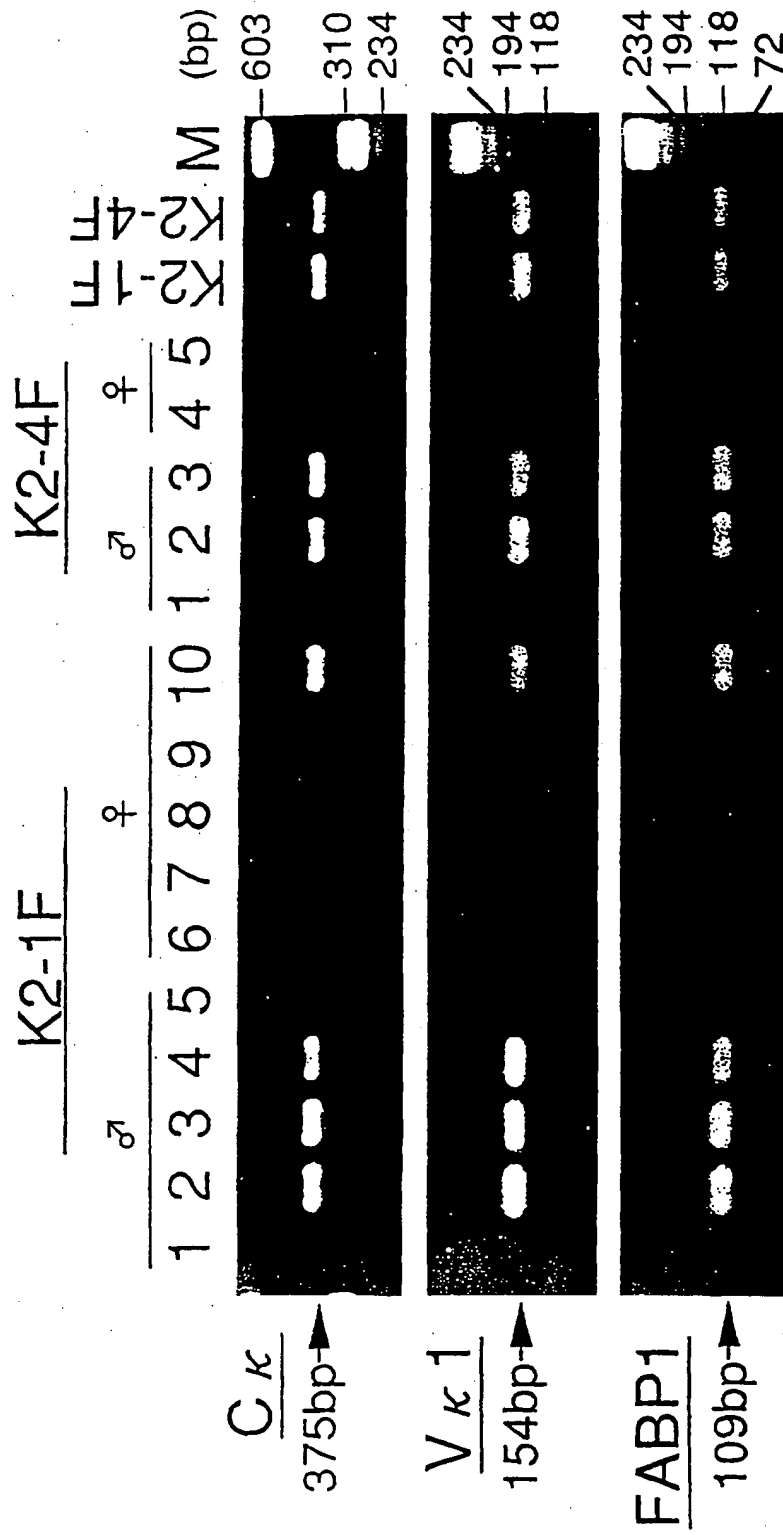


FIG. 21



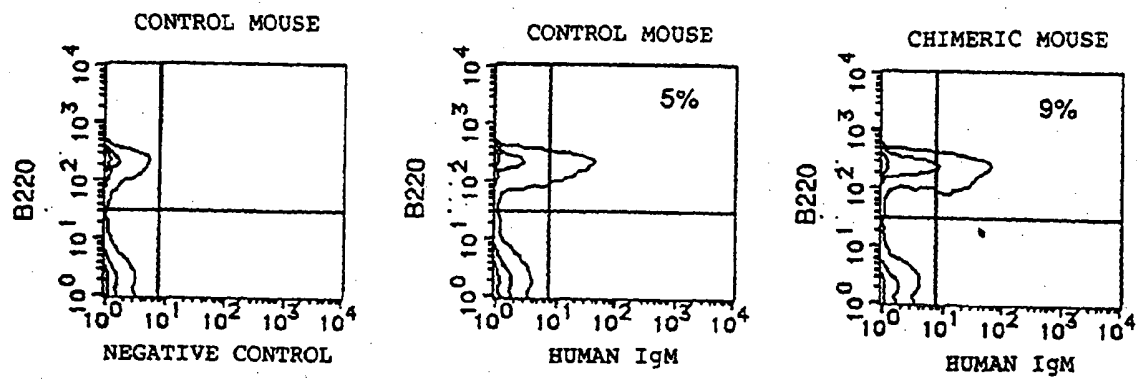


FIG. 22

FIG. 23

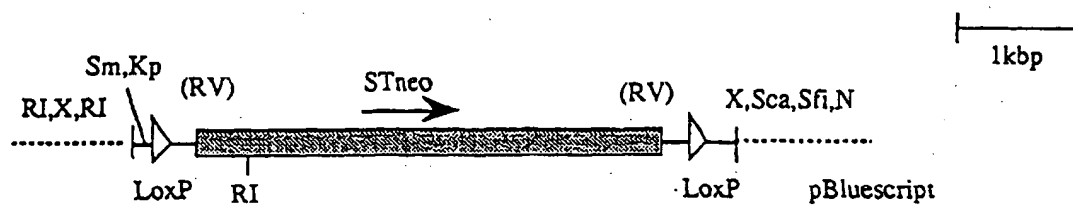


FIG. 24

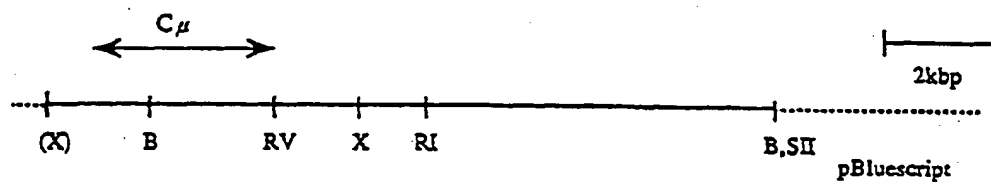


FIG. 25

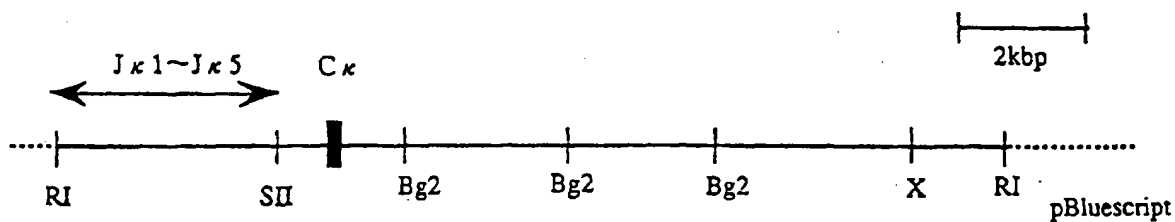


FIG. 26

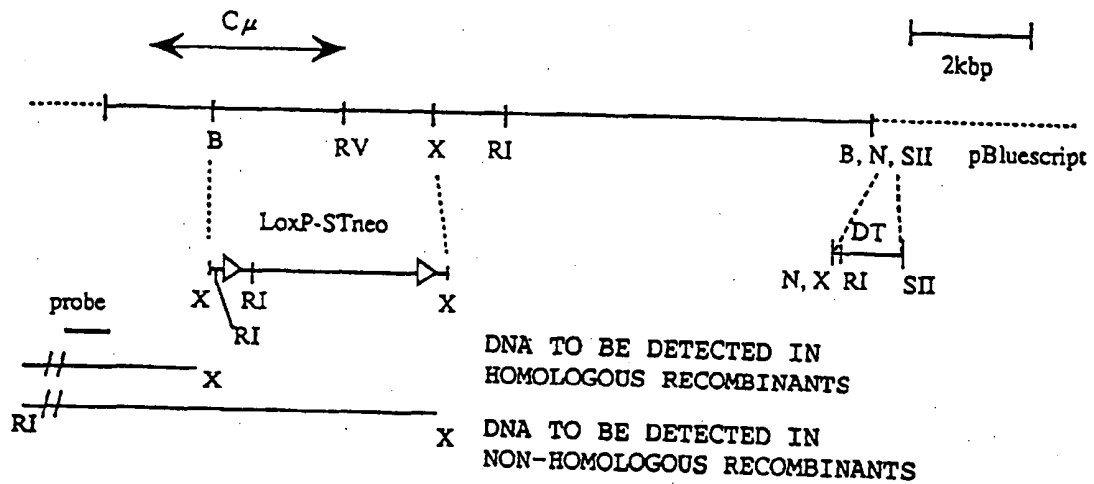


FIG. 27

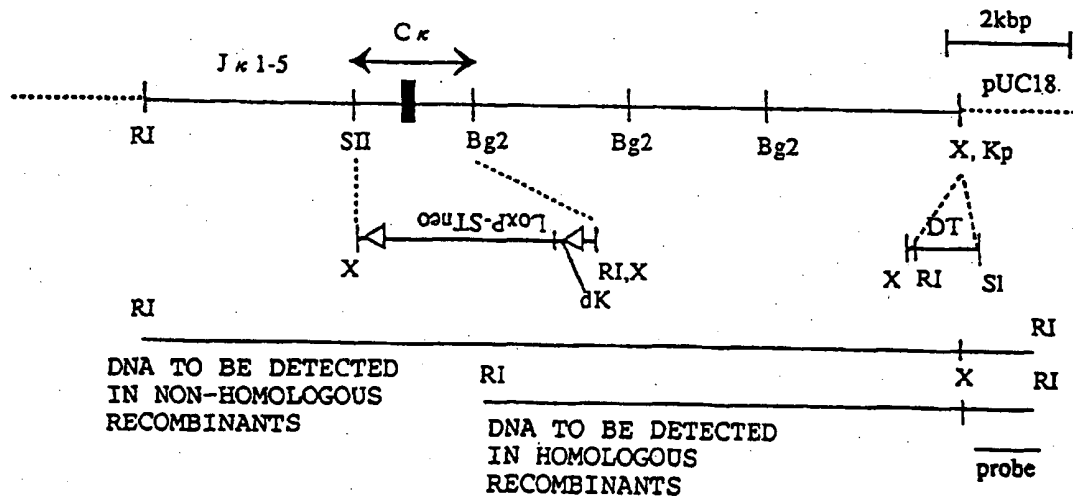


FIG. 28

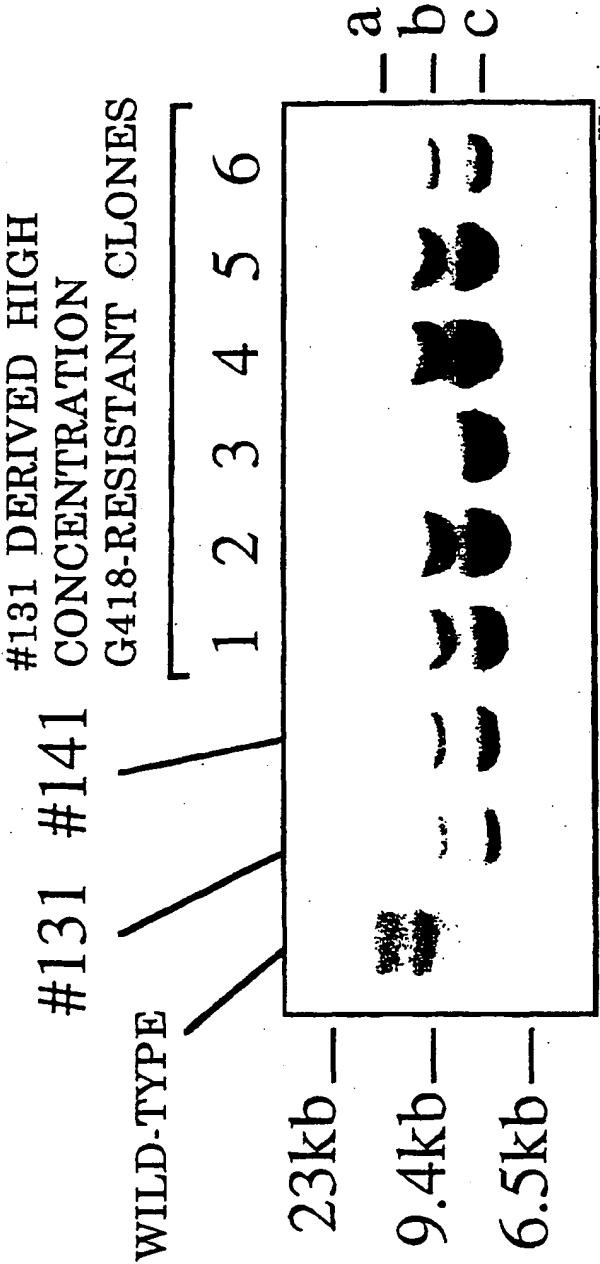


FIG. 29 TRANSFORMANT TT2F CELL CLONES

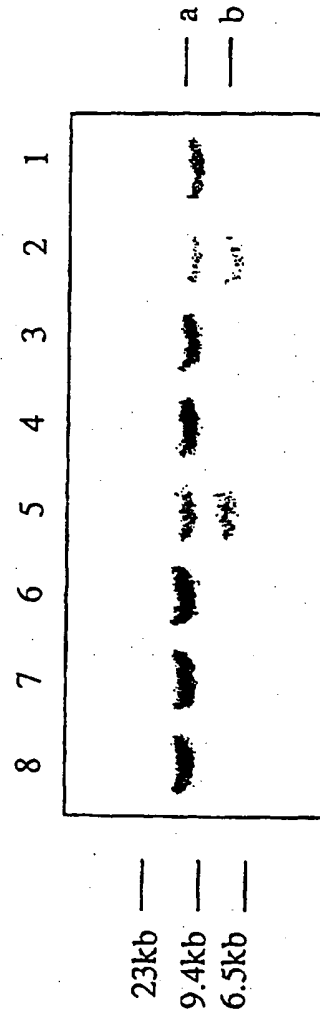




FIG. 30

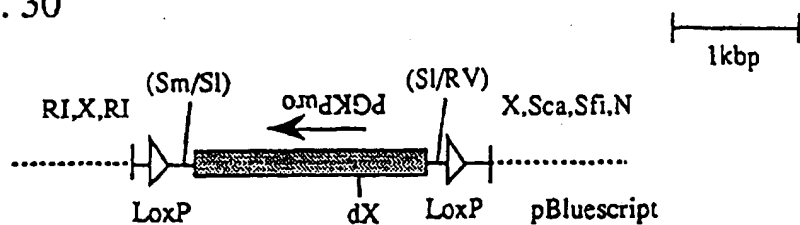
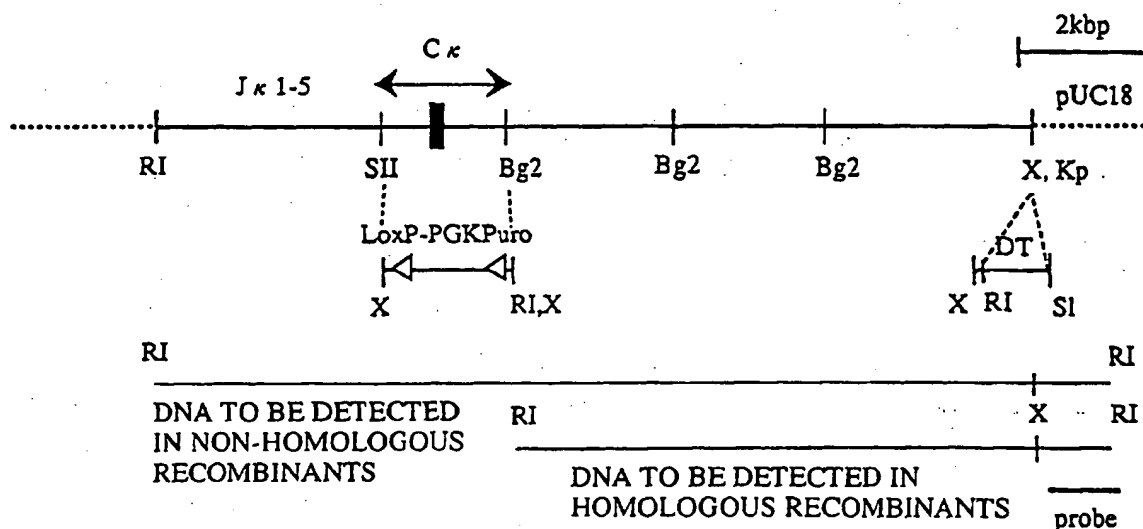


FIG. 31

**RESTRICTION ENZYMES:**

Kp: KpnI, B: BamHI, Bg2:Bgl2, RI: EcoRI, RV: EcoRV, N: Not, SI: SalI, Sca: ScaI, Sfi: SfiI,  
 Sm: Sma, X: XhoI, (X): XhoI RESTRICTION SITE FROM  $\lambda$  VECTOR  
 dK: KpnI RESTRICTION SITE DELETED, dX: XhoI RESTRICTION SITE DELETED  
 (Sm/SI): LIGATED TO SmaI RESTRICTION SITE AFTER SalI BLUNTING  
 (SI/RV): LIGATED TO EcoRV RESTRICTION SITE AFTER SalI BLUNTING  
 DOTTED PORTION: pBluescript SKII(+) OR pUC18 PLASMID DNA

◁▷ : LoxP SEQUENCE

FIG. 32  
HIGH CONCENTRATION G418 RESISTANT TT2F CELL CLONES

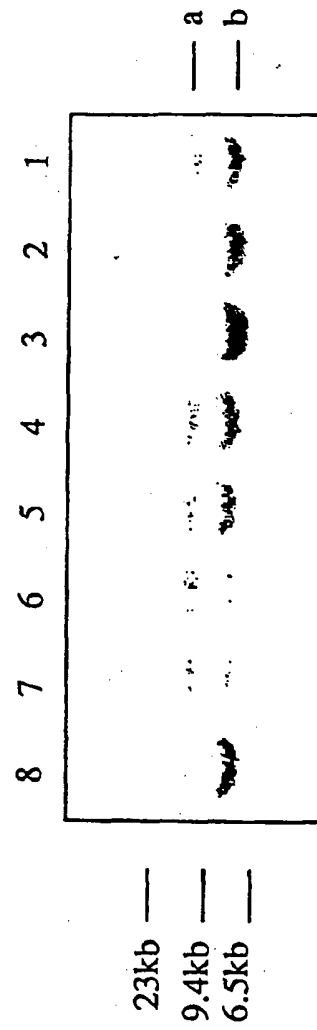
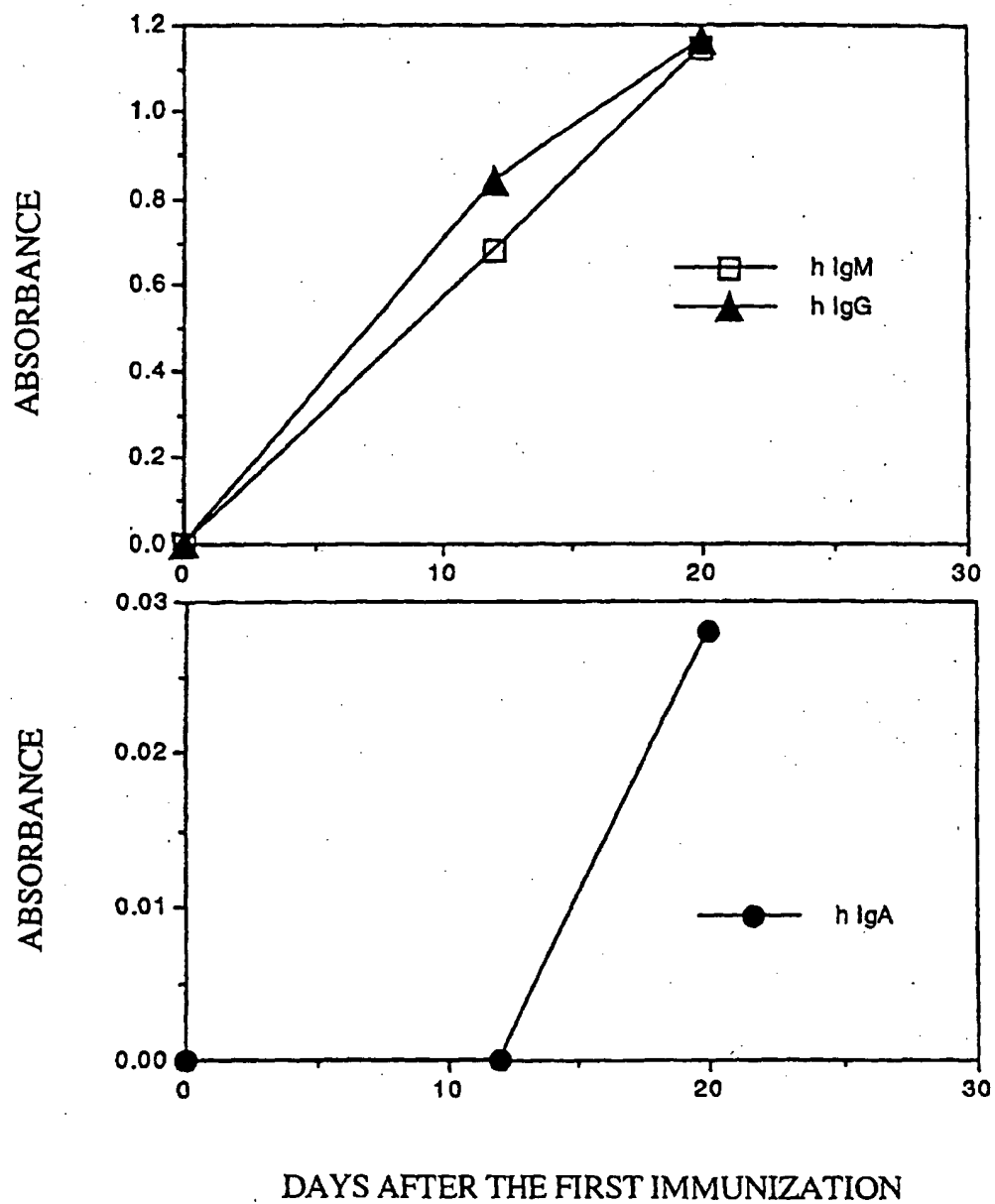


FIG. 33



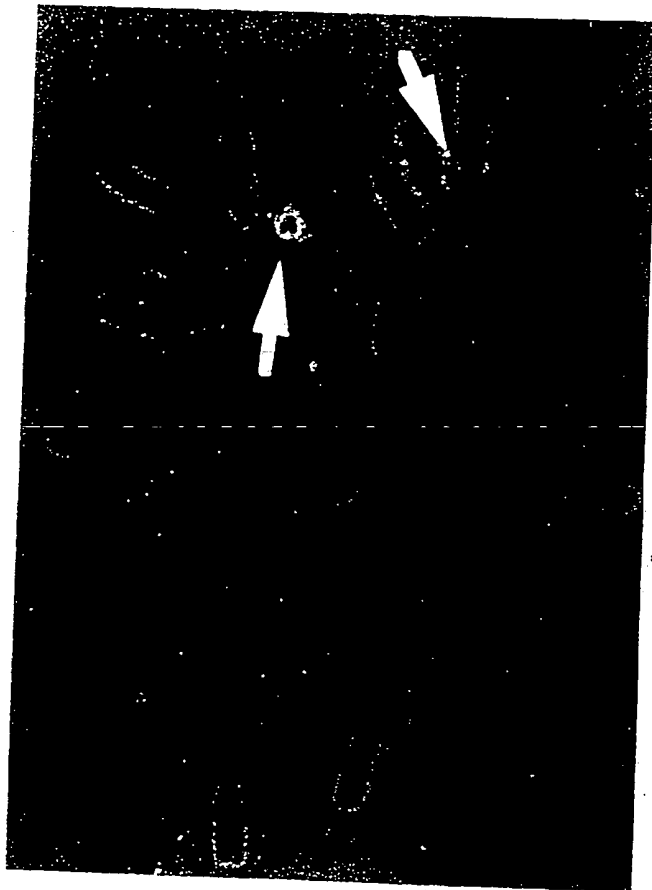


FIG. 34

FIG. 35

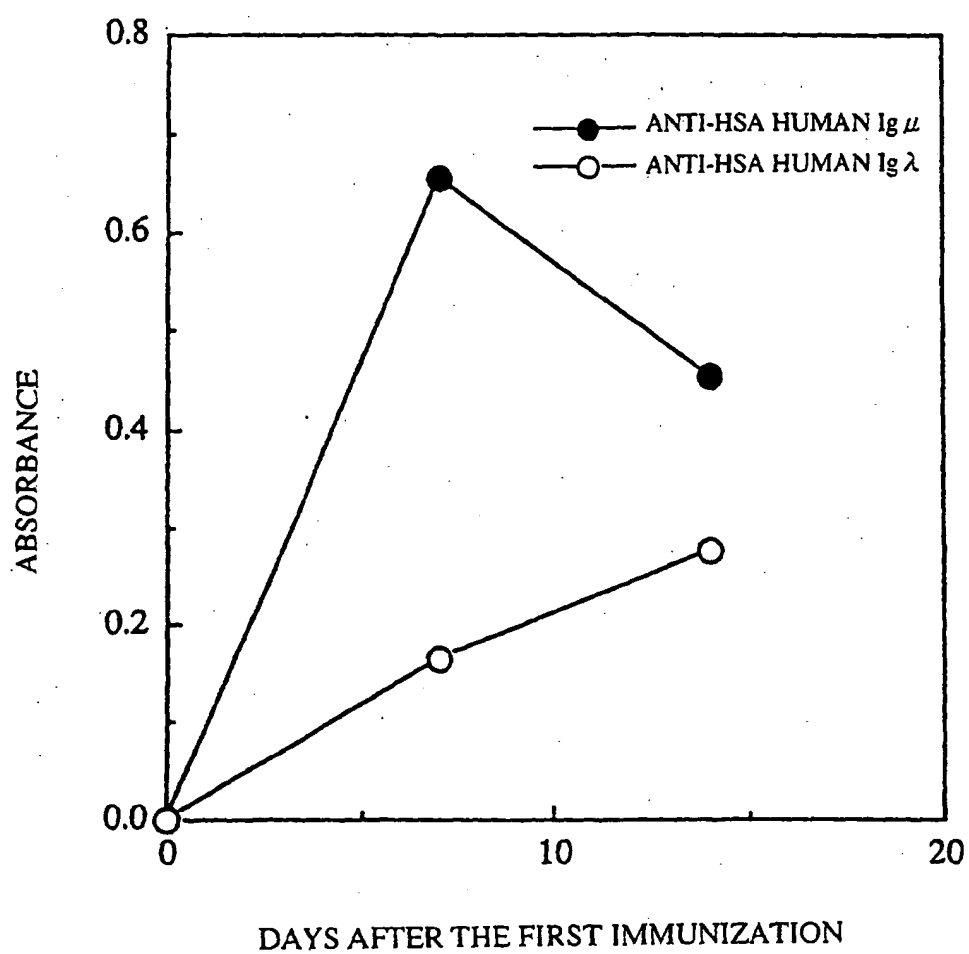




FIG. 36

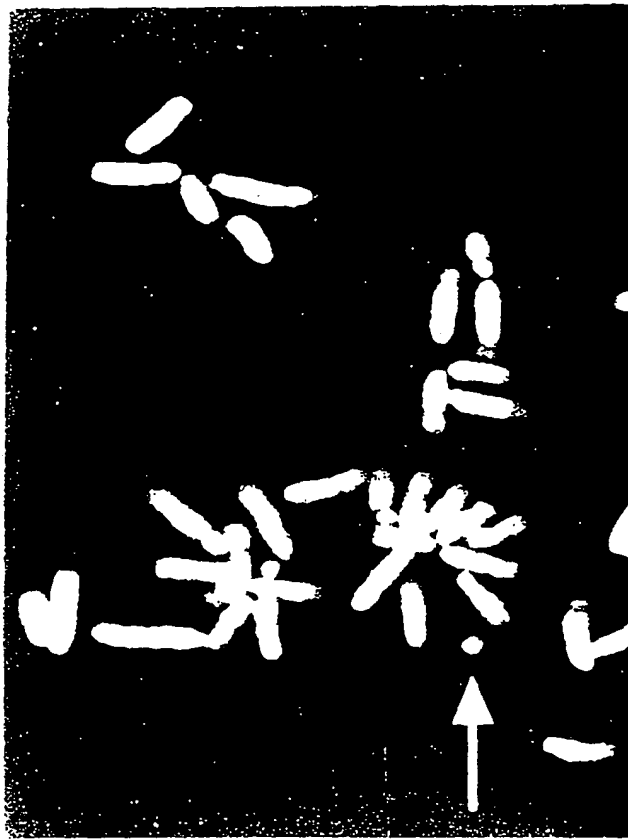


FIG. 37

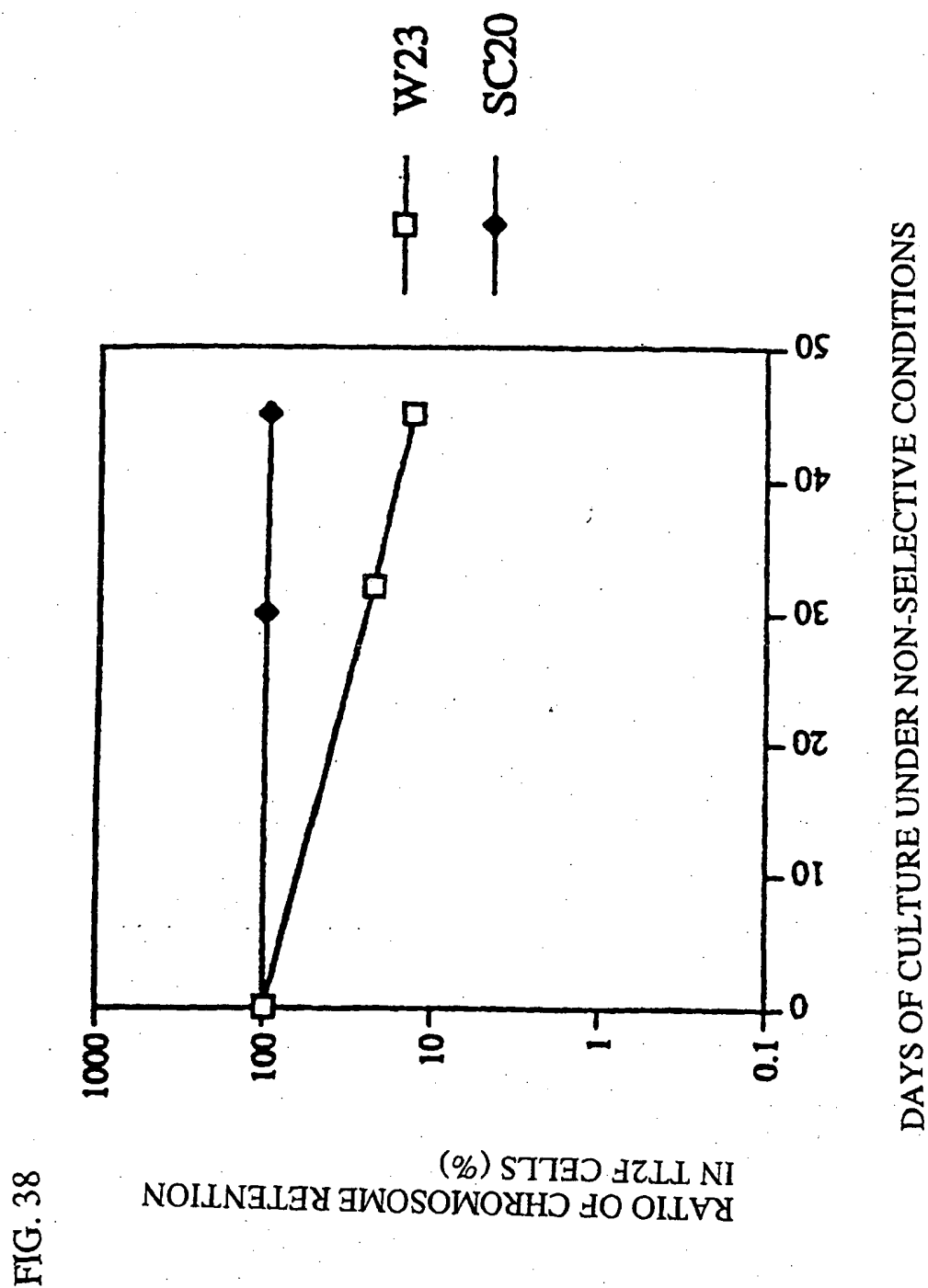




FIG. 39

	BRAIN		SPLEEN		LIVER		BONE MARROW		TESTIS			
									1ST MEIOSIS		2ND MEIOSIS	
	+	-	+	-	+	-	+	-	+	-	+	-
16-5 (F1)	30 (100%)	0	11 (21%)	42	30 (97%)	1	30	15 (100%)	0 (94%)	15 (94%)	11 (32%)	23
17-8 (F1)	30 (100%)	0	12 (30%)	28	30 (100%)	0	28	16 (100%)	0 (53%)	9 (53%)	14 (34%)	27
17-23 (F1)	41 (95%)	2	7 (17%)	34	31 (97%)	1	25	15 (100%)	0 (81%)	13 (81%)	13 (52%)	12

RETENTION OF SC20 FRAGMENT IN MICE

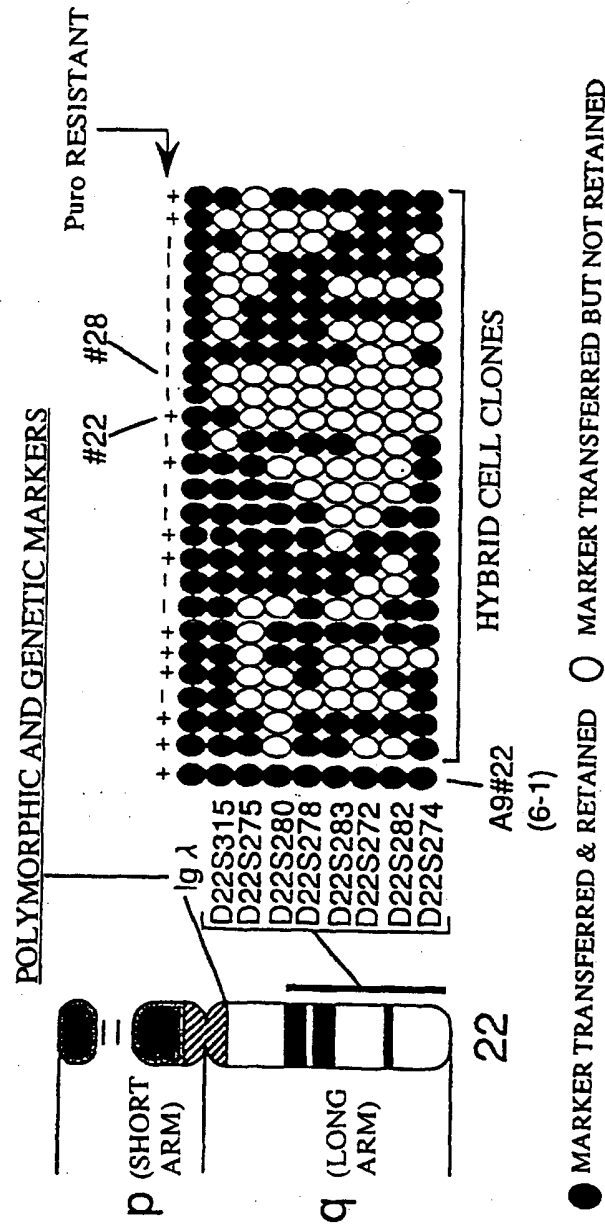
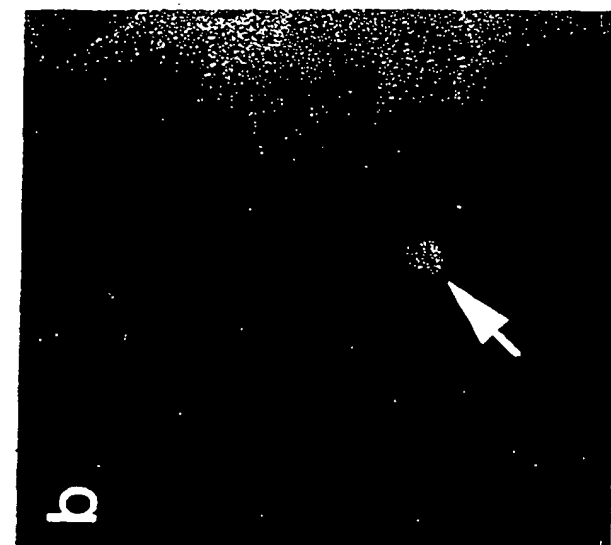
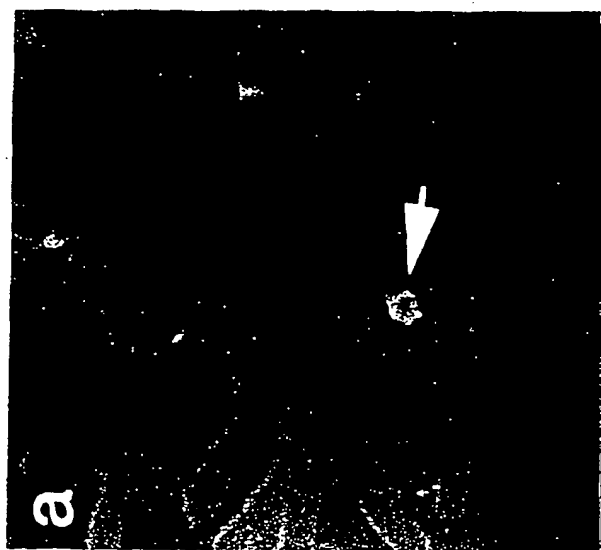


FIG. 40



b) CONTROL



a) CLONE #28

FIG. 41

	HK23	HK28	HK29
ANTIBODY HEAVY-CHAIN KNOCKOUT	HOMO	WILD-TYPE OR HETERO	WILD-TYPE OR HETERO
ANTIBODY LIGHT-CHAIN KNOCKOUT	HETERO	WILD-TYPE	HETERO
W23 FRAGMENT	+	+	+
SC20 FRAGMENT	+	+	+
HUMAN $\mu$ CHAIN IN SERUM (mg/l)	100	5.9	14
HUMAN $\kappa$ CHAIN IN SERUM (mg/l)	8.6	8.4	25
HUMAN $\mu/\kappa$ CHAINS IN SERUM (mg/l)	18	0.13	BELOW DETECTION LIMIT

\* SINCE "ANTIBODY HEAVY-CHAIN KNOCKOUT" IS JUDGED BY THE PRESENCE OR ABSENCE OF THE EXPRESSION OF MOUSE  $\mu$  CHAIN, IT IS IMPOSSIBLE TO DISCRIMINATE HETERO FROM WILD-TYPE.

\* "ANTIBODY LIGHT-CHAIN KNOCKOUT" IS JUDGED BY SOUTHERN BLOT ANALYSIS.

FIG. 42

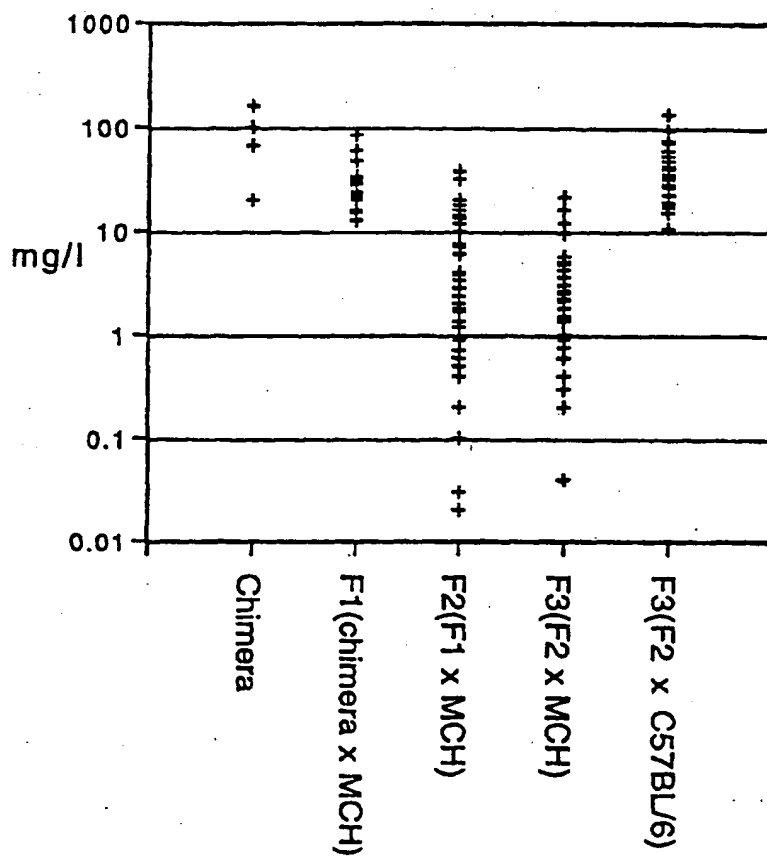
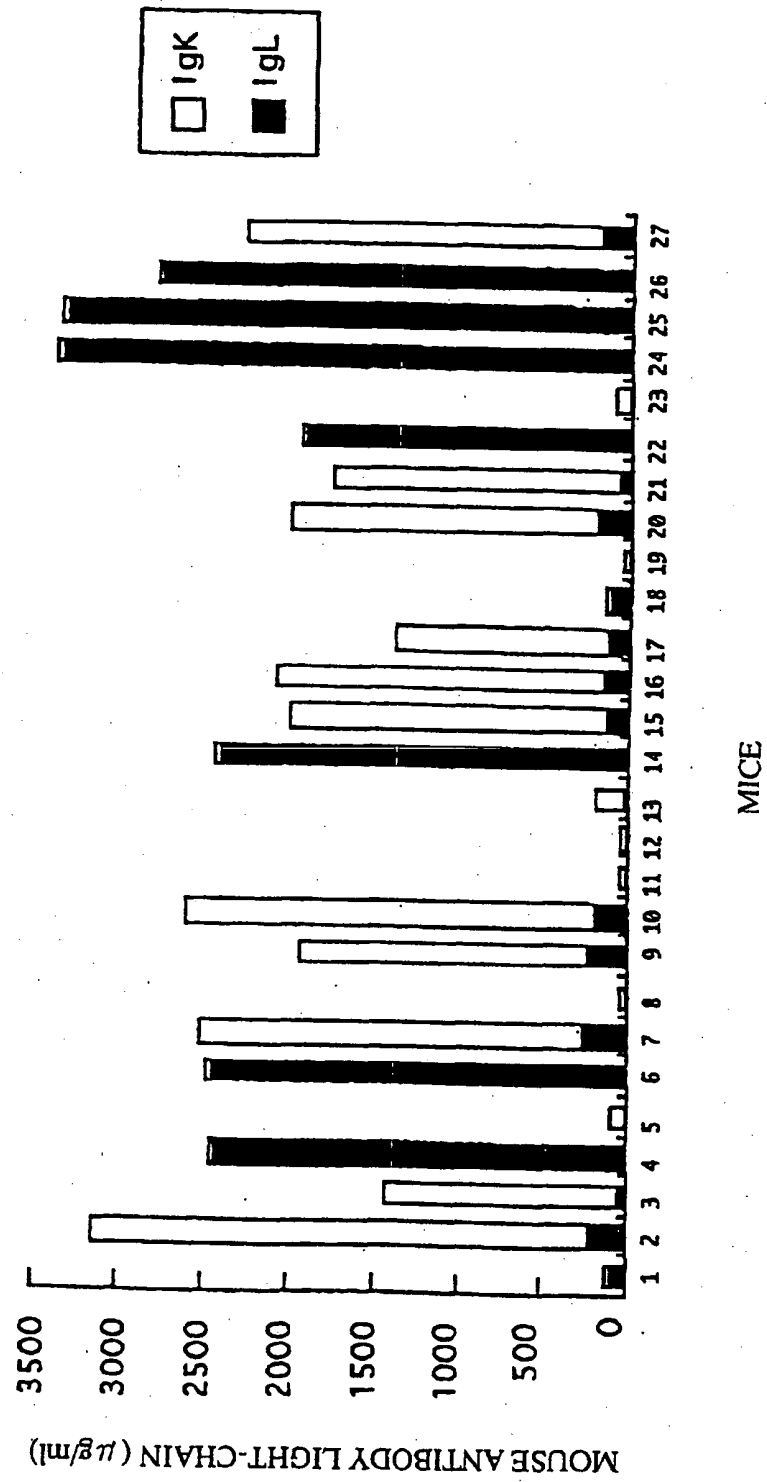


FIG. 43

FIG. 44



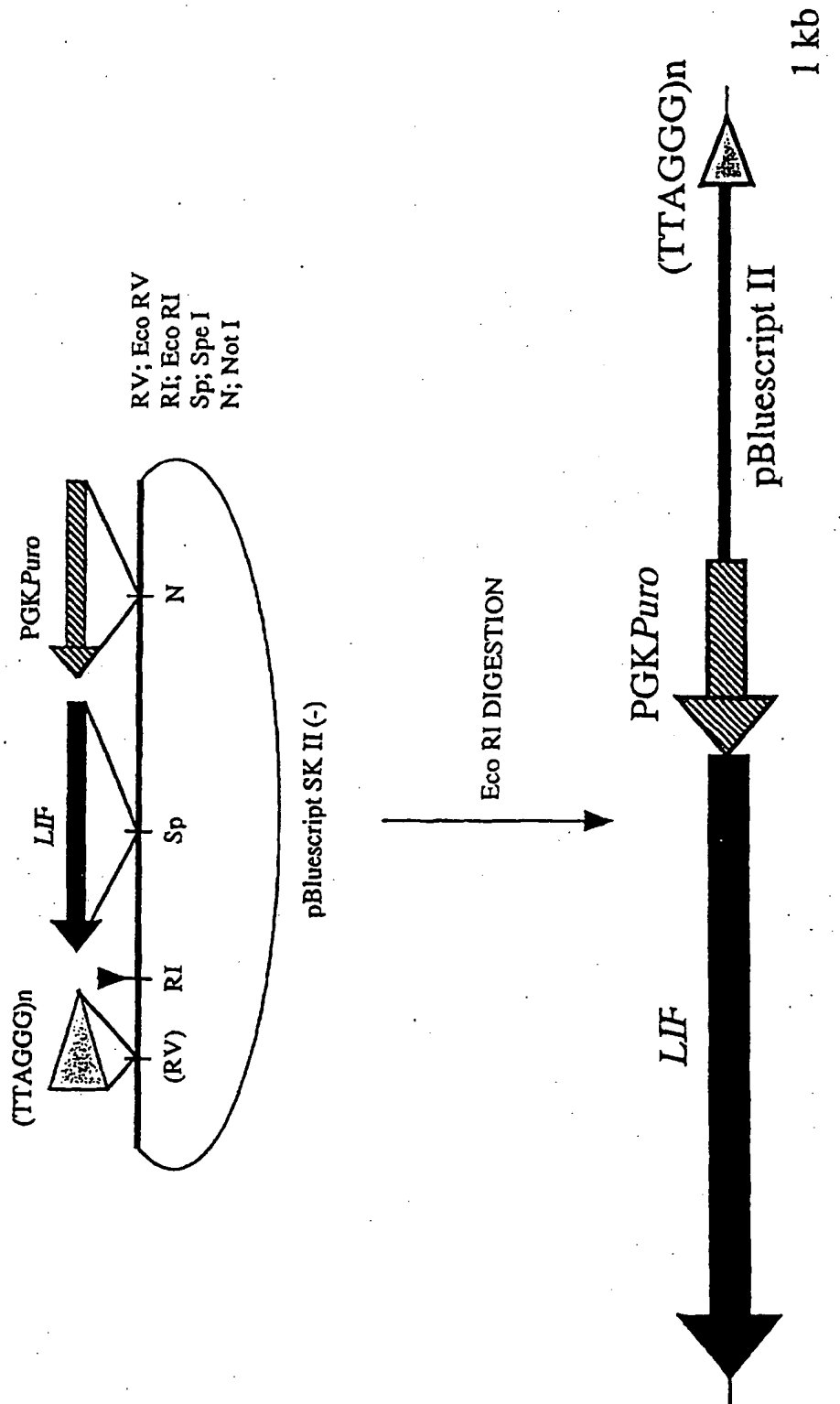


FIG. 45

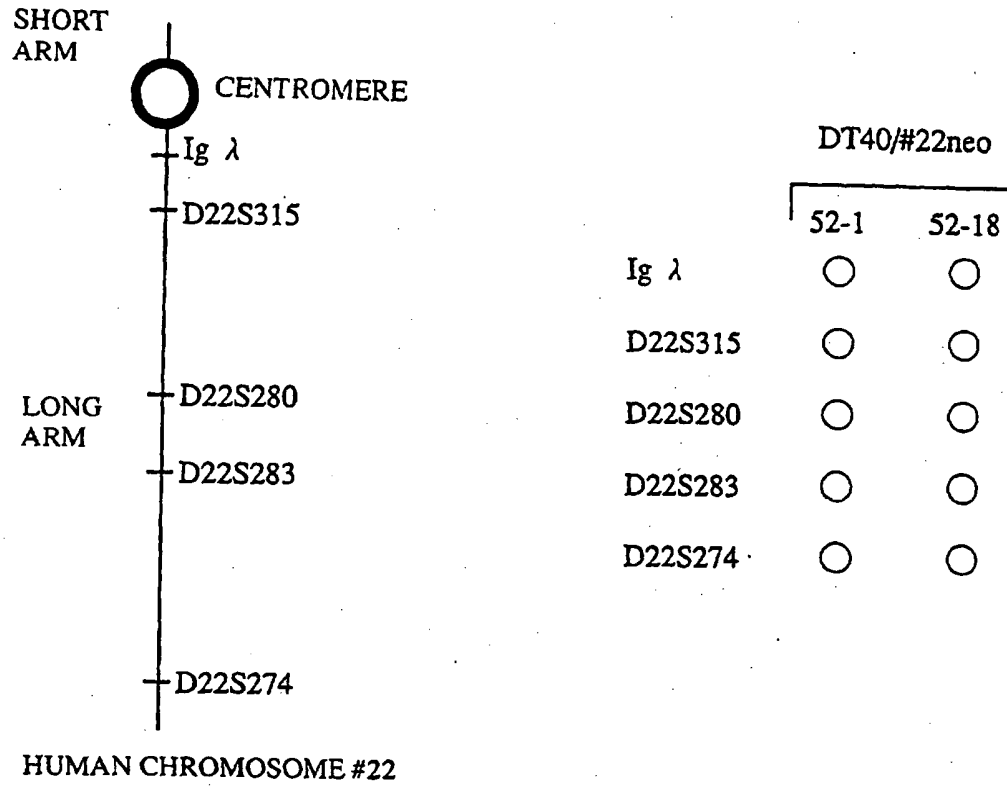


FIG. 46



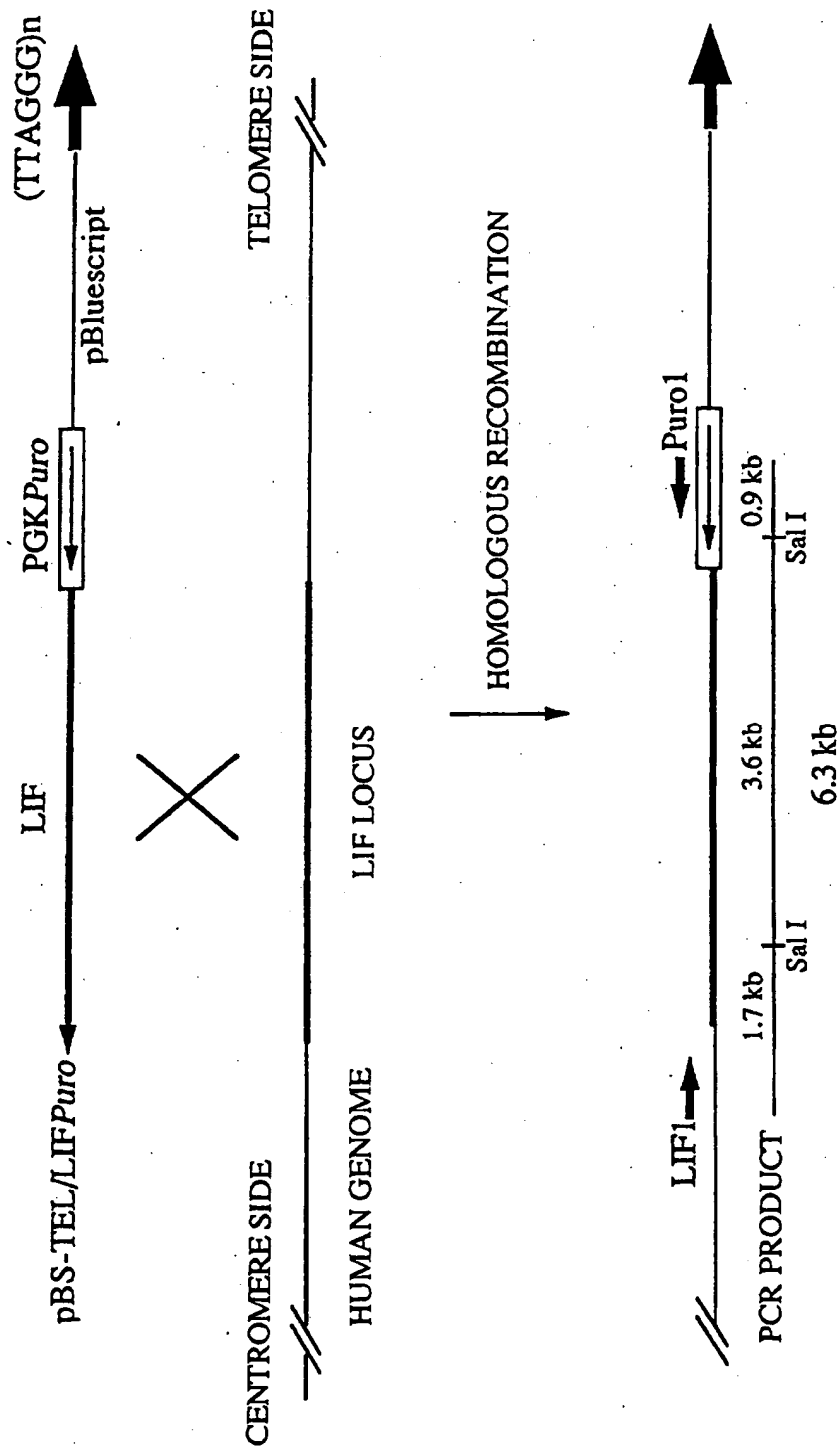


FIG. 47

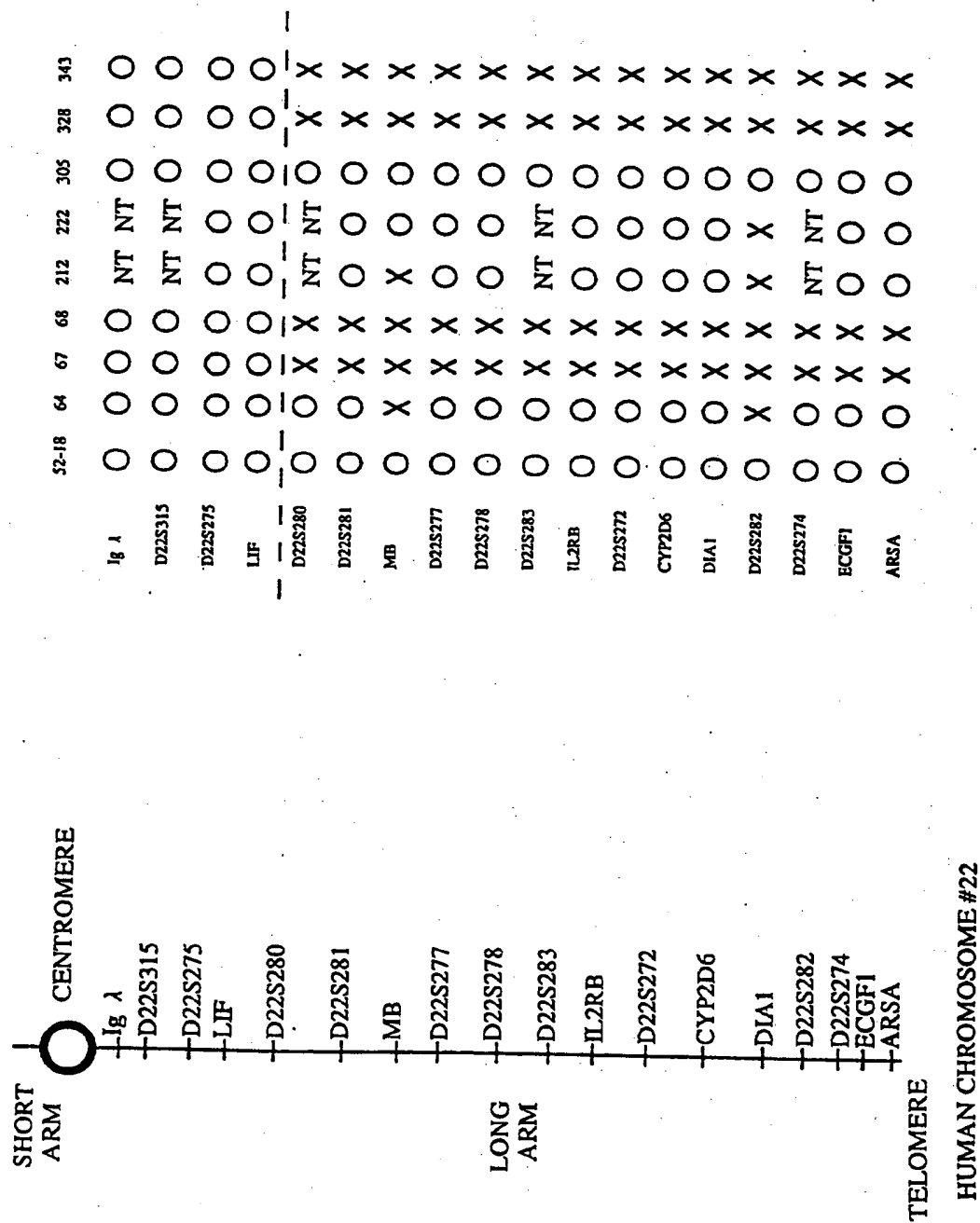


FIG. 48

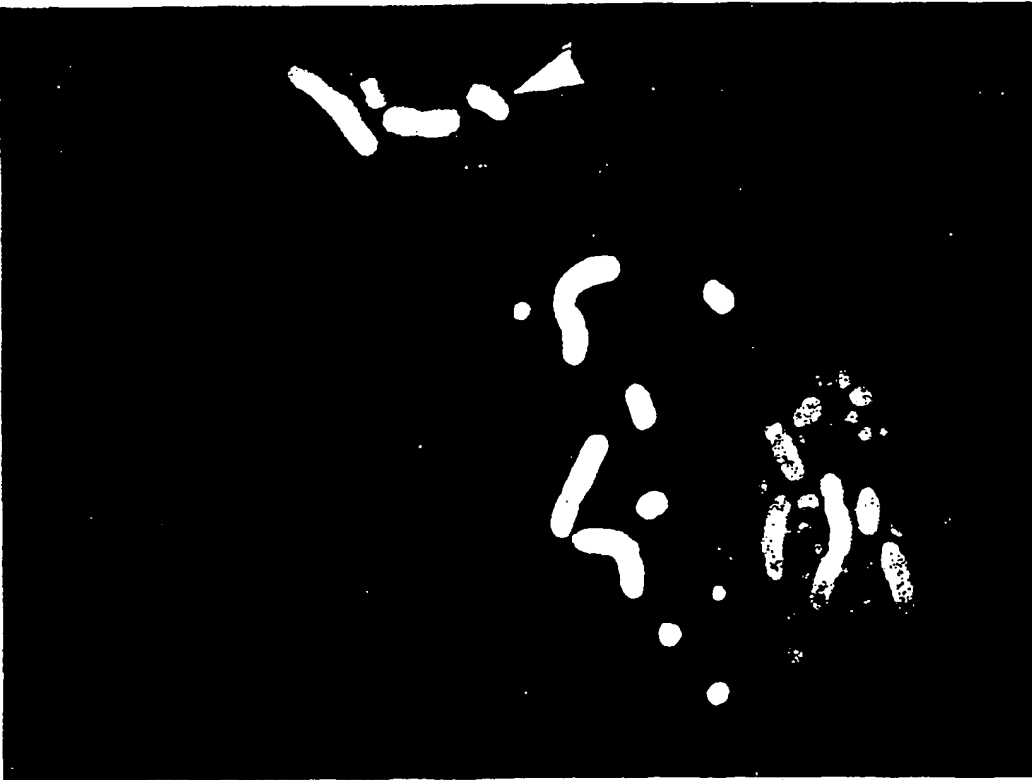
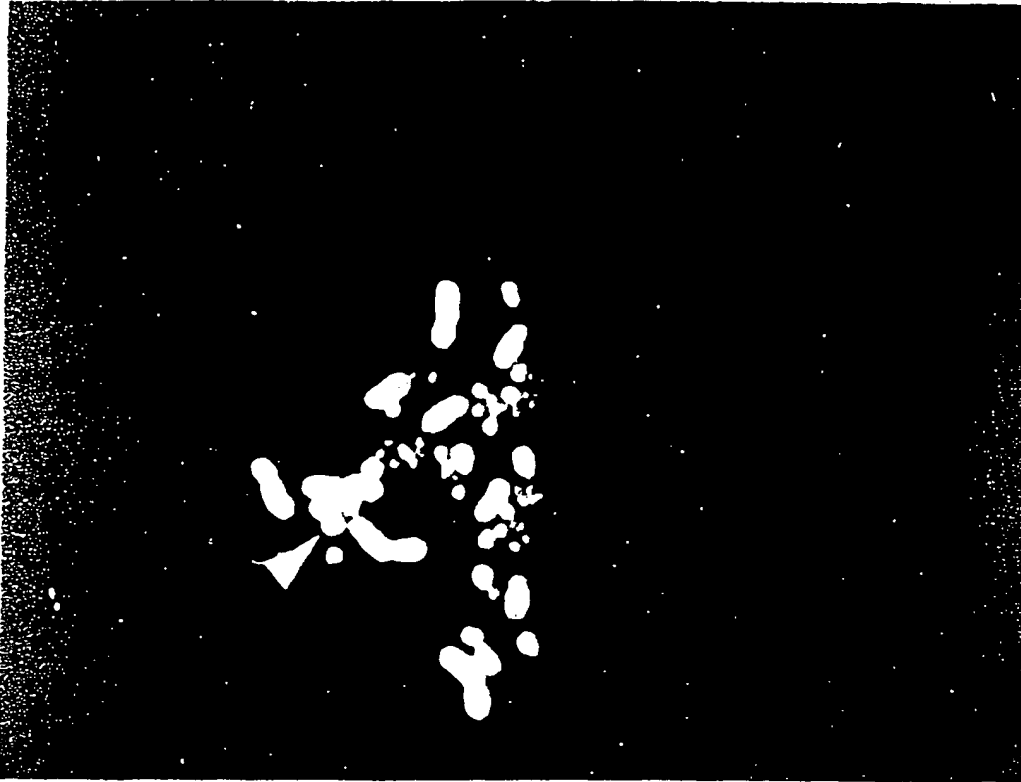


FIG. 49

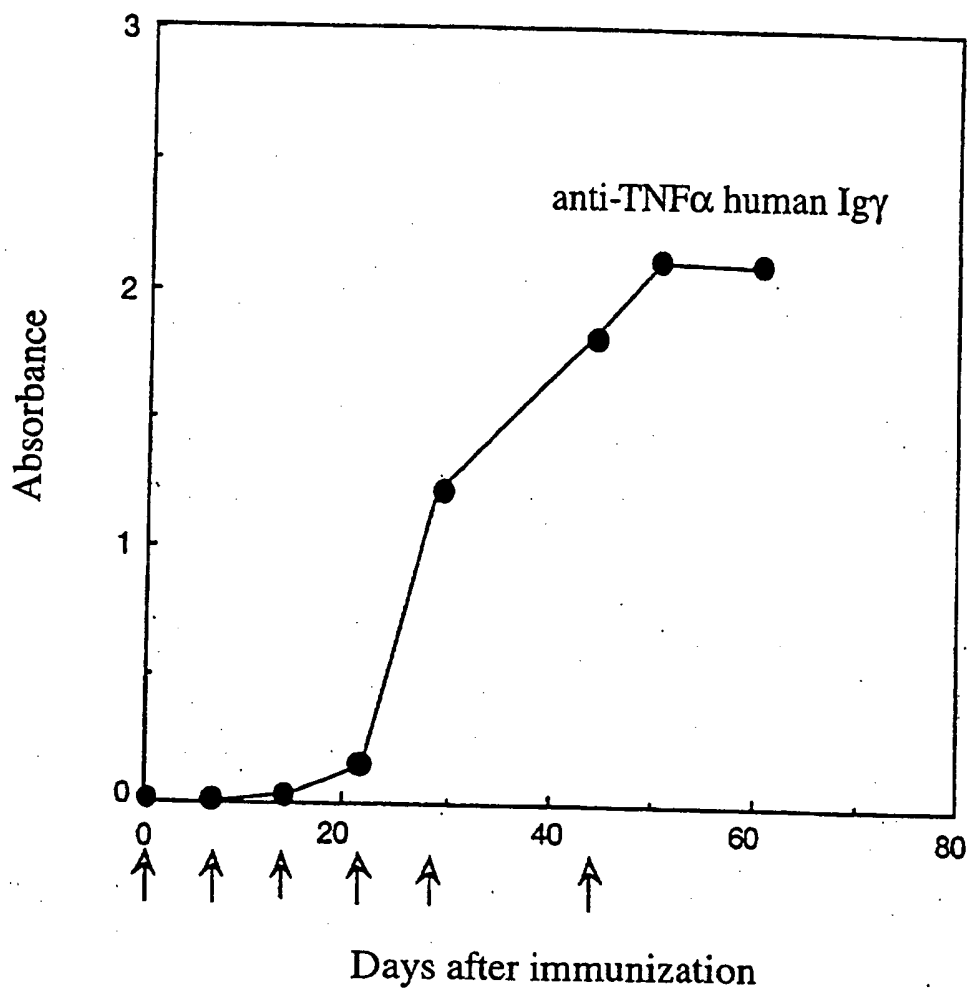


FIG. 50

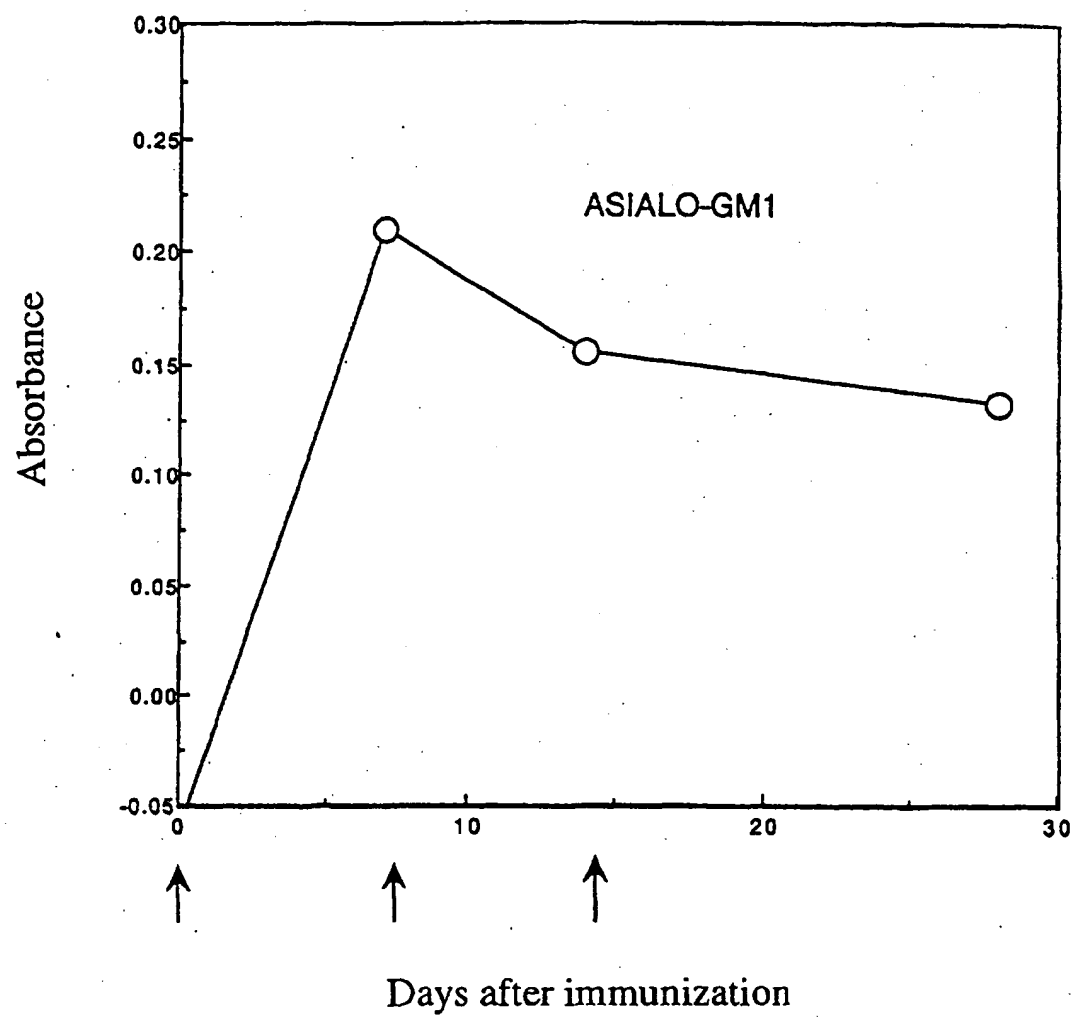


FIG. 51

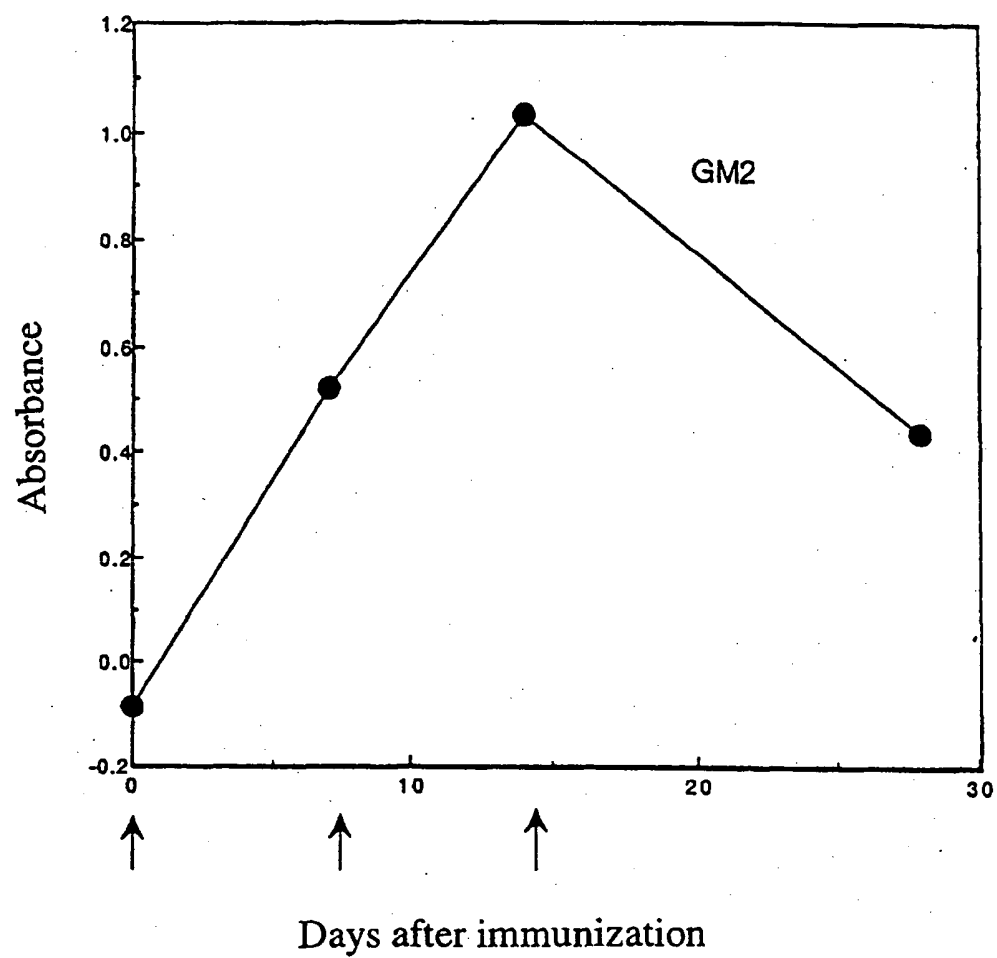
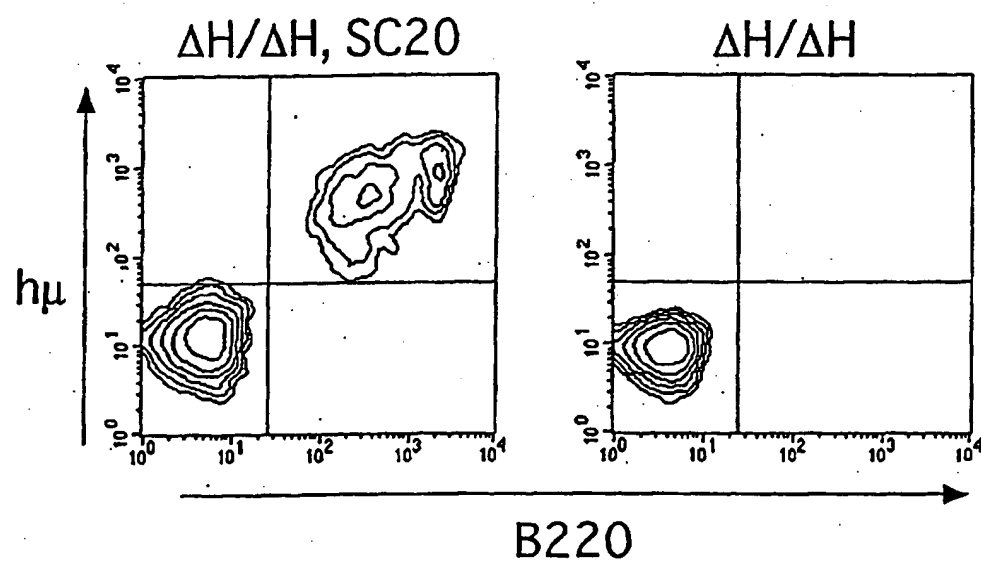


FIG. 52



$\Delta H/\Delta H, SC20$ : Single Tc/KO mouse

$\Delta H/\Delta H$ : antibody heavy-chain KO mouse

FIG. 53

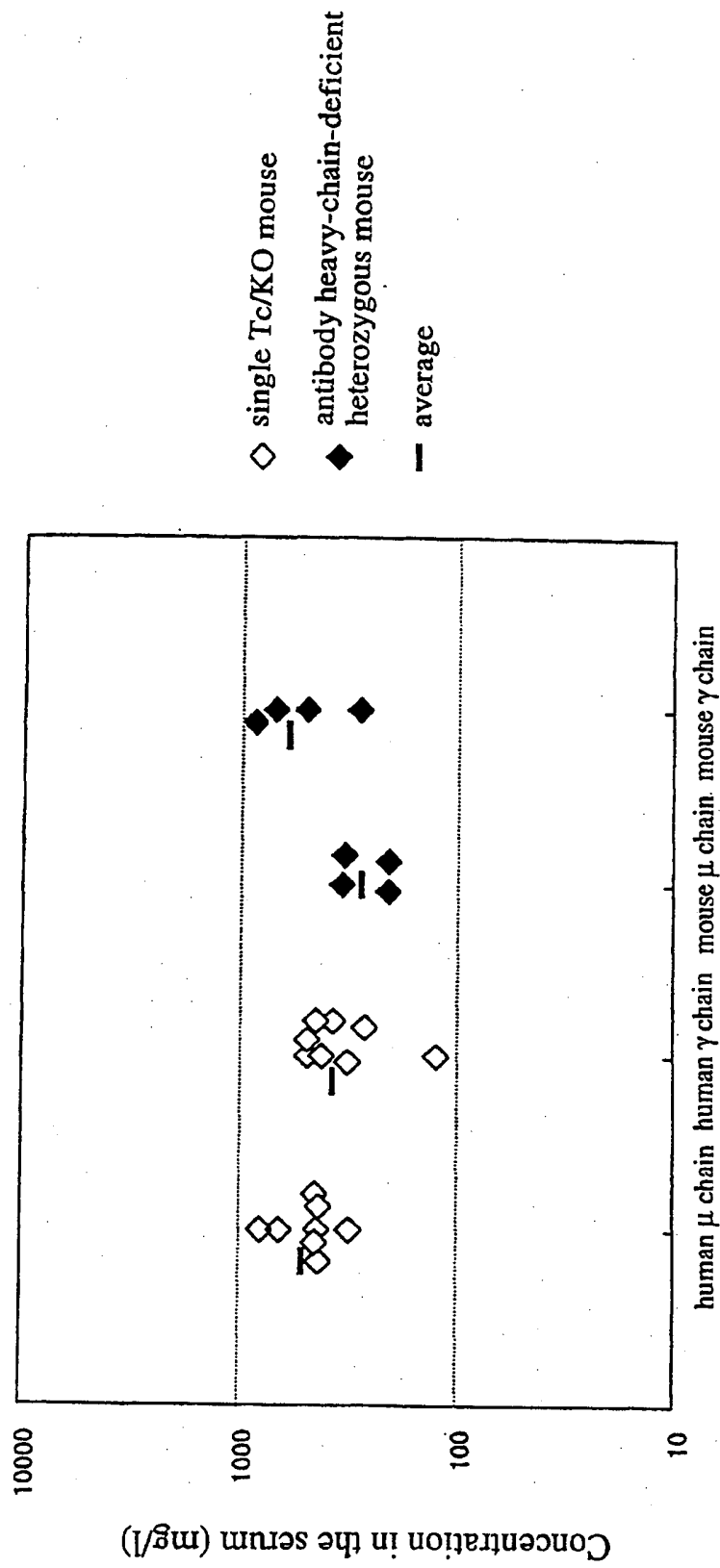


FIG. 54



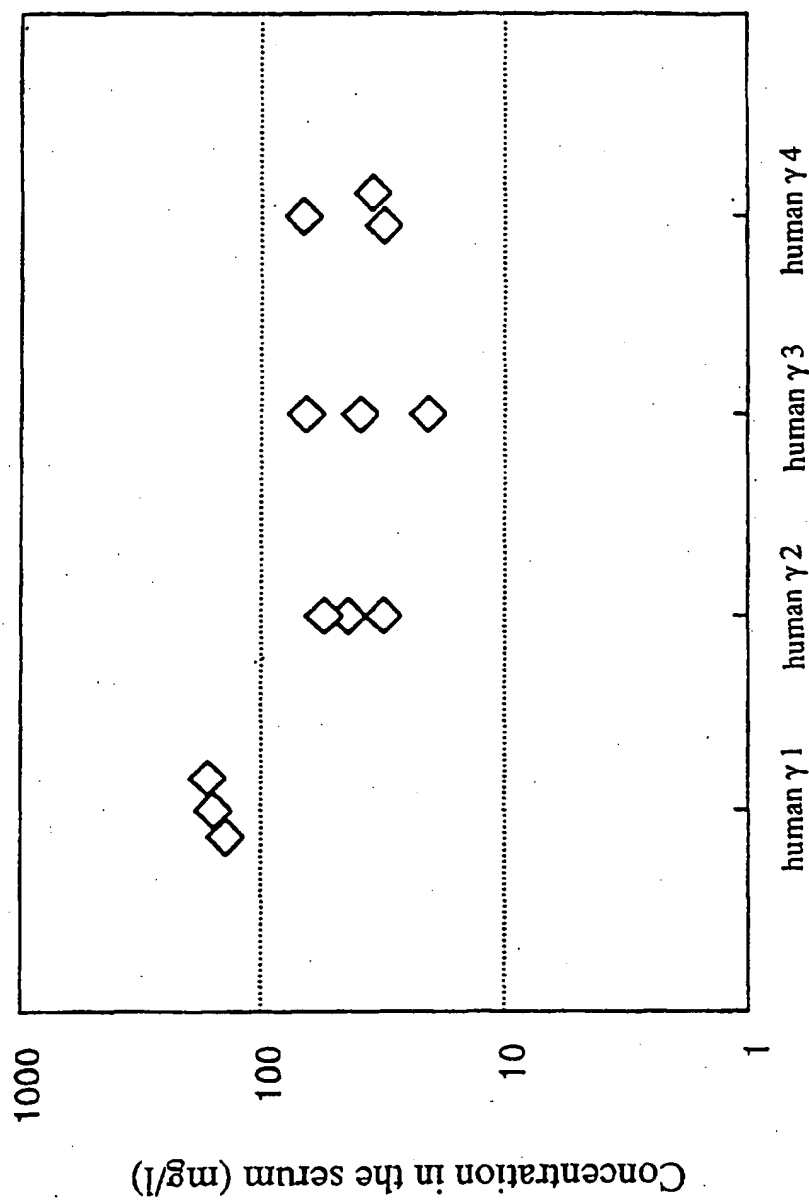


FIG. 55

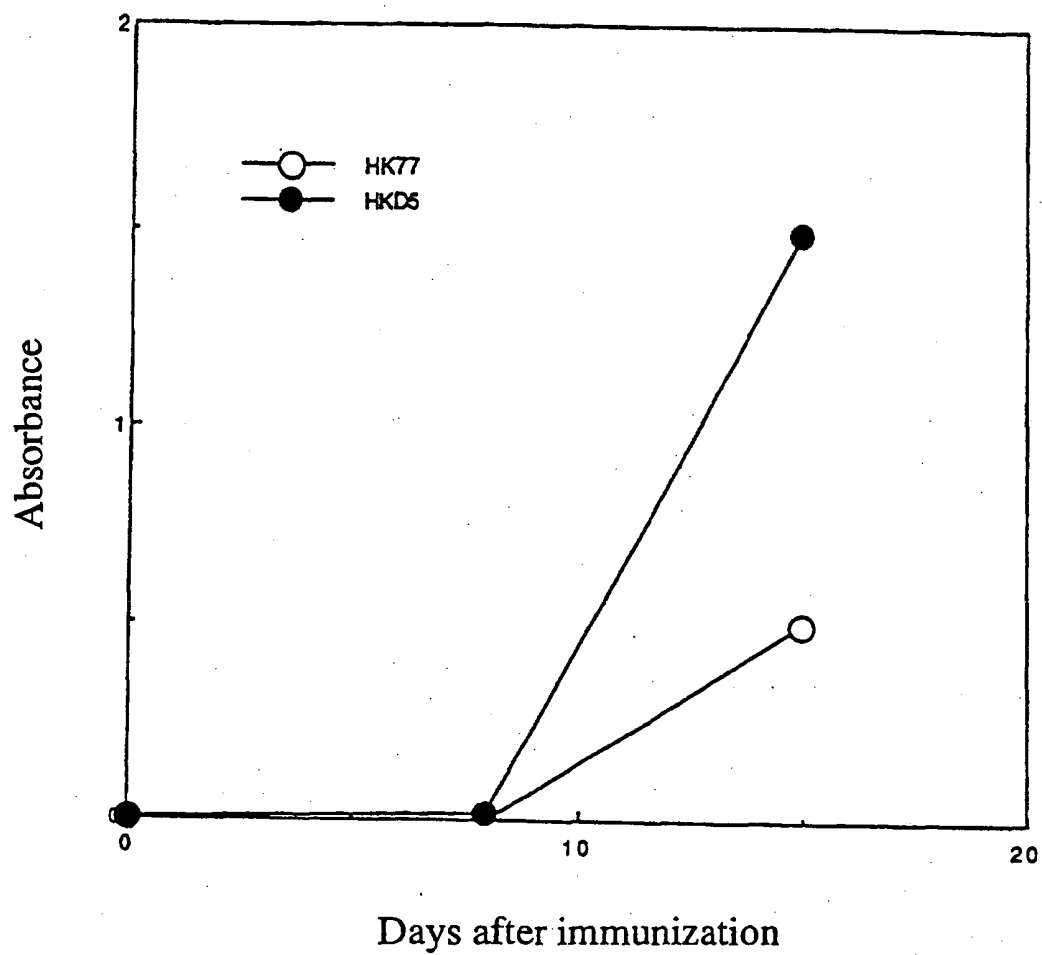


FIG. 56

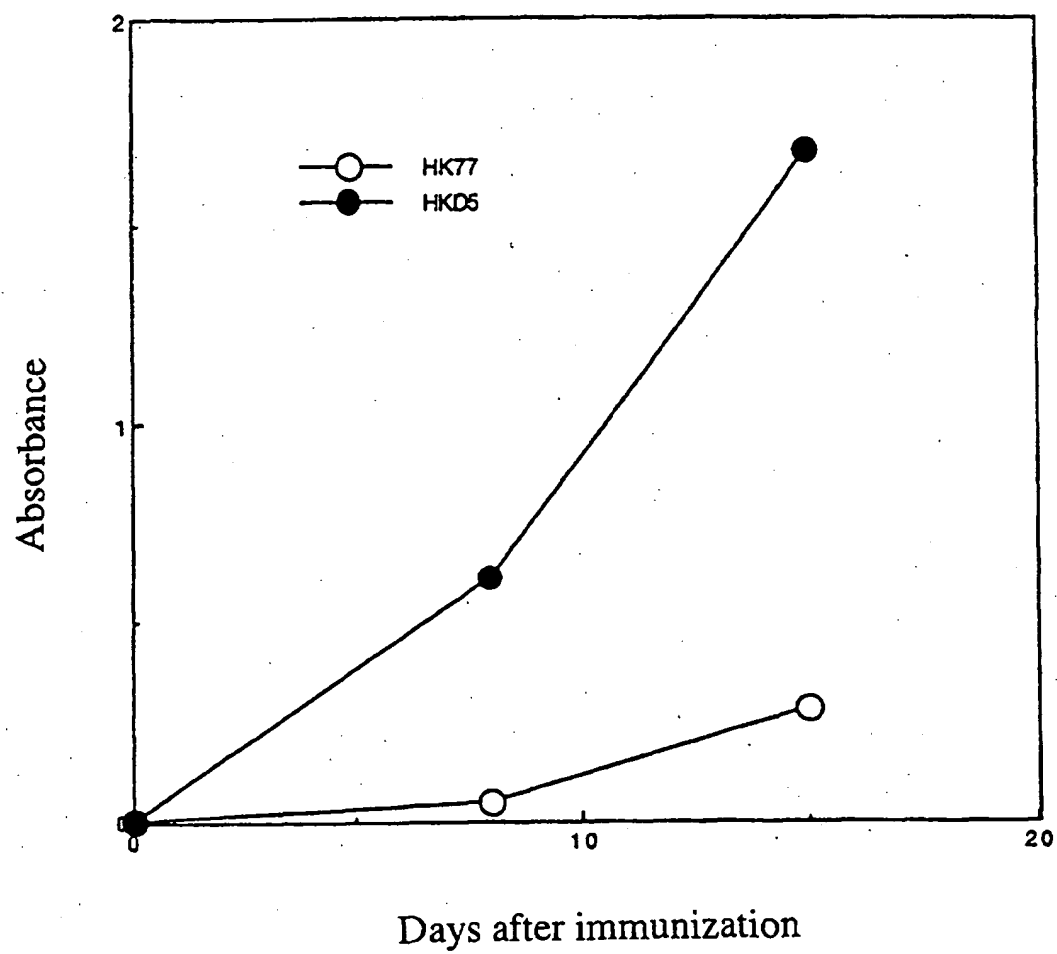


FIG. 57

FIG. 58

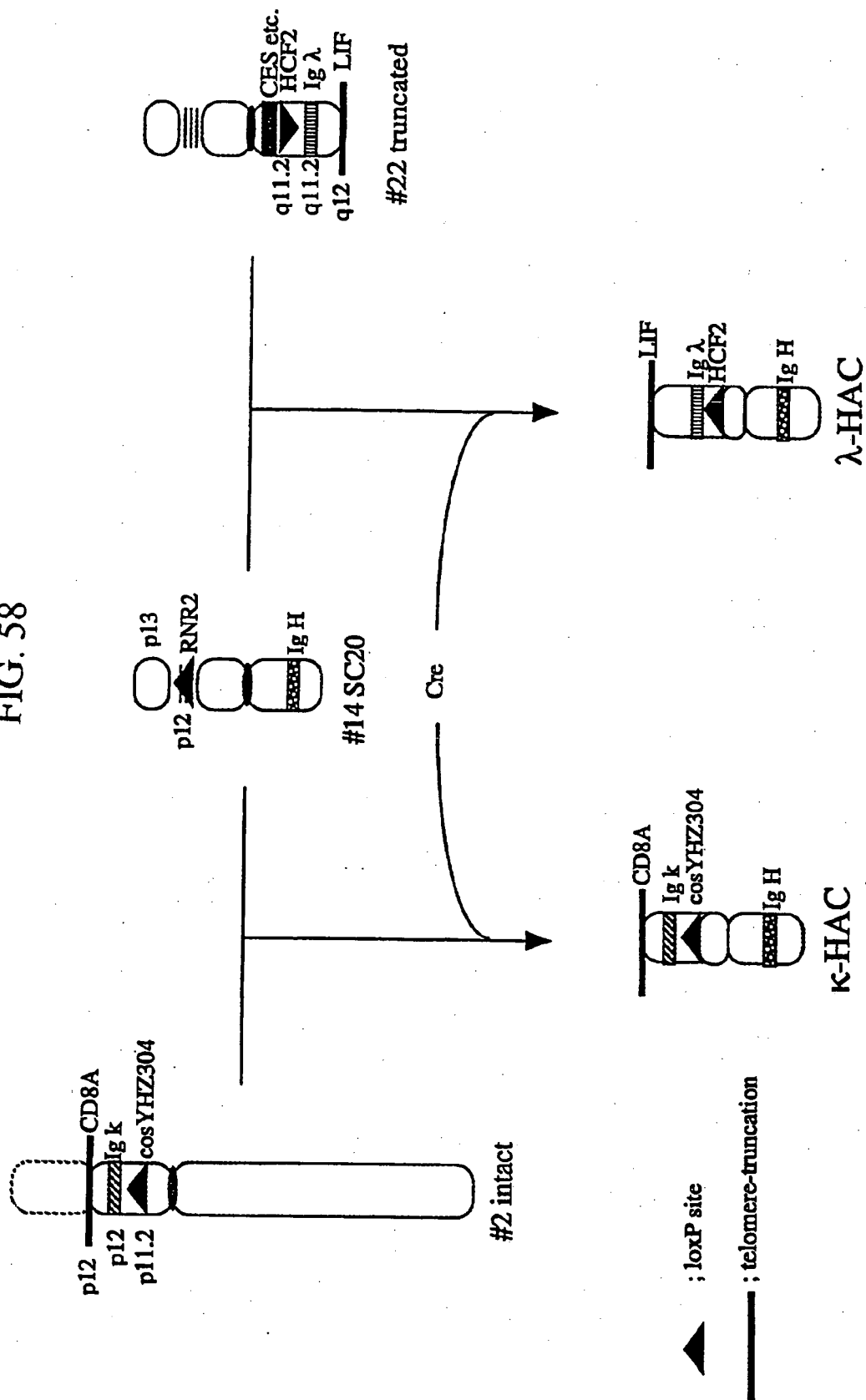


FIG. 59

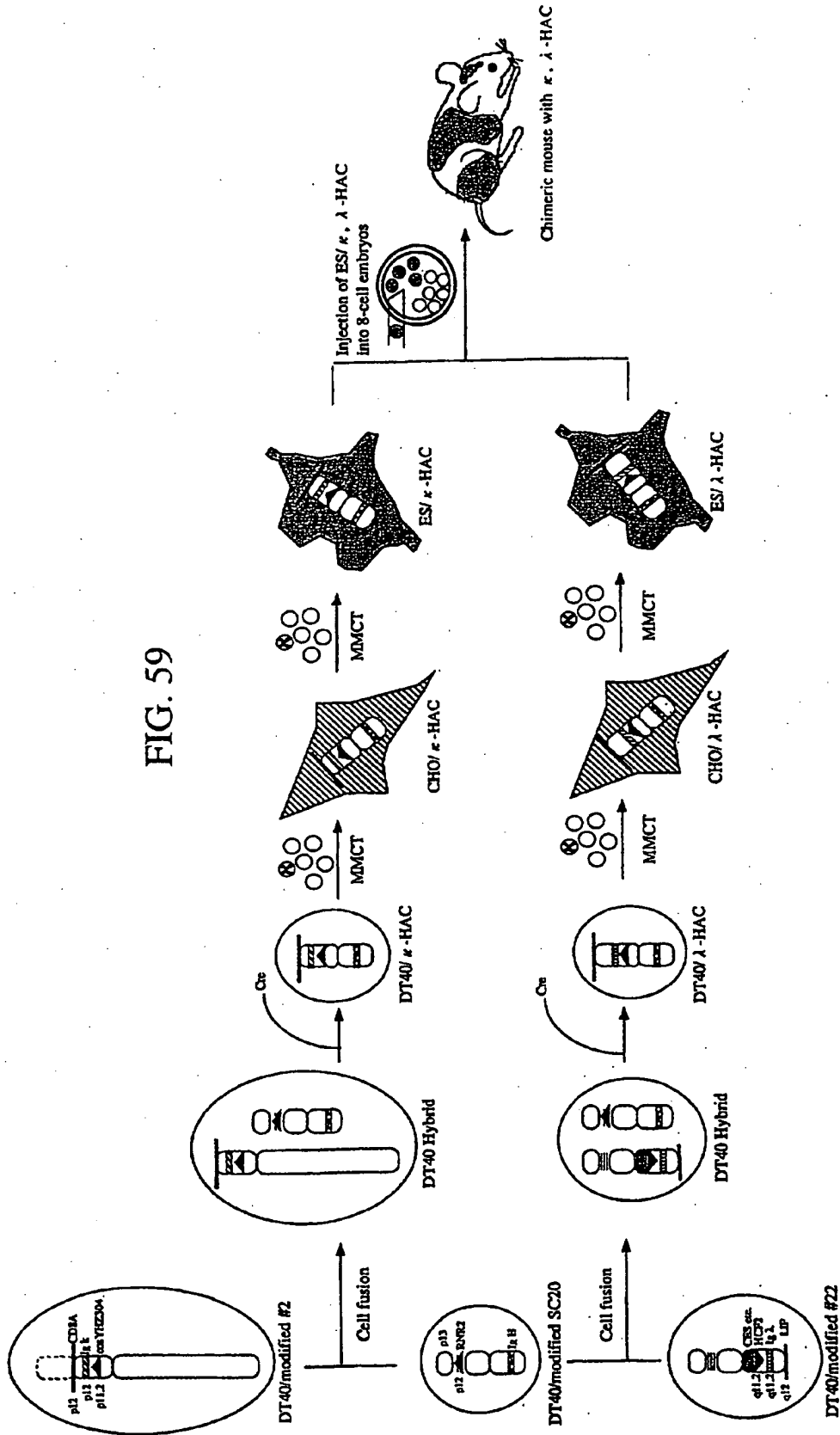
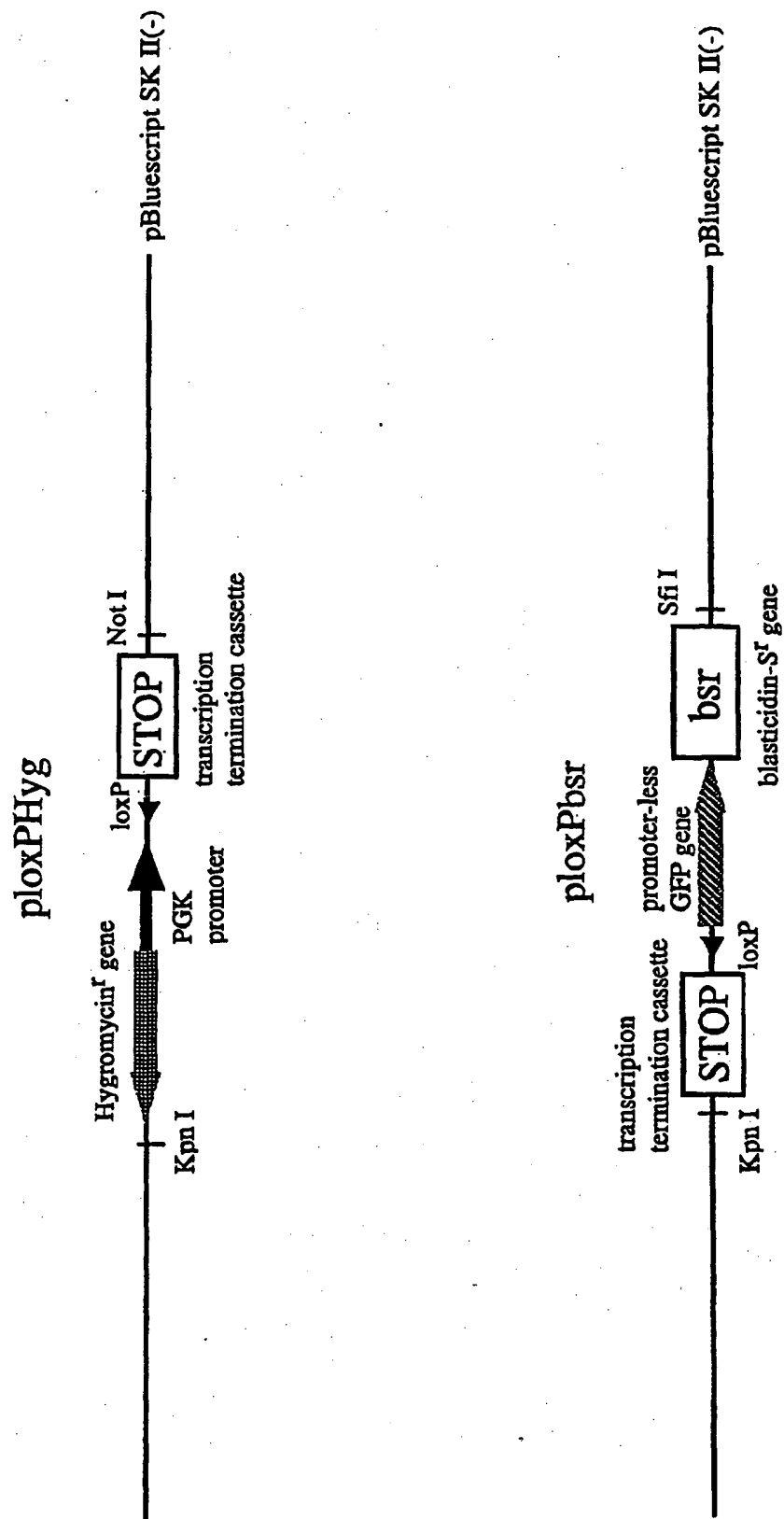


FIG. 60



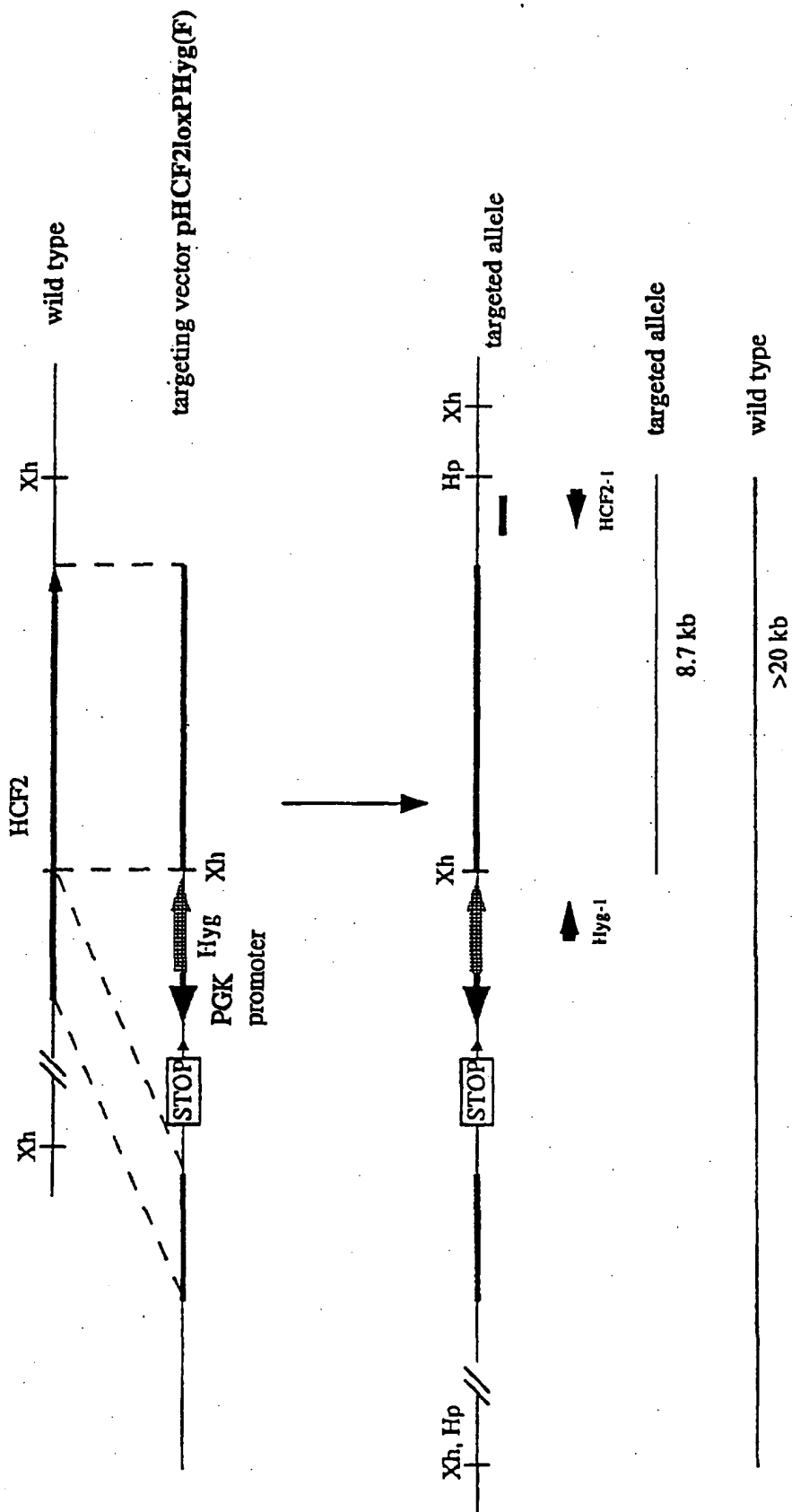


FIG. 61

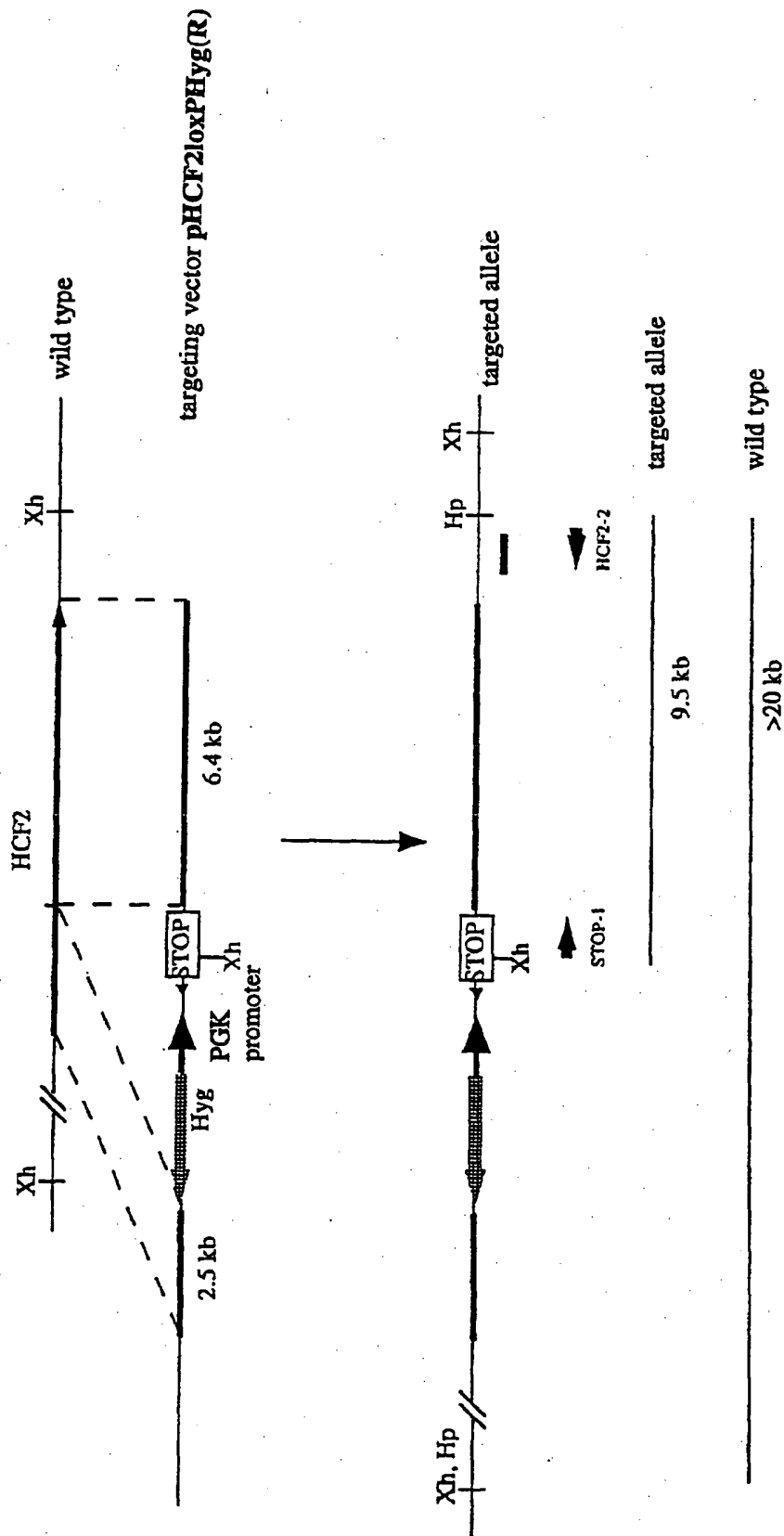


FIG. 62



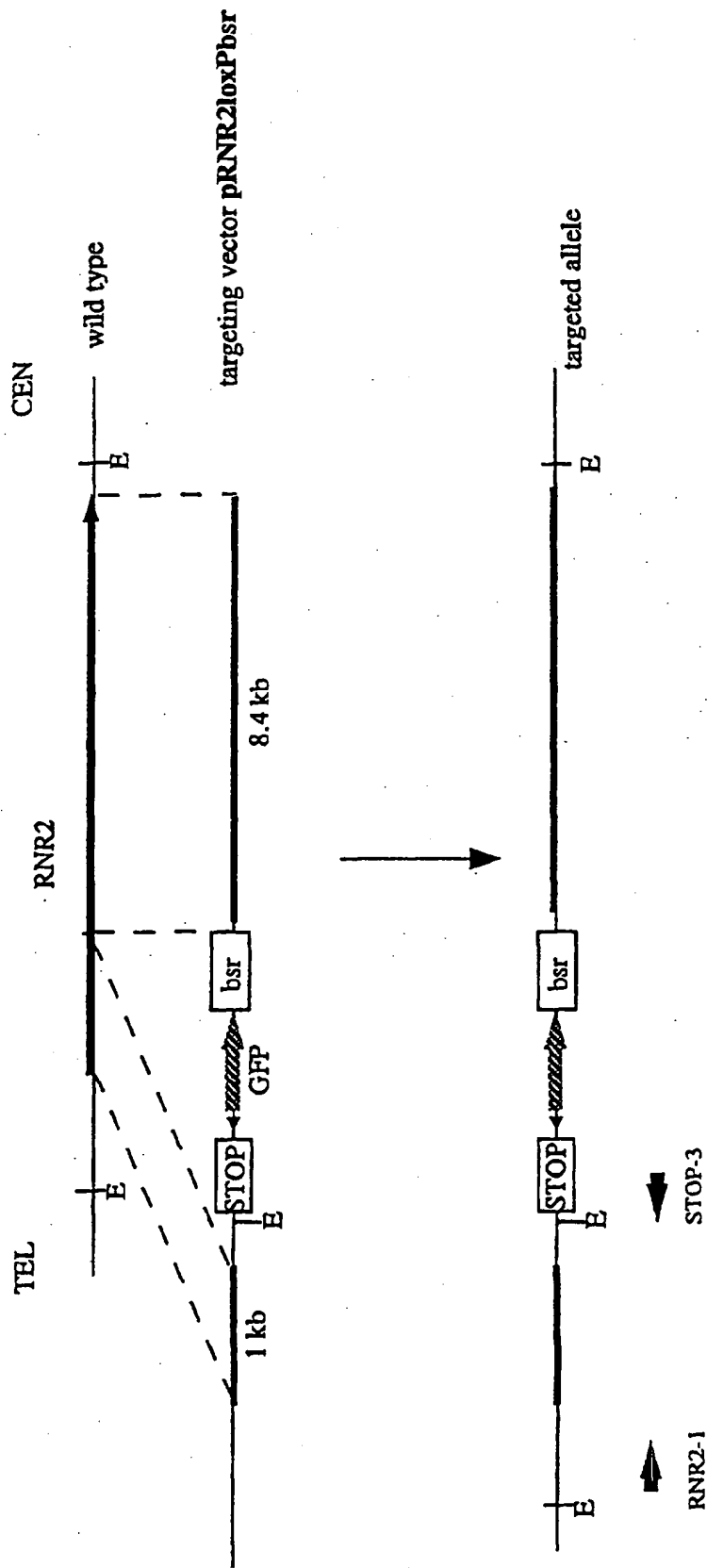


FIG. 63

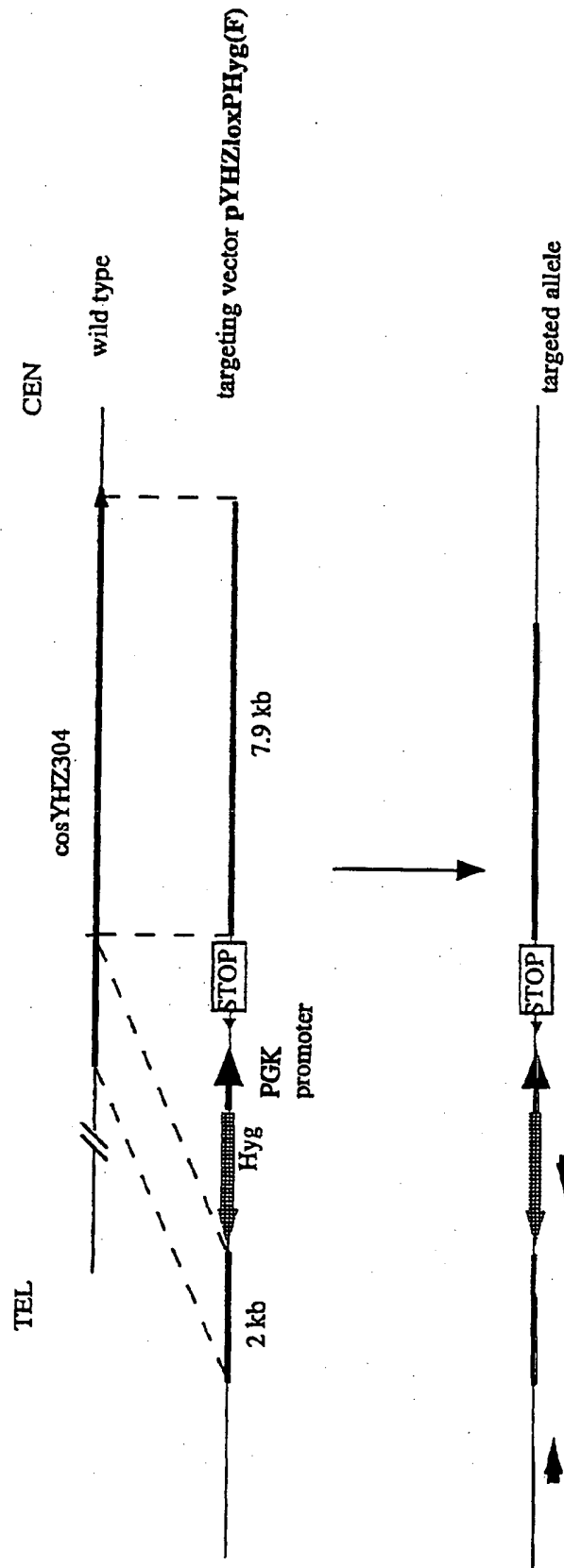
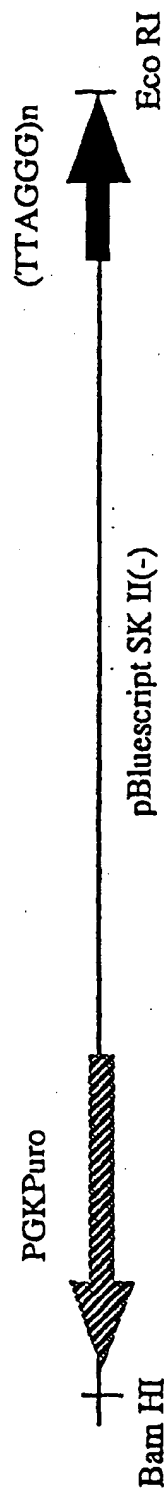


FIG. 64

FIG. 65



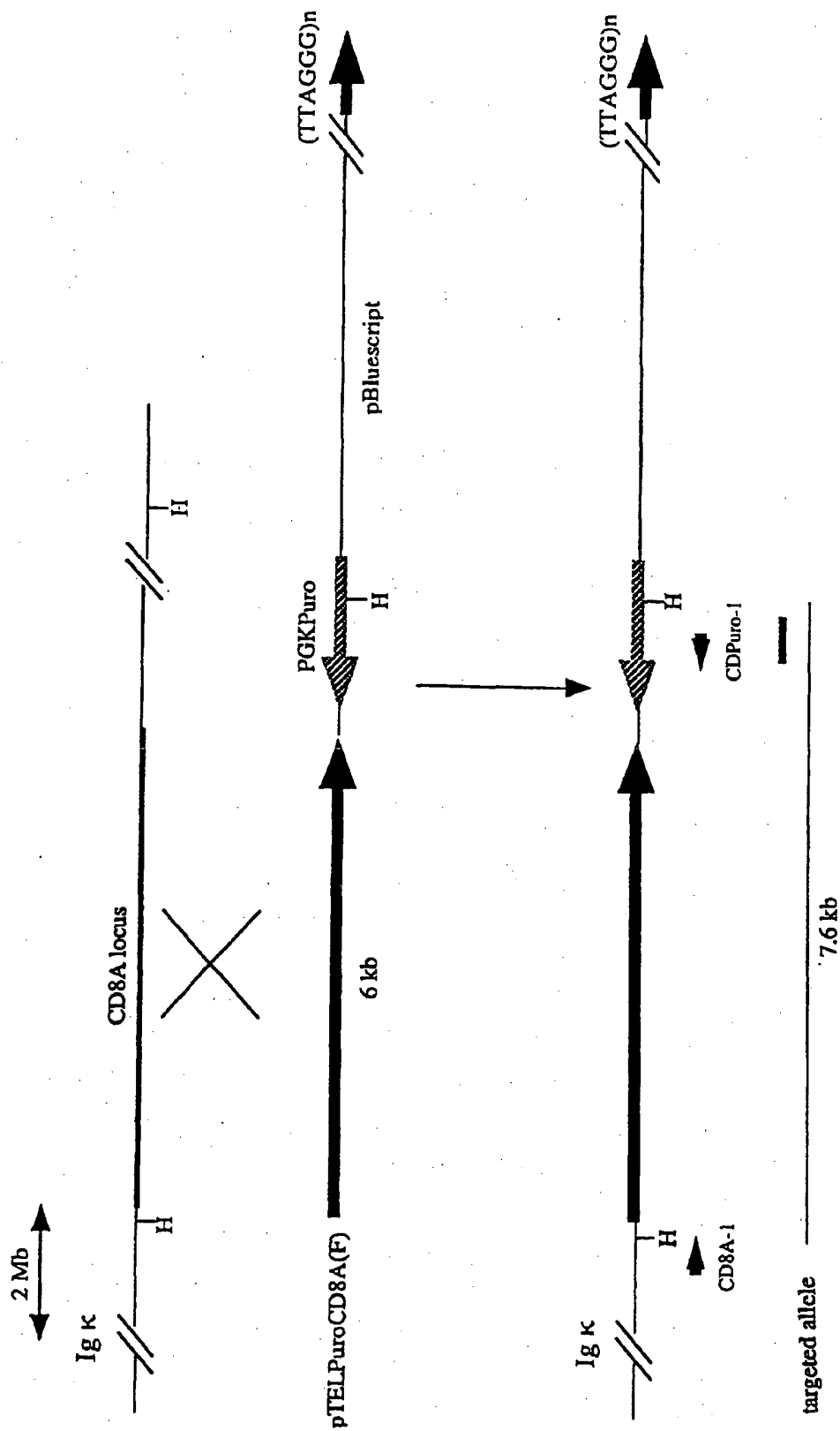


FIG. 66

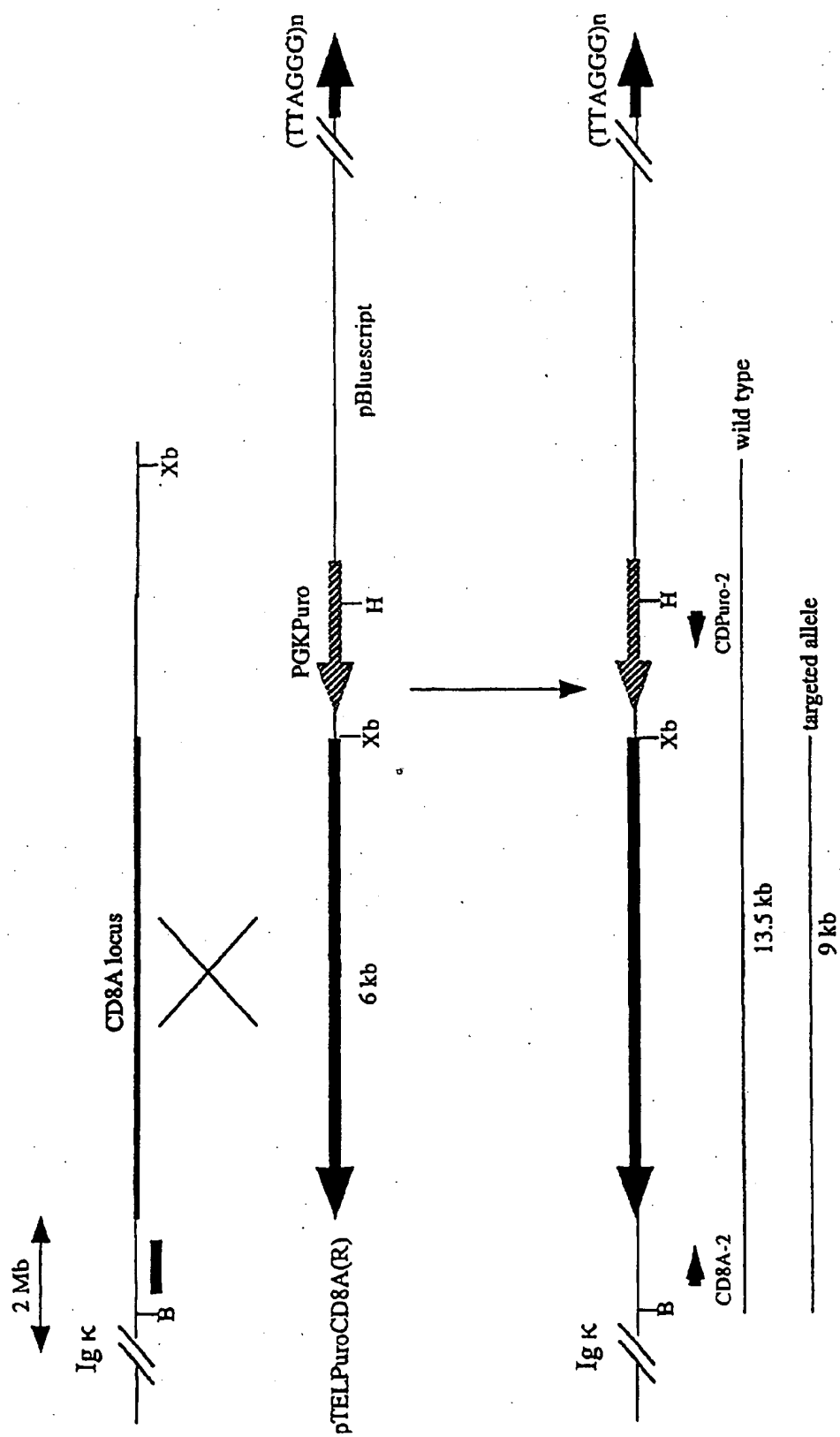
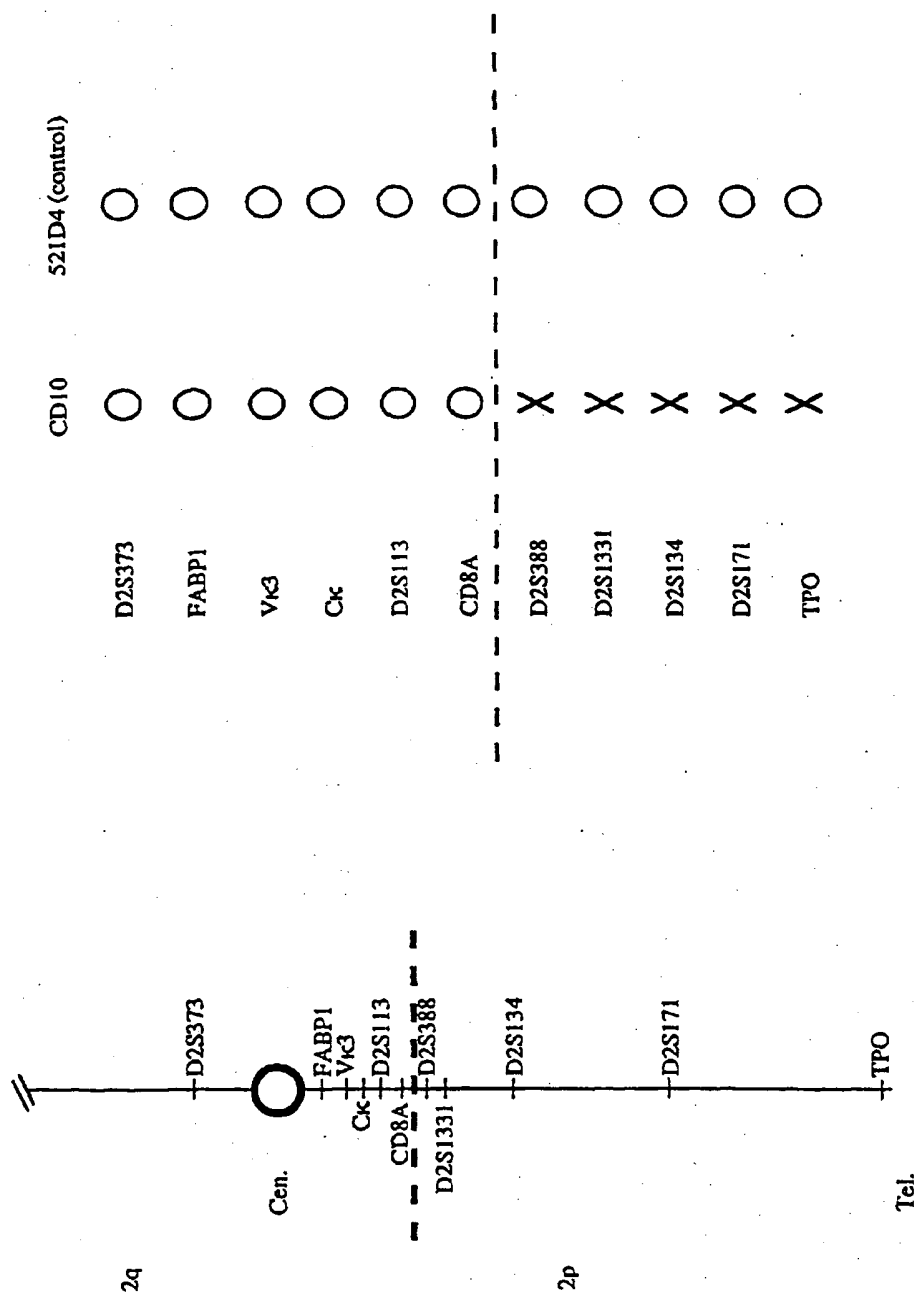


FIG. 67



Human chromosome 2

FIG. 68

FIG. 69

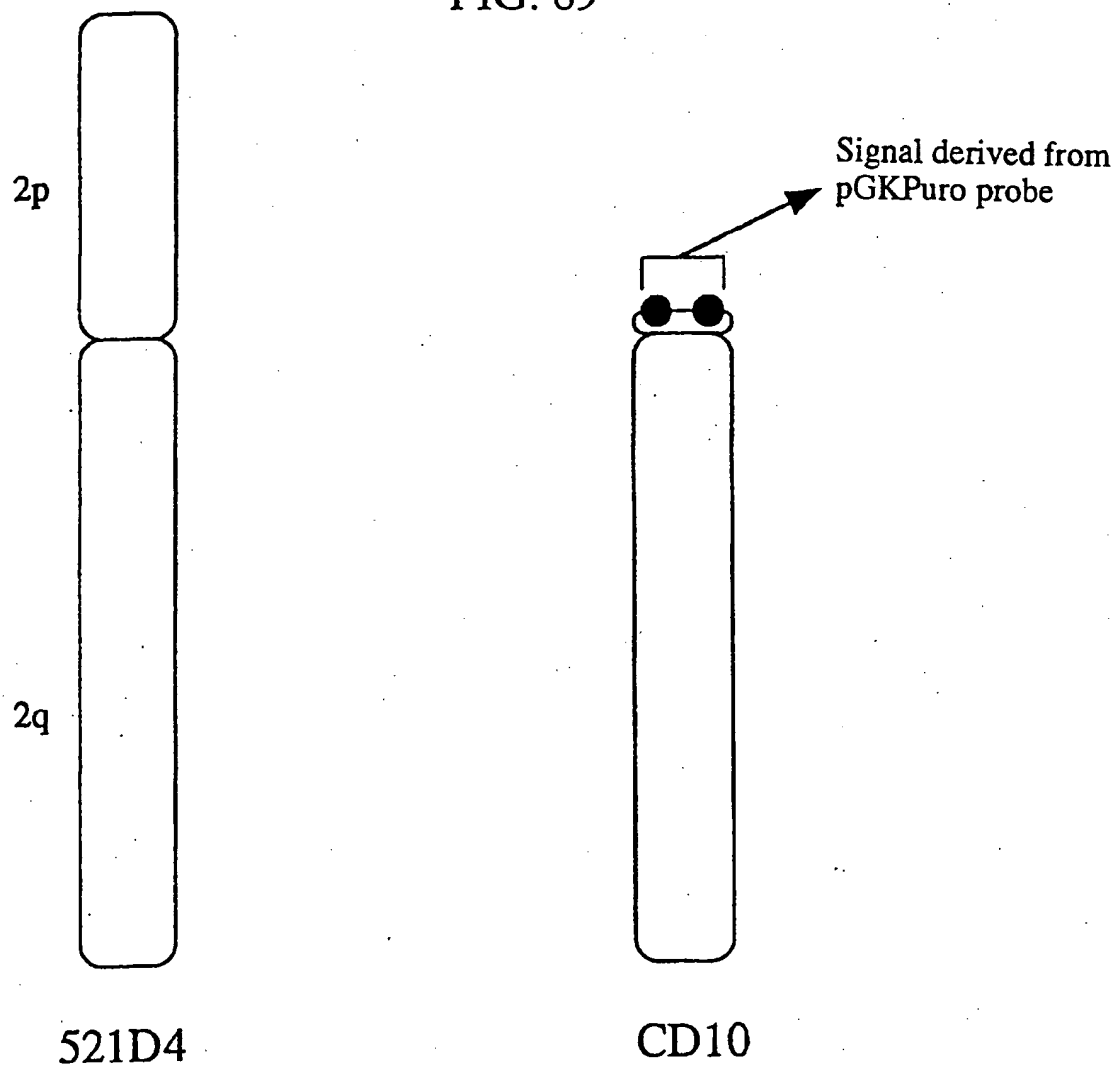


FIG. 70

pBS185hisD

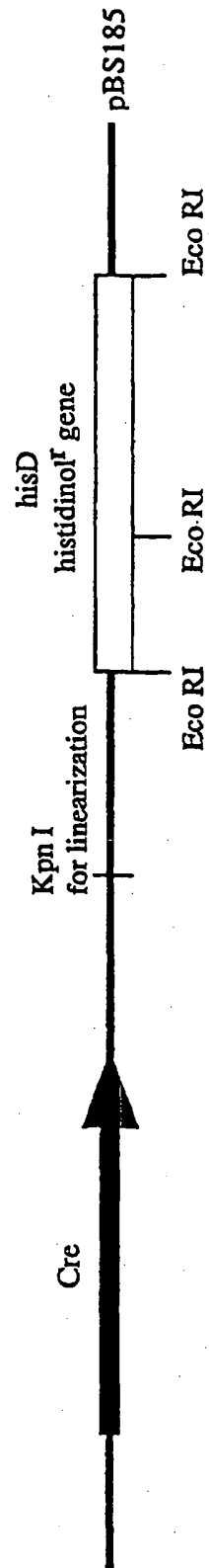
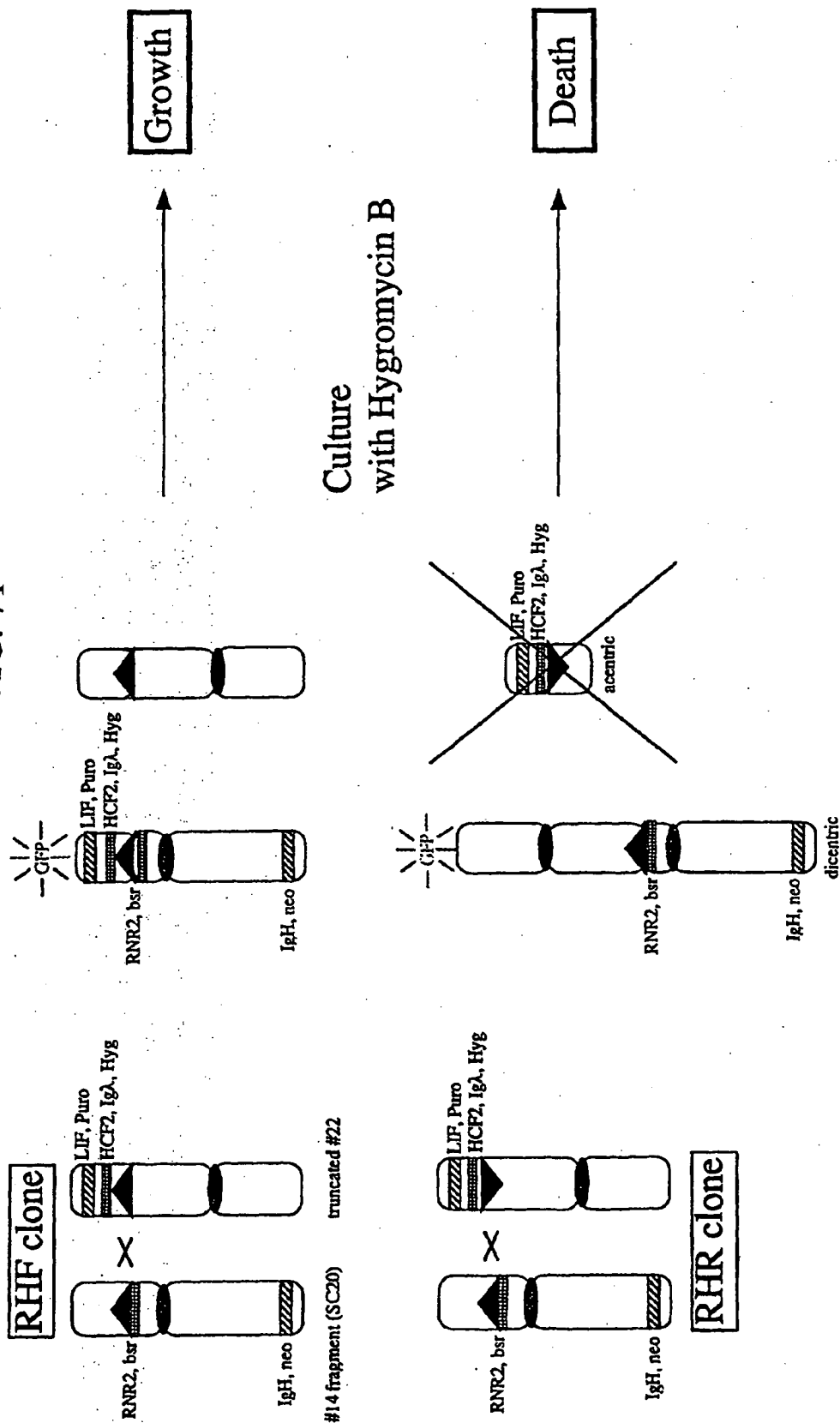




FIG. 71



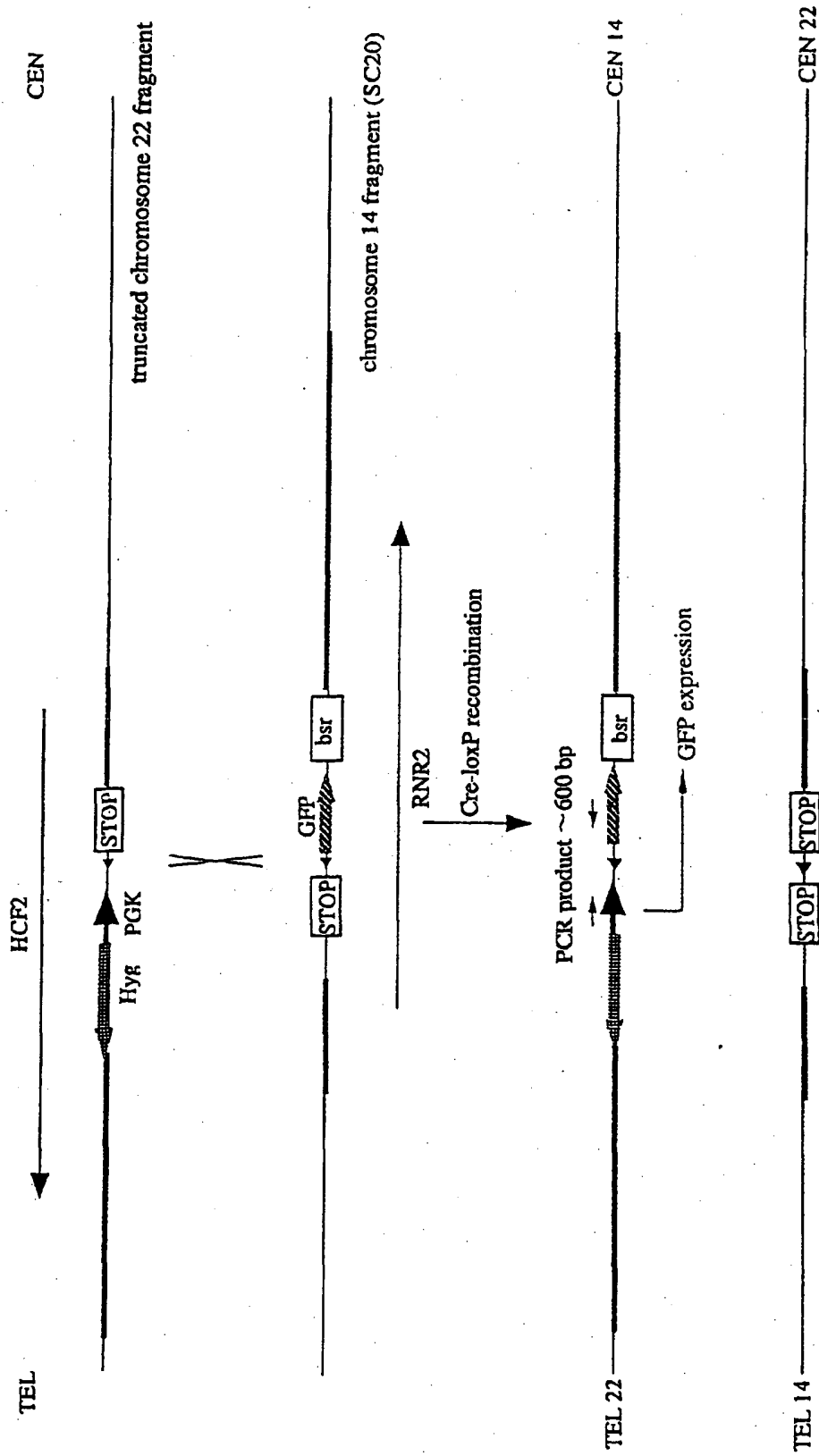


FIG. 72

FIG. 73

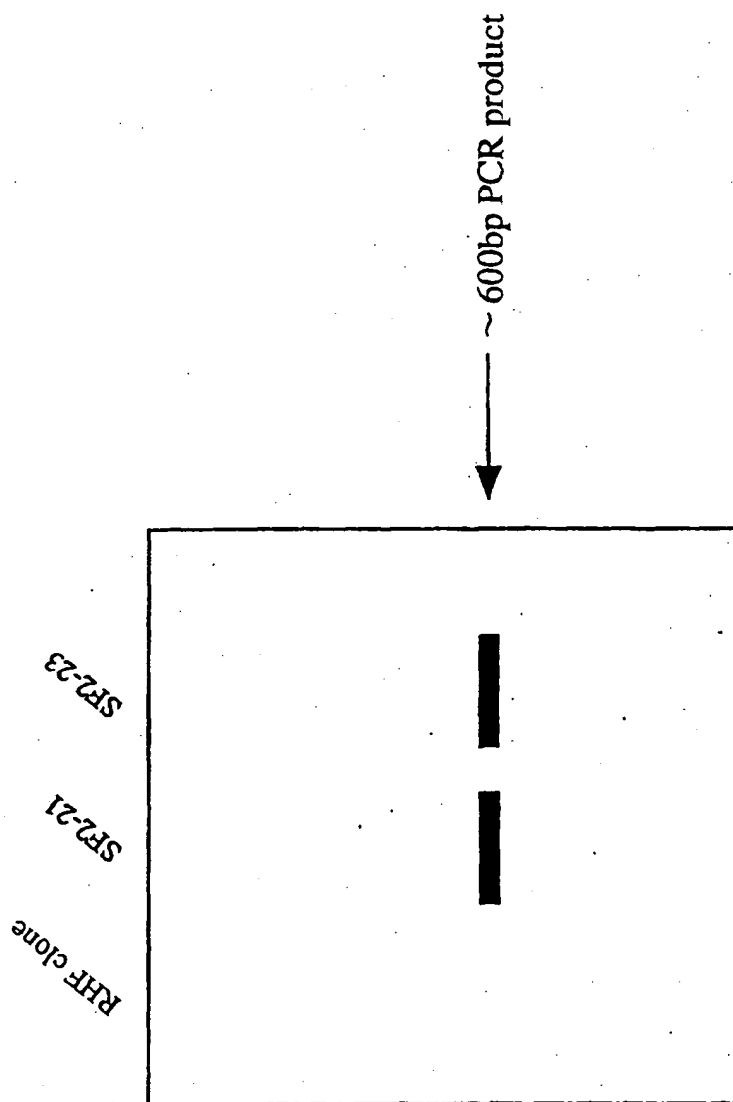


FIG. 74

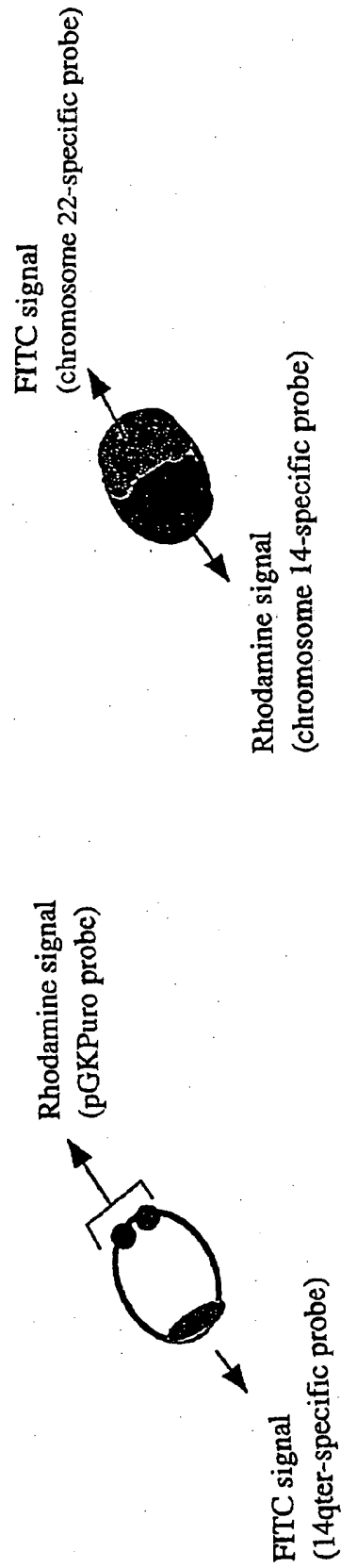
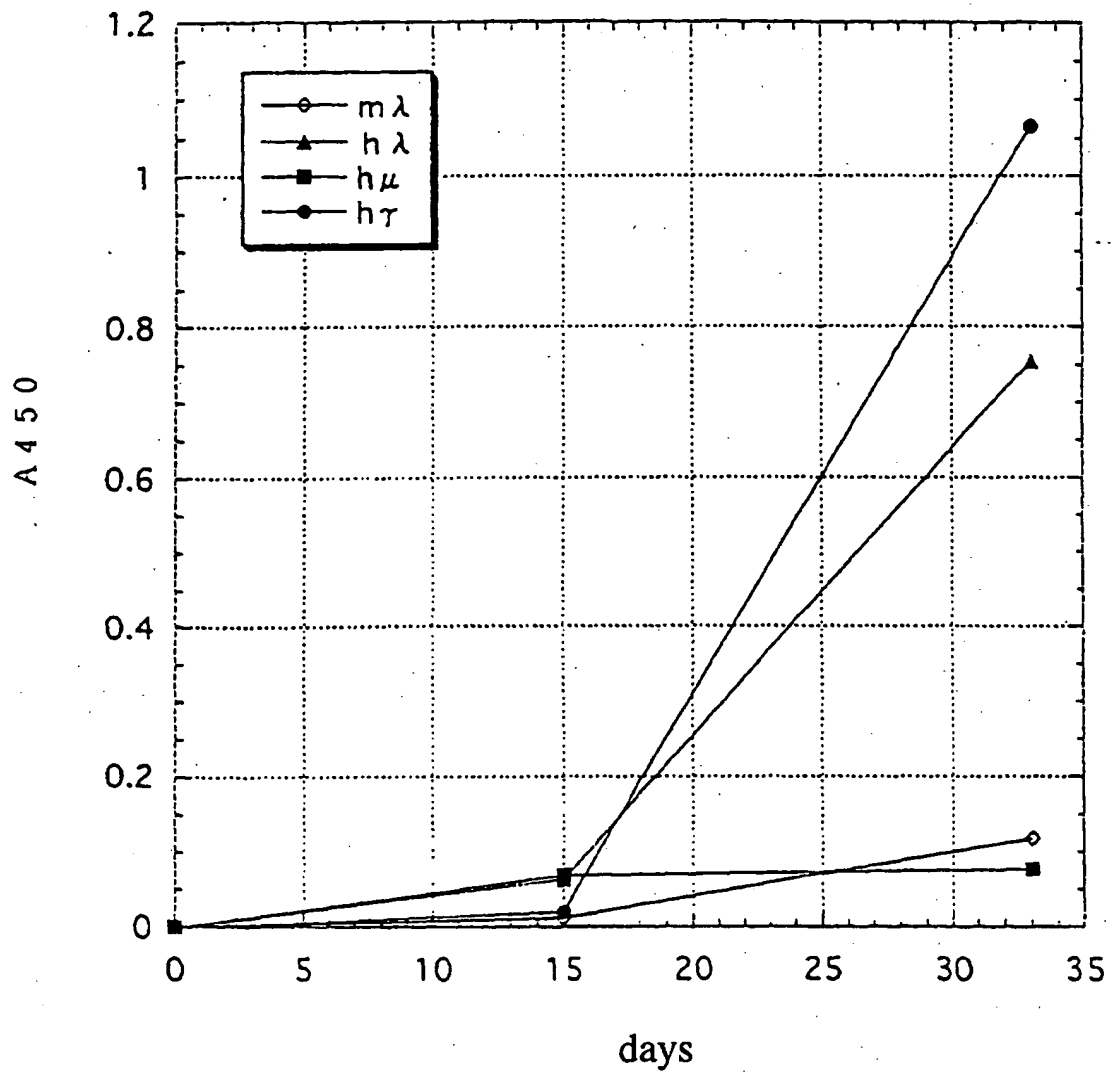


FIG. 75



## INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP99/04518

## A. CLASSIFICATION OF SUBJECT MATTER

Int.Cl.<sup>6</sup> A01K 67/027, C12N 5/10, C12N 15/06

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

Int.Cl.<sup>6</sup> A01K 67/027, C12N 5/10, C12N 15/06

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

BIOSIS (DIALOG), JOIS (JICST)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X Y	Tomizuka K., et al., Nature Genetics, vol.16, p.133-143 (1997)	68,70 26-67,69,71-83
X Y	WO, 97/07671, A1 (Kirin Brewery Company, limited), 06 March, 1997 (06.03.97) & AU, 6837696, A1 & EP, 843961, A1	68,70 26-67,69,71-83
Y	Dieken E.S. et al., Nature Genetics, vol.12, p.174-182 (1996)	1-92
Y	Smith A.J.H. et al., Nature Genetics, vol.8, p.376-385 (1995)	1-92
Y	Willmut et al., Nature, vol.385, p.810 (1997)	46-67
A	Ramirez-Solis R. et al., Nature, vol.378, p.720-724 (1995)	1-92
A	Mills W. et al., Human Mol.Gen., vol.8, p.751-761 (1999)	1-92

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Date of the actual completion of the international search  
15 November, 1999 (15.11.99)Date of mailing of the international search report  
24 November, 1999 (24.11.99)Name and mailing address of the ISA/  
Japanese Patent Office

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## INTERNATIONAL SEARCH REPORT

International application No.

PCT/JP99/04518

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO, 98/54348, A1 (Bruggmann M.), 03 December, 1998 (03.12.98) & GB, 97-11167, A1 & AU, 7667698, A1	1-92
A	WO, 97/49804, A1 (Baylor College of Medicine), 31 December, 1997 (31.12.97) & AU, 3507297, A1 & EP, 907726, A1	1-92

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